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(54) Title: COMPOSITIONS AND METHODS FOR THERAPY AND DIAGNOSIS OF BREAST CANCER

(57) Abstract: Compositions and methods for the therapy and diagnosis of cancer, such as breast cancer, are disclosed. Compositions may comprise one or more breast tumor proteins, immunogenic portions thereof, or polynucleotides that encode such portions. Alternatively, a therapeutic composition may comprise an antigen presenting cell that expresses a breast tumor protein, or a T cell that is specific for cells expressing such a protein. Such compositions may be used, for example, for the prevention and treatment of diseases such as breast cancer. Diagnostic methods based on detecting a breast tumor protein, or mRNA encoding such a protein, in a sample are also provided.

## COMPOSITIONS AND METHODS FOR THERAPY AND DIAGNOSIS OF BREAST CANCER

### TECHNICAL FIELD

The present invention relates generally to therapy and diagnosis of  
5 cancer, such as breast cancer. The invention is more specifically related to polypeptides  
comprising at least a portion of a breast tumor protein, and to polynucleotides encoding  
such polypeptides. Such polypeptides and polynucleotides may be used in vaccines and  
pharmaceutical compositions for prevention and treatment of breast cancer, and for the  
diagnosis and monitoring of such cancers.

### 10 BACKGROUND OF THE INVENTION

Breast cancer is a significant health problem for women in the United  
States and throughout the world. Although advances have been made in detection and  
treatment of the disease, breast cancer remains the second leading cause of cancer-  
related deaths in women, affecting more than 180,000 women in the United States each  
15 year. For women in North America, the life-time odds of getting breast cancer are now  
one in eight.

No vaccine or other universally successful method for the prevention or  
treatment of breast cancer is currently available. Management of the disease currently  
relies on a combination of early diagnosis (through routine breast screening procedures)  
20 and aggressive treatment, which may include one or more of a variety of treatments  
such as surgery, radiotherapy, chemotherapy and hormone therapy. The course of  
treatment for a particular breast cancer is often selected based on a variety of prognostic  
parameters, including an analysis of specific tumor markers. *See, e.g., Porter-Jordan  
and Lippman, Breast Cancer 8:73-100 (1994).* However, the use of established markers  
25 often leads to a result that is difficult to interpret, and the high mortality observed in  
breast cancer patients indicates that improvements are needed in the treatment,  
diagnosis and prevention of the disease.

Accordingly, there is a need in the art for improved methods for therapy and diagnosis of breast cancer. The present invention fulfills these needs and further provides other related advantages.

#### SUMMARY OF THE INVENTION

5                    Briefly stated, the present invention provides compositions and methods for the diagnosis and therapy of cancer, such as breast cancer. In one aspect, the present invention provides polypeptides comprising at least a portion of a breast tumor protein, or a variant thereof. Certain portions and other variants are immunogenic, such that the ability of the variant to react with antigen-specific antisera is not substantially  
10 diminished. Within certain embodiments, the polypeptide comprises a sequence that is encoded by a polynucleotide sequence selected from the group consisting of: (a) sequences recited in SEQ ID NO: 1-38, 42-204, 205, 207 and 210-290; (b) variants of a sequence recited in SEQ ID NO: 1-38, 42-204, 205, 207 and 210-290; and (c) complements of a sequence of (a) or (b).

15                    The present invention further provides polynucleotides that encode a polypeptide as described above, or a portion thereof (such as a portion encoding at least 15 amino acid residues of a breast tumor protein), expression vectors comprising such polynucleotides and host cells transformed or transfected with such expression vectors.

                    Within other aspects, the present invention provides pharmaceutical  
20 compositions comprising a polypeptide or polynucleotide as described above and a physiologically acceptable carrier.

                    Within a related aspect of the present invention, vaccines for prophylactic or therapeutic use are provided. Such vaccines comprise a polypeptide or polynucleotide as described above and an immunostimulant.

25                    The present invention further provides pharmaceutical compositions that comprise: (a) an antibody or antigen-binding fragment thereof that specifically binds to a breast tumor protein; and (b) a physiologically acceptable carrier.

                    Within further aspects, the present invention provides pharmaceutical compositions comprising: (a) an antigen presenting cell that expresses a polypeptide as  
30 described above and (b) a pharmaceutically acceptable carrier or excipient. Antigen

presenting cells include dendritic cells, macrophages, monocytes, fibroblasts and B cells.

Within related aspects, vaccines are provided that comprise: (a) an antigen presenting cell that expresses a polypeptide as described above and (b) an immunostimulant.

The present invention further provides, in other aspects, fusion proteins that comprise at least one polypeptide as described above, as well as polynucleotides encoding such fusion proteins.

Within related aspects, pharmaceutical compositions comprising a fusion protein, or a polynucleotide encoding a fusion protein, in combination with a physiologically acceptable carrier are provided.

Vaccines are further provided, within other aspects, that comprise a fusion protein, or a polynucleotide encoding a fusion protein, in combination with an immunostimulant.

Within further aspects, the present invention provides methods for inhibiting the development of a cancer in a patient, comprising administering to a patient a pharmaceutical composition or vaccine as recited above.

The present invention further provides, within other aspects, methods for removing tumor cells from a biological sample, comprising contacting a biological sample with T cells that specifically react with a breast tumor protein, wherein the step of contacting is performed under conditions and for a time sufficient to permit the removal of cells expressing the protein from the sample.

Within related aspects, methods are provided for inhibiting the development of a cancer in a patient, comprising administering to a patient a biological sample treated as described above.

Methods are further provided, within other aspects, for stimulating and/or expanding T cells specific for a breast tumor protein, comprising contacting T cells with one or more of: (i) a polypeptide as described above; (ii) a polynucleotide encoding such a polypeptide; and/or (iii) an antigen presenting cell that expresses such a polypeptide; under conditions and for a time sufficient to permit the stimulation and/or



expansion of T cells. Isolated T cell populations comprising T cells prepared as described above are also provided.

Within further aspects, the present invention provides methods for inhibiting the development of a cancer in a patient, comprising administering to a  
5 patient an effective amount of a T cell population as described above.

The present invention further provides methods for inhibiting the development of a cancer in a patient, comprising the steps of: (a) incubating CD4<sup>+</sup> and/or CD8<sup>+</sup> T cells isolated from a patient with one or more of: (i) a polypeptide comprising at least an immunogenic portion of a breast tumor protein; (ii) a  
10 polynucleotide encoding such a polypeptide; and (iii) an antigen-presenting cell that expresses such a polypeptide; and (b) administering to the patient an effective amount of the proliferated T cells, and thereby inhibiting the development of a cancer in the patient. Proliferated cells may, but need not, be cloned prior to administration to the patient.

15 Within further aspects, the present invention provides methods for determining the presence or absence of a cancer in a patient, comprising: (a) contacting a biological sample obtained from a patient with a binding agent that binds to a polypeptide as recited above; (b) detecting in the sample an amount of polypeptide that binds to the binding agent; and (c) comparing the amount of polypeptide with a  
20 predetermined cut-off value, and therefrom determining the presence or absence of a cancer in the patient. Within preferred embodiments, the binding agent is an antibody, more preferably a monoclonal antibody. The cancer may be breast cancer.

The present invention also provides, within other aspects, methods for monitoring the progression of a cancer in a patient. Such methods comprise the steps  
25 of: (a) contacting a biological sample obtained from a patient at a first point in time with a binding agent that binds to a polypeptide as recited above; (b) detecting in the sample an amount of polypeptide that binds to the binding agent; (c) repeating steps (a) and (b) using a biological sample obtained from the patient at a subsequent point in time; and (d) comparing the amount of polypeptide detected in step (c) with the amount  
30 detected in step (b) and therefrom monitoring the progression of the cancer in the patient.

The present invention further provides, within other aspects, methods for determining the presence or absence of a cancer in a patient, comprising the steps of: (a) contacting a biological sample obtained from a patient with an oligonucleotide that hybridizes to a polynucleotide that encodes a breast tumor protein; (b) detecting in the sample a level of a polynucleotide, preferably mRNA, that hybridizes to the oligonucleotide; and (c) comparing the level of polynucleotide that hybridizes to the oligonucleotide with a predetermined cut-off value, and therefrom determining the presence or absence of a cancer in the patient. Within certain embodiments, the amount of mRNA is detected via polymerase chain reaction using, for example, at least one oligonucleotide primer that hybridizes to a polynucleotide encoding a polypeptide as recited above, or a complement of such a polynucleotide. Within other embodiments, the amount of mRNA is detected using a hybridization technique, employing an oligonucleotide probe that hybridizes to a polynucleotide that encodes a polypeptide as recited above, or a complement of such a polynucleotide.

In related aspects, methods are provided for monitoring the progression of a cancer in a patient, comprising the steps of: (a) contacting a biological sample obtained from a patient with an oligonucleotide that hybridizes to a polynucleotide that encodes a breast tumor protein; (b) detecting in the sample an amount of a polynucleotide that hybridizes to the oligonucleotide; (c) repeating steps (a) and (b) using a biological sample obtained from the patient at a subsequent point in time; and (d) comparing the amount of polynucleotide detected in step (c) with the amount detected in step (b) and therefrom monitoring the progression of the cancer in the patient.

Within further aspects, the present invention provides antibodies, such as monoclonal antibodies, that bind to a polypeptide as described above, as well as diagnostic kits comprising such antibodies. Diagnostic kits comprising one or more oligonucleotide probes or primers as described above are also provided.

These and other aspects of the present invention will become apparent upon reference to the following detailed description. All references disclosed herein are hereby incorporated by reference in their entirety as if each was incorporated individually.

**SEQUENCE IDENTIFIERS**

- SEQ ID NO: 1 is the determined cDNA sequence for clone 26915.
- SEQ ID NO: 2 is the determined cDNA sequence for clone 26914.
- SEQ ID NO: 3 is the determined cDNA sequence for clone 26673.
- 5 SEQ ID NO: 4 is the determined cDNA sequence for clone 26672.
- SEQ ID NO: 5 is the determined cDNA sequence for clone 26671.
- SEQ ID NO: 6 is the determined cDNA sequence for clone 26670.
- SEQ ID NO: 7 is the determined cDNA sequence for clone 26669.
- SEQ ID NO: 8 is a first determined cDNA sequence for clone 26668.
- 10 SEQ ID NO: 9 is a second determined cDNA sequence for clone 26668.
- SEQ ID NO: 10 is the determined cDNA sequence for clone 26667.
- SEQ ID NO: 11 is the determined cDNA sequence for clone 26666.
- SEQ ID NO: 12 is the determined cDNA sequence for clone 26665.
- SEQ ID NO: 13 is the determined cDNA sequence for clone 26664.
- 15 SEQ ID NO: 14 is the determined cDNA sequence for clone 26662.
- SEQ ID NO: 15 is the determined cDNA sequence for clone 26661.
- SEQ ID NO: 16 is the determined cDNA sequence for clone 26660.
- SEQ ID NO: 17 is the determined cDNA sequence for clone 26603.
- SEQ ID NO: 18 is the determined cDNA sequence for clone 26601.
- 20 SEQ ID NO: 19 is the determined cDNA sequence for clone 26600.
- SEQ ID NO: 20 is the determined cDNA sequence for clone 26587.
- SEQ ID NO: 21 is the determined cDNA sequence for clone 26586.
- SEQ ID NO: 22 is the determined cDNA sequence for clone 26584.
- SEQ ID NO: 23 is the determined cDNA sequence for clone 26583.
- 25 SEQ ID NO: 24 is the determined cDNA sequence for clone 26580.
- SEQ ID NO: 25 is the determined cDNA sequence for clone 26579.
- SEQ ID NO: 26 is the determined cDNA sequence for clone 26577.
- SEQ ID NO: 27 is the determined cDNA sequence for clone 26575.
- SEQ ID NO: 28 is the determined cDNA sequence for clone 26574.
- 30 SEQ ID NO: 29 is the determined cDNA sequence for clone 26573.
- SEQ ID NO: 30 is the determined cDNA sequence for clone 25612.

- SEQ ID NO: 31 is the determined cDNA sequence for clone 22295.  
SEQ ID NO: 32 is the determined cDNA sequence for clone 22301.  
SEQ ID NO: 33 is the determined cDNA sequence for clone 22298.  
SEQ ID NO: 34 is the determined cDNA sequence for clone 22297.  
5 SEQ ID NO: 35 is the determined cDNA sequence for clone 22303.  
SEQ ID NO: 36 is the determined cDNA sequence for a first GABA<sub>A</sub> receptor clone.  
SEQ ID NO: 37 is the determined cDNA sequence for a second GABA<sub>A</sub> receptor clone.  
10 SEQ ID NO: 38 is the determined cDNA sequence for a third GABA<sub>A</sub> receptor clone.  
SEQ ID NO: 39 is the amino acid sequence encoded by SEQ ID NO: 36.  
SEQ ID NO: 40 is the amino acid sequence encoded by SEQ ID NO: 37.  
SEQ ID NO: 41 is the amino acid sequence encoded by SEQ ID NO: 38.  
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SEQ ID NO: 43 is the determined cDNA sequence for contig 2.  
SEQ ID NO: 44 is the determined cDNA sequence for contig 3.  
SEQ ID NO: 45 is the determined cDNA sequence for contig 4.  
SEQ ID NO: 46 is the determined cDNA sequence for contig 5.  
20 SEQ ID NO: 47 is the determined cDNA sequence for contig 6.  
SEQ ID NO: 48 is the determined cDNA sequence for contig 7.  
SEQ ID NO: 49 is the determined cDNA sequence for contig 8.  
SEQ ID NO: 50 is the determined cDNA sequence for contig 9.  
SEQ ID NO: 51 is the determined cDNA sequence for contig 10.  
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SEQ ID NO: 53 is the determined cDNA sequence for contig 12.  
SEQ ID NO: 54 is the determined cDNA sequence for contig 13.  
SEQ ID NO: 55 is the determined cDNA sequence for contig 14.  
SEQ ID NO: 56 is the determined cDNA sequence for contig 15.  
30 SEQ ID NO: 57 is the determined cDNA sequence for contig 16.  
SEQ ID NO: 58 is the determined cDNA sequence for contig 17.

SEQ ID NO: 59 is the determined cDNA sequence for contig 18.  
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SEQ ID NO: 64 is the determined cDNA sequence for contig 23.  
SEQ ID NO: 65 is the determined cDNA sequence for contig 24.  
SEQ ID NO: 66 is the determined cDNA sequence for contig 25.  
SEQ ID NO: 67 is the determined cDNA sequence for contig 26.  
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SEQ ID NO: 70 is the determined cDNA sequence for contig 29.  
SEQ ID NO: 71 is the determined cDNA sequence for contig 30.  
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SEQ ID NO: 74 is the determined cDNA sequence for contig 33.  
SEQ ID NO: 75 is the determined cDNA sequence for contig 34.  
SEQ ID NO: 76 is the determined cDNA sequence for contig 35.  
SEQ ID NO: 77 is the determined cDNA sequence for contig 36.  
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SEQ ID NO: 81 is the determined cDNA sequence for contig 40.  
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25 SEQ ID NO: 83 is the determined cDNA sequence for contig 42.  
SEQ ID NO: 84 is the determined cDNA sequence for contig 43.  
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SEQ ID NO: 85 is the determined cDNA sequence for contig 45.  
SEQ ID NO: 85 is the determined cDNA sequence for contig 46.  
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SEQ ID NO: 89 is the determined cDNA sequence for contig 48.

SEQ ID NO: 90 is the determined cDNA sequence for contig 49.  
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SEQ ID NO: 93 is the determined cDNA sequence for contig 52.  
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SEQ ID NO: 95 is the determined cDNA sequence for contig 54.  
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SEQ ID NO: 97 is the determined cDNA sequence for contig 56.  
SEQ ID NO: 98 is the determined cDNA sequence for contig 57.  
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SEQ ID NO: 100 is the determined cDNA sequence for contig 59.  
SEQ ID NO: 101 is the determined cDNA sequence for contig 60.  
SEQ ID NO: 102 is the determined cDNA sequence for contig 61.  
SEQ ID NO: 103 is the determined cDNA sequence for contig 62.  
15 SEQ ID NO: 104 is the determined cDNA sequence for contig 63.  
SEQ ID NO: 105 is the determined cDNA sequence for contig 64.  
SEQ ID NO: 106 is the determined cDNA sequence for contig 65.  
SEQ ID NO: 107 is the determined cDNA sequence for contig 66.  
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SEQ ID NO: 110 is the determined cDNA sequence for contig 69.  
SEQ ID NO: 111 is the determined cDNA sequence for contig 70.  
SEQ ID NO: 112 is the determined cDNA sequence for contig 71.  
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25 SEQ ID NO: 114 is the determined cDNA sequence for contig 73.  
SEQ ID NO: 115 is the determined cDNA sequence for contig 74.  
SEQ ID NO: 116 is the determined cDNA sequence for contig 75.  
SEQ ID NO: 117 is the determined cDNA sequence for contig 76.  
SEQ ID NO: 118 is the determined cDNA sequence for contig 77.  
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SEQ ID NO: 120 is the determined cDNA sequence for contig 79.

SEQ ID NO: 121 is the determined cDNA sequence for contig 80.  
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SEQ ID NO: 123 is the determined cDNA sequence for contig 82.  
SEQ ID NO: 124 is the determined cDNA sequence for contig 83.  
5 SEQ ID NO: 125 is the determined cDNA sequence for contig 84.  
SEQ ID NO: 126 is the determined cDNA sequence for contig 85.  
SEQ ID NO: 127 is the determined cDNA sequence for contig 86.  
SEQ ID NO: 128 is the determined cDNA sequence for contig 87.  
SEQ ID NO: 129 is the determined cDNA sequence for contig 88.  
10 SEQ ID NO: 130 is the determined cDNA sequence for contig 89.  
SEQ ID NO: 131 is the determined cDNA sequence for contig 90.  
SEQ ID NO: 132 is the determined cDNA sequence for contig 91.  
SEQ ID NO: 133 is the determined cDNA sequence for contig 92.  
SEQ ID NO: 134 is the determined cDNA sequence for contig 93.  
15 SEQ ID NO: 135 is the determined cDNA sequence for contig 94.  
SEQ ID NO: 136 is the determined cDNA sequence for contig 95.  
SEQ ID NO: 137 is the determined cDNA sequence for contig 96.  
SEQ ID NO: 138 is the determined cDNA sequence for clone 47589.  
SEQ ID NO: 139 is the determined cDNA sequence for clone 47578.  
20 SEQ ID NO: 140 is the determined cDNA sequence for clone 47602.  
SEQ ID NO: 141 is the determined cDNA sequence for clone 47593.  
SEQ ID NO: 142 is the determined cDNA sequence for clone 47583.  
SEQ ID NO: 143 is the determined cDNA sequence for clone 47624.  
SEQ ID NO: 144 is the determined cDNA sequence for clone 47622.  
25 SEQ ID NO: 145 is the determined cDNA sequence for clone 47649.  
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SEQ ID NO: 147 is the determined cDNA sequence for clone 48962.  
SEQ ID NO: 148 is the determined cDNA sequence for clone 48964.  
SEQ ID NO: 149 is the determined cDNA sequence for clone 48987.  
30 SEQ ID NO: 150 is the determined cDNA sequence for clone 49002.  
SEQ ID NO: 151 is the determined cDNA sequence for clone 48950.

SEQ ID NO: 152 is the determined cDNA sequence for clone 48934.  
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SEQ ID NO: 162 is the determined cDNA sequence for clone 48965.  
SEQ ID NO: 163 is the determined cDNA sequence for clone 48970.  
SEQ ID NO: 164 is the determined cDNA sequence for clone 48984.  
SEQ ID NO: 165 is the determined cDNA sequence for clone 48969.  
15 SEQ ID NO: 166 is the determined cDNA sequence for clone 48978.  
SEQ ID NO: 167 is the determined cDNA sequence for clone 48968.  
SEQ ID NO: 168 is the determined cDNA sequence for clone 48929.  
SEQ ID NO: 169 is the determined cDNA sequence for clone 48937.  
SEQ ID NO: 170 is the determined cDNA sequence for clone 48982.  
20 SEQ ID NO: 171 is the determined cDNA sequence for clone 48983.  
SEQ ID NO: 172 is the determined cDNA sequence for clone 48997.  
SEQ ID NO: 173 is the determined cDNA sequence for clone 48992.  
SEQ ID NO: 174 is the determined cDNA sequence for clone 49006.  
SEQ ID NO: 175 is the determined cDNA sequence for clone 48994.  
25 SEQ ID NO: 176 is the determined cDNA sequence for clone 49013.  
SEQ ID NO: 177 is the determined cDNA sequence for clone 49008.  
SEQ ID NO: 178 is the determined cDNA sequence for clone 48990.  
SEQ ID NO: 179 is the determined cDNA sequence for clone 48989.  
SEQ ID NO: 180 is the determined cDNA sequence for clone 49014.  
30 SEQ ID NO: 181 is the determined cDNA sequence for clone 48988.  
SEQ ID NO: 182 is the determined cDNA sequence for clone 49018.



SEQ ID NO: 183 is the determined cDNA sequence for clone 6921.  
SEQ ID NO: 184 is the determined cDNA sequence for clone 6837.  
SEQ ID NO: 185 is the determined cDNA sequence for clone 6840.  
SEQ ID NO: 186 is the determined cDNA sequence for clone 6844.  
5 SEQ ID NO: 187 is the determined cDNA sequence for clone 6854.  
SEQ ID NO: 188 is the determined cDNA sequence for clone 6872.  
SEQ ID NO: 189 is the determined cDNA sequence for clone 6906.  
SEQ ID NO: 190 is the determined cDNA sequence for clone 6908.  
SEQ ID NO: 191 is the determined cDNA sequence for clone 6910.  
10 SEQ ID NO: 192 is the determined cDNA sequence for clone 6912.  
SEQ ID NO: 193 is the determined cDNA sequence for clone 6913.  
SEQ ID NO: 194 is the determined cDNA sequence for clone 6914.  
SEQ ID NO: 195 is the determined cDNA sequence for clone 6916.  
SEQ ID NO: 196 is the determined cDNA sequence for clone 6918.  
15 SEQ ID NO: 197 is the determined cDNA sequence for clone 6924.  
SEQ ID NO: 198 is the determined cDNA sequence for clone 6928.  
SEQ ID NO: 199 is the determined cDNA sequence for clone 6978A.  
SEQ ID NO: 200 is the determined cDNA sequence for clone 6978B.  
SEQ ID NO: 201 is the determined cDNA sequence for clone 6982A.  
20 SEQ ID NO: 202 is the determined cDNA sequence for clone 6982B.  
SEQ ID NO: 203 is the determined cDNA sequence for clone 6850.  
SEQ ID NO: 204 is the determined cDNA sequence for clone 6860.  
SEQ ID NO: 205 is the determined cDNA sequence for O772P.  
SEQ ID NO: 206 is the amino acid sequence encoded by SEQ ID NO:  
25 205.  
  
SEQ ID NO: 207 is the full-length cDNA sequence for O8E.  
SEQ ID NO: 208 is a first amino acid sequence encoded by SEQ ID NO:  
207.  
  
SEQ ID NO: 209 is a second amino acid sequence encoded by SEQ ID  
30 NO: 209.

SEQ ID NO: 210-290 are determined cDNA sequence of breast-tumor specific clones.

#### DETAILED DESCRIPTION OF THE INVENTION

As noted above, the present invention is generally directed to compositions and methods for using the compositions, for example in the therapy and diagnosis of cancer, such as breast cancer. Certain illustrative compositions described herein include breast tumor polypeptides, polynucleotides encoding such polypeptides, binding agents such as antibodies, antigen presenting cells (APCs) and/or immune system cells (*e.g.*, T cells). A "breast tumor protein," as the term is used herein, refers generally to a protein that is expressed in breast tumor cells at a level that is at least two fold, and preferably at least five fold, greater than the level of expression in other normal tissues, as determined using a representative assay provided herein. Certain breast tumor proteins are tumor proteins that react detectably (within an immunoassay, such as an ELISA or Western blot) with antisera of a patient afflicted with breast cancer.

Therefore, in accordance with the above, and as described further below, the present invention provides illustrative polynucleotide compositions having sequences set forth in SEQ ID NO:1-38, 42-204, 205, 207 and 210-290, polypeptides encoded by such polynucleotides, antibody compositions capable of binding such polypeptides, and numerous additional embodiments employing such compositions, for example in the detection, diagnosis and/or therapy of human breast cancer.

#### POLYNUCLEOTIDE COMPOSITIONS

As used herein, the terms "DNA segment" and "polynucleotide" refer to a DNA molecule that has been isolated free of total genomic DNA of a particular species. Therefore, a DNA segment encoding a polypeptide refers to a DNA segment that contains one or more coding sequences yet is substantially isolated away from, or purified free from, total genomic DNA of the species from which the DNA segment is obtained. Included within the terms "DNA segment" and "polynucleotide" are DNA

segments and smaller fragments of such segments, and also recombinant vectors, including, for example, plasmids, cosmids, phagemids, phage, viruses, and the like.

As will be understood by those skilled in the art, the DNA segments of this invention can include genomic sequences, extra-genomic and plasmid-encoded  
5 sequences and smaller engineered gene segments that express, or may be adapted to express, proteins, polypeptides, peptides and the like. Such segments may be naturally isolated, or modified synthetically by the hand of man.

"Isolated," as used herein, means that a polynucleotide is substantially away from other coding sequences, and that the DNA segment does not contain large  
10 portions of unrelated coding DNA, such as large chromosomal fragments or other functional genes or polypeptide coding regions. Of course, this refers to the DNA segment as originally isolated, and does not exclude genes or coding regions later added to the segment by the hand of man.

As will be recognized by the skilled artisan, polynucleotides may be  
15 single-stranded (coding or antisense) or double-stranded, and may be DNA (genomic, cDNA or synthetic) or RNA molecules. RNA molecules include HnRNA molecules, which contain introns and correspond to a DNA molecule in a one-to-one manner, and mRNA molecules, which do not contain introns. Additional coding or non-coding sequences may, but need not, be present within a polynucleotide of the present  
20 invention, and a polynucleotide may, but need not, be linked to other molecules and/or support materials.

Polynucleotides may comprise a native sequence (*i.e.*, an endogenous sequence that encodes a breast tumor protein or a portion thereof) or may comprise a variant, or a biological or antigenic functional equivalent of such a sequence.  
25 Polynucleotide variants may contain one or more substitutions, additions, deletions and/or insertions, as further described below, preferably such that the immunogenicity of the encoded polypeptide is not diminished, relative to a native tumor protein. The effect on the immunogenicity of the encoded polypeptide may generally be assessed as described herein. The term "variants" also encompasses homologous genes of  
30 xenogenic origin.

When comparing polynucleotide or polypeptide sequences, two sequences are said to be "identical" if the sequence of nucleotides or amino acids in the two sequences is the same when aligned for maximum correspondence, as described below. Comparisons between two sequences are typically performed by comparing the sequences over a comparison window to identify and compare local regions of sequence similarity. A "comparison window" as used herein, refers to a segment of at least about 20 contiguous positions, usually 30 to about 75, 40 to about 50, in which a sequence may be compared to a reference sequence of the same number of contiguous positions after the two sequences are optimally aligned.

Optimal alignment of sequences for comparison may be conducted using the Megalign program in the Lasergene suite of bioinformatics software (DNASTAR, Inc., Madison, WI), using default parameters. This program embodies several alignment schemes described in the following references: Dayhoff, M.O. (1978) A model of evolutionary change in proteins – Matrices for detecting distant relationships. In Dayhoff, M.O. (ed.) Atlas of Protein Sequence and Structure, National Biomedical Research Foundation, Washington DC Vol. 5, Suppl. 3, pp. 345-358; Hein J. (1990) Unified Approach to Alignment and Phylogenies pp. 626-645 *Methods in Enzymology* vol. 183, Academic Press, Inc., San Diego, CA; Higgins, D.G. and Sharp, P.M. (1989) *CABIOS* 5:151-153; Myers, E.W. and Muller W. (1988) *CABIOS* 4:11-17; Robinson, E.D. (1971) *Comb. Theor* 11:105; Santou, N. Nes, M. (1987) *Mol. Biol. Evol.* 4:406-425; Sneath, P.H.A. and Sokal, R.R. (1973) *Numerical Taxonomy – the Principles and Practice of Numerical Taxonomy*, Freeman Press, San Francisco, CA; Wilbur, W.J. and Lipman, D.J. (1983) *Proc. Natl. Acad. Sci. USA* 80:726-730.

Alternatively, optimal alignment of sequences for comparison may be conducted by the local identity algorithm of Smith and Waterman (1981) *Add. APL. Math* 2:482, by the identity alignment algorithm of Needleman and Wunsch (1970) *J. Mol. Biol.* 48:443, by the search for similarity methods of Pearson and Lipman (1988) *Proc. Natl. Acad. Sci. USA* 85: 2444, by computerized implementations of these algorithms (GAP, BESTFIT, BLAST, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group (GCG), 575 Science Dr., Madison, WI), or by inspection.

One preferred example of algorithms that are suitable for determining percent sequence identity and sequence similarity are the BLAST and BLAST 2.0 algorithms, which are described in Altschul et al. (1977) *Nucl. Acids Res.* 25:3389-3402 and Altschul et al. (1990) *J. Mol. Biol.* 215:403-410, respectively. BLAST and BLAST 2.0 can be used, for example with the parameters described herein, to determine percent sequence identity for the polynucleotides and polypeptides of the invention. Software for performing BLAST analyses is publicly available through the National Center for Biotechnology Information. In one illustrative example, cumulative scores can be calculated using, for nucleotide sequences, the parameters M (reward score for a pair of matching residues; always >0) and N (penalty score for mismatching residues; always <0). For amino acid sequences, a scoring matrix can be used to calculate the cumulative score. Extension of the word hits in each direction are halted when: the cumulative alignment score falls off by the quantity X from its maximum achieved value; the cumulative score goes to zero or below, due to the accumulation of one or more negative-scoring residue alignments; or the end of either sequence is reached. The BLAST algorithm parameters W, T and X determine the sensitivity and speed of the alignment. The BLASTN program (for nucleotide sequences) uses as defaults a wordlength (W) of 11, and expectation (E) of 10, and the BLOSUM62 scoring matrix (see Henikoff and Henikoff (1989) *Proc. Natl. Acad. Sci. USA* 89:10915) alignments, (B) of 50, expectation (E) of 10, M=5, N=-4 and a comparison of both strands.

Preferably, the "percentage of sequence identity" is determined by comparing two optimally aligned sequences over a window of comparison of at least 20 positions, wherein the portion of the polynucleotide or polypeptide sequence in the comparison window may comprise additions or deletions (*i.e.*, gaps) of 20 percent or less, usually 5 to 15 percent, or 10 to 12 percent, as compared to the reference sequences (which does not comprise additions or deletions) for optimal alignment of the two sequences. The percentage is calculated by determining the number of positions at which the identical nucleic acid bases or amino acid residue occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the reference sequence (*i.e.*, the window size) and multiplying the results by 100 to yield the percentage of sequence identity.

Therefore, the present invention encompasses polynucleotide and polypeptide sequences having substantial identity to the sequences disclosed herein, for example those comprising at least 50% sequence identity, preferably at least 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% or higher, sequence  
5 identity compared to a polynucleotide or polypeptide sequence of this invention using the methods described herein, (e.g., BLAST analysis using standard parameters, as described below). One skilled in this art will recognize that these values can be appropriately adjusted to determine corresponding identity of proteins encoded by two nucleotide sequences by taking into account codon degeneracy, amino acid similarity,  
10 reading frame positioning and the like.

In additional embodiments, the present invention provides isolated polynucleotides and polypeptides comprising various lengths of contiguous stretches of sequence identical to or complementary to one or more of the sequences disclosed herein. For example, polynucleotides are provided by this invention that comprise at  
15 least about 15, 20, 30, 40, 50, 75, 100, 150, 200, 300, 400, 500 or 1000 or more contiguous nucleotides of one or more of the sequences disclosed herein as well as all intermediate lengths there between. It will be readily understood that "intermediate lengths", in this context, means any length between the quoted values, such as 16, 17, 18, 19, *etc.*; 21, 22, 23, *etc.*; 30, 31, 32, *etc.*; 50, 51, 52, 53, *etc.*; 100, 101, 102, 103,  
20 *etc.*; 150, 151, 152, 153, *etc.*; including all integers through 200-500; 500-1,000, and the like.

The polynucleotides of the present invention, or fragments thereof, regardless of the length of the coding sequence itself, may be combined with other DNA sequences, such as promoters, polyadenylation signals, additional restriction  
25 enzyme sites, multiple cloning sites, other coding segments, and the like, such that their overall length may vary considerably. It is therefore contemplated that a nucleic acid fragment of almost any length may be employed, with the total length preferably being limited by the ease of preparation and use in the intended recombinant DNA protocol. For example, illustrative DNA segments with total lengths of about 10,000, about 5000,  
30 about 3000, about 2,000, about 1,000, about 500, about 200, about 100, about 50 base

pairs in length, and the like, (including all intermediate lengths) are contemplated to be useful in many implementations of this invention.

In other embodiments, the present invention is directed to polynucleotides that are capable of hybridizing under moderately stringent conditions to  
5 a polynucleotide sequence provided herein, or a fragment thereof, or a complementary sequence thereof. Hybridization techniques are well known in the art of molecular biology. For purposes of illustration, suitable moderately stringent conditions for testing the hybridization of a polynucleotide of this invention with other polynucleotides include prewashing in a solution of 5 X SSC, 0.5% SDS, 1.0 mM  
10 EDTA (pH 8.0); hybridizing at 50°C-65°C, 5 X SSC, overnight; followed by washing twice at 65°C for 20 minutes with each of 2X, 0.5X and 0.2X SSC containing 0.1% SDS.

Moreover, it will be appreciated by those of ordinary skill in the art that, as a result of the degeneracy of the genetic code, there are many nucleotide sequences  
15 that encode a polypeptide as described herein. Some of these polynucleotides bear minimal homology to the nucleotide sequence of any native gene. Nonetheless, polynucleotides that vary due to differences in codon usage are specifically contemplated by the present invention. Further, alleles of the genes comprising the polynucleotide sequences provided herein are within the scope of the present invention.  
20 Alleles are endogenous genes that are altered as a result of one or more mutations, such as deletions, additions and/or substitutions of nucleotides. The resulting mRNA and protein may, but need not, have an altered structure or function. Alleles may be identified using standard techniques (such as hybridization, amplification and/or database sequence comparison).

## 25 PROBES AND PRIMERS

In other embodiments of the present invention, the polynucleotide sequences provided herein can be advantageously used as probes or primers for nucleic acid hybridization. As such, it is contemplated that nucleic acid segments that comprise a sequence region of at least about 15 nucleotide long contiguous sequence that has the  
30 same sequence as, or is complementary to, a 15 nucleotide long contiguous sequence

disclosed herein will find particular utility. Longer contiguous identical or complementary sequences, *e.g.*, those of about 20, 30, 40, 50, 100, 200, 500, 1000 (including all intermediate lengths) and even up to full length sequences will also be of use in certain embodiments.

5                   The ability of such nucleic acid probes to specifically hybridize to a sequence of interest will enable them to be of use in detecting the presence of complementary sequences in a given sample. However, other uses are also envisioned, such as the use of the sequence information for the preparation of mutant species primers, or primers for use in preparing other genetic constructions.

10                  Polynucleotide molecules having sequence regions consisting of contiguous nucleotide stretches of 10-14, 15-20, 30, 50, or even of 100-200 nucleotides or so (including intermediate lengths as well), identical or complementary to a polynucleotide sequence disclosed herein, are particularly contemplated as hybridization probes for use in, *e.g.*, Southern and Northern blotting. This would allow  
15 a gene product, or fragment thereof, to be analyzed, both in diverse cell types and also in various bacterial cells. The total size of fragment, as well as the size of the complementary stretch(es), will ultimately depend on the intended use or application of the particular nucleic acid segment. Smaller fragments will generally find use in hybridization embodiments, wherein the length of the contiguous complementary  
20 region may be varied, such as between about 15 and about 100 nucleotides, but larger contiguous complementarity stretches may be used, according to the length complementary sequences one wishes to detect.

                  The use of a hybridization probe of about 15-25 nucleotides in length allows the formation of a duplex molecule that is both stable and selective. Molecules  
25 having contiguous complementary sequences over stretches greater than 15 bases in length are generally preferred, though, in order to increase stability and selectivity of the hybrid, and thereby improve the quality and degree of specific hybrid molecules obtained. One will generally prefer to design nucleic acid molecules having gene-complementary stretches of 15 to 25 contiguous nucleotides, or even longer where  
30 desired.



Hybridization probes may be selected from any portion of any of the sequences disclosed herein. All that is required is to review the sequence set forth in SEQ ID NO: 1-38, 42-204, 205, 207 and 210-290, or to any continuous portion of the sequence, from about 15-25 nucleotides in length up to and including the full length  
5 sequence, that one wishes to utilize as a probe or primer. The choice of probe and primer sequences may be governed by various factors. For example, one may wish to employ primers from towards the termini of the total sequence.

Small polynucleotide segments or fragments may be readily prepared by, for example, directly synthesizing the fragment by chemical means, as is commonly  
10 practiced using an automated oligonucleotide synthesizer. Also, fragments may be obtained by application of nucleic acid reproduction technology, such as the PCR<sup>TM</sup> technology of U. S. Patent 4,683,202 (incorporated herein by reference), by introducing selected sequences into recombinant vectors for recombinant production, and by other recombinant DNA techniques generally known to those of skill in the art of molecular  
15 biology.

The nucleotide sequences of the invention may be used for their ability to selectively form duplex molecules with complementary stretches of the entire gene or gene fragments of interest. Depending on the application envisioned, one will typically desire to employ varying conditions of hybridization to achieve varying degrees of  
20 selectivity of probe towards target sequence. For applications requiring high selectivity, one will typically desire to employ relatively stringent conditions to form the hybrids, *e.g.*, one will select relatively low salt and/or high temperature conditions, such as provided by a salt concentration of from about 0.02 M to about 0.15 M salt at temperatures of from about 50°C to about 70°C. Such selective conditions tolerate  
25 little, if any, mismatch between the probe and the template or target strand, and would be particularly suitable for isolating related sequences.

Of course, for some applications, for example, where one desires to prepare mutants employing a mutant primer strand hybridized to an underlying template, less stringent (reduced stringency) hybridization conditions will typically be  
30 needed in order to allow formation of the heteroduplex. In these circumstances, one may desire to employ salt conditions such as those of from about 0.15 M to about 0.9 M

salt, at temperatures ranging from about 20°C to about 55°C. Cross-hybridizing species can thereby be readily identified as positively hybridizing signals with respect to control hybridizations. In any case, it is generally appreciated that conditions can be rendered more stringent by the addition of increasing amounts of formamide, which serves to  
5 destabilize the hybrid duplex in the same manner as increased temperature. Thus, hybridization conditions can be readily manipulated, and thus will generally be a method of choice depending on the desired results.

#### POLYNUCLEOTIDE IDENTIFICATION AND CHARACTERIZATION

Polynucleotides may be identified, prepared and/or manipulated using  
10 any of a variety of well established techniques. For example, a polynucleotide may be identified, as described in more detail below, by screening a microarray of cDNAs for tumor-associated expression (*i.e.*, expression that is at least two fold greater in a tumor than in normal tissue, as determined using a representative assay provided herein). Such screens may be performed, for example, using a Synteni microarray (Palo Alto,  
15 CA) according to the manufacturer's instructions (and essentially as described by Schena et al., *Proc. Natl. Acad. Sci. USA* 93:10614-10619, 1996 and Heller et al., *Proc. Natl. Acad. Sci. USA* 94:2150-2155, 1997). Alternatively, polynucleotides may be amplified from cDNA prepared from cells expressing the proteins described herein, such as breast tumor cells. Such polynucleotides may be amplified via polymerase  
20 chain reaction (PCR). For this approach, sequence-specific primers may be designed based on the sequences provided herein, and may be purchased or synthesized.

An amplified portion of a polynucleotide of the present invention may be used to isolate a full length gene from a suitable library (*e.g.*, a breast tumor cDNA library) using well known techniques. Within such techniques, a library (cDNA or  
25 genomic) is screened using one or more polynucleotide probes or primers suitable for amplification. Preferably, a library is size-selected to include larger molecules. Random primed libraries may also be preferred for identifying 5' and upstream regions of genes. Genomic libraries are preferred for obtaining introns and extending 5' sequences.

For hybridization techniques, a partial sequence may be labeled (*e.g.*, by nick-translation or end-labeling with  $^{32}\text{P}$ ) using well known techniques. A bacterial or bacteriophage library is then generally screened by hybridizing filters containing denatured bacterial colonies (or lawns containing phage plaques) with the labeled probe  
5 (*see* Sambrook et al., *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Laboratories, Cold Spring Harbor, NY, 1989). Hybridizing colonies or plaques are selected and expanded, and the DNA is isolated for further analysis. cDNA clones may be analyzed to determine the amount of additional sequence by, for example, PCR using a primer from the partial sequence and a primer from the vector. Restriction maps and  
10 partial sequences may be generated to identify one or more overlapping clones. The complete sequence may then be determined using standard techniques, which may involve generating a series of deletion clones. The resulting overlapping sequences can then be assembled into a single contiguous sequence. A full length cDNA molecule can be generated by ligating suitable fragments, using well known techniques.

15 Alternatively, there are numerous amplification techniques for obtaining a full length coding sequence from a partial cDNA sequence. Within such techniques, amplification is generally performed via PCR. Any of a variety of commercially available kits may be used to perform the amplification step. Primers may be designed using, for example, software well known in the art. Primers are preferably 22-30  
20 nucleotides in length, have a GC content of at least 50% and anneal to the target sequence at temperatures of about 68°C to 72°C. The amplified region may be sequenced as described above, and overlapping sequences assembled into a contiguous sequence.

One such amplification technique is inverse PCR (*see* Triglia et al., *Nucl.*  
25 *Acids Res.* 16:8186, 1988), which uses restriction enzymes to generate a fragment in the known region of the gene. The fragment is then circularized by intramolecular ligation and used as a template for PCR with divergent primers derived from the known region. Within an alternative approach, sequences adjacent to a partial sequence may be retrieved by amplification with a primer to a linker sequence and a primer specific to a  
30 known region. The amplified sequences are typically subjected to a second round of amplification with the same linker primer and a second primer specific to the known

region. A variation on this procedure, which employs two primers that initiate extension in opposite directions from the known sequence, is described in WO 96/38591. Another such technique is known as "rapid amplification of cDNA ends" or RACE. This technique involves the use of an internal primer and an external primer, 5 which hybridizes to a polyA region or vector sequence, to identify sequences that are 5' and 3' of a known sequence. Additional techniques include capture PCR (Lagerstrom et al., *PCR Methods Applic. 1*:111-19, 1991) and walking PCR (Parker et al., *Nucl. Acids. Res. 19*:3055-60, 1991). Other methods employing amplification may also be employed to obtain a full length cDNA sequence.

10 In certain instances, it is possible to obtain a full length cDNA sequence by analysis of sequences provided in an expressed sequence tag (EST) database, such as that available from GenBank. Searches for overlapping ESTs may generally be performed using well known programs (*e.g.*, NCBI BLAST searches), and such ESTs may be used to generate a contiguous full length sequence. Full length DNA sequences 15 may also be obtained by analysis of genomic fragments.

#### POLYNUCLEOTIDE EXPRESSION IN HOST CELLS

In other embodiments of the invention, polynucleotide sequences or fragments thereof which encode polypeptides of the invention, or fusion proteins or functional equivalents thereof, may be used in recombinant DNA molecules to direct 20 expression of a polypeptide in appropriate host cells. Due to the inherent degeneracy of the genetic code, other DNA sequences that encode substantially the same or a functionally equivalent amino acid sequence may be produced and these sequences may be used to clone and express a given polypeptide.

As will be understood by those of skill in the art, it may be advantageous 25 in some instances to produce polypeptide-encoding nucleotide sequences possessing non-naturally occurring codons. For example, codons preferred by a particular prokaryotic or eukaryotic host can be selected to increase the rate of protein expression or to produce a recombinant RNA transcript having desirable properties, such as a half-life which is longer than that of a transcript generated from the naturally occurring 30 sequence.

Moreover, the polynucleotide sequences of the present invention can be engineered using methods generally known in the art in order to alter polypeptide encoding sequences for a variety of reasons, including but not limited to, alterations which modify the cloning, processing, and/or expression of the gene product. For example, DNA shuffling by random fragmentation and PCR reassembly of gene fragments and synthetic oligonucleotides may be used to engineer the nucleotide sequences. In addition, site-directed mutagenesis may be used to insert new restriction sites, alter glycosylation patterns, change codon preference, produce splice variants, or introduce mutations, and so forth.

10 In another embodiment of the invention, natural, modified, or recombinant nucleic acid sequences may be ligated to a heterologous sequence to encode a fusion protein. For example, to screen peptide libraries for inhibitors of polypeptide activity, it may be useful to encode a chimeric protein that can be recognized by a commercially available antibody. A fusion protein may also be engineered to contain a cleavage site located between the polypeptide-encoding sequence and the heterologous protein sequence, so that the polypeptide may be cleaved and purified away from the heterologous moiety.

Sequences encoding a desired polypeptide may be synthesized, in whole or in part, using chemical methods well known in the art (see Caruthers, M. H. et al. (1980) *Nucl. Acids Res. Symp. Ser.* 215-223, Horn, T. et al. (1980) *Nucl. Acids Res. Symp. Ser.* 225-232). Alternatively, the protein itself may be produced using chemical methods to synthesize the amino acid sequence of a polypeptide, or a portion thereof. For example, peptide synthesis can be performed using various solid-phase techniques (Roberge, J. Y. et al. (1995) *Science* 269:202-204) and automated synthesis may be achieved, for example, using the ABI 431A Peptide Synthesizer (Perkin Elmer, Palo Alto, CA).

A newly synthesized peptide may be substantially purified by preparative high performance liquid chromatography (e.g., Creighton, T. (1983) *Proteins, Structures and Molecular Principles*, WH Freeman and Co., New York, N.Y.) or other comparable techniques available in the art. The composition of the synthetic peptides may be confirmed by amino acid analysis or sequencing (e.g., the Edman

degradation procedure). Additionally, the amino acid sequence of a polypeptide, or any part thereof, may be altered during direct synthesis and/or combined using chemical methods with sequences from other proteins, or any part thereof, to produce a variant polypeptide.

5                   In order to express a desired polypeptide, the nucleotide sequences encoding the polypeptide, or functional equivalents, may be inserted into appropriate expression vector, i.e., a vector which contains the necessary elements for the transcription and translation of the inserted coding sequence. Methods which are well known to those skilled in the art may be used to construct expression vectors containing  
10 sequences encoding a polypeptide of interest and appropriate transcriptional and translational control elements. These methods include in vitro recombinant DNA techniques, synthetic techniques, and in vivo genetic recombination. Such techniques are described in Sambrook, J. et al. (1989) *Molecular Cloning, A Laboratory Manual*, Cold Spring Harbor Press, Plainview, N.Y., and Ausubel, F. M. et al. (1989) *Current*  
15 *Protocols in Molecular Biology*, John Wiley & Sons, New York. N.Y.

A variety of expression vector/host systems may be utilized to contain and express polynucleotide sequences. These include, but are not limited to, microorganisms such as bacteria transformed with recombinant bacteriophage, plasmid, or cosmid DNA expression vectors; yeast transformed with yeast expression vectors;  
20 insect cell systems infected with virus expression vectors (e.g., baculovirus); plant cell systems transformed with virus expression vectors (e.g., cauliflower mosaic virus, CaMV; tobacco mosaic virus, TMV) or with bacterial expression vectors (e.g., Ti or pBR322 plasmids); or animal cell systems.

The "control elements" or "regulatory sequences" present in an  
25 expression vector are those non-translated regions of the vector--enhancers, promoters, 5' and 3' untranslated regions--which interact with host cellular proteins to carry out transcription and translation. Such elements may vary in their strength and specificity. Depending on the vector system and host utilized, any number of suitable transcription and translation elements, including constitutive and inducible promoters, may be used.  
30 For example, when cloning in bacterial systems, inducible promoters such as the hybrid lacZ promoter of the PBLUESCRIPT phagemid (Stratagene, La Jolla, Calif.) or

PSPORT1 plasmid (Gibco BRL, Gaithersburg, MD) and the like may be used. In mammalian cell systems, promoters from mammalian genes or from mammalian viruses are generally preferred. If it is necessary to generate a cell line that contains multiple copies of the sequence encoding a polypeptide, vectors based on SV40 or EBV  
5 may be advantageously used with an appropriate selectable marker.

In bacterial systems, a number of expression vectors may be selected depending upon the use intended for the expressed polypeptide. For example, when large quantities are needed, for example for the induction of antibodies, vectors which direct high level expression of fusion proteins that are readily purified may be used.  
10 Such vectors include, but are not limited to, the multifunctional *E. coli* cloning and expression vectors such as BLUESCRIPT (Stratagene), in which the sequence encoding the polypeptide of interest may be ligated into the vector in frame with sequences for the amino-terminal Met and the subsequent 7 residues of  $\beta$ -galactosidase so that a hybrid protein is produced; pIN vectors (Van Heeke, G. and S. M. Schuster (1989) *J.*  
15 *Biol. Chem.* 264:5503-5509); and the like. pGEX Vectors (Promega, Madison, Wis.) may also be used to express foreign polypeptides as fusion proteins with glutathione S-transferase (GST). In general, such fusion proteins are soluble and can easily be purified from lysed cells by adsorption to glutathione-agarose beads followed by elution in the presence of free glutathione. Proteins made in such systems may be designed to include  
20 heparin, thrombin, or factor XA protease cleavage sites so that the cloned polypeptide of interest can be released from the GST moiety at will.

In the yeast, *Saccharomyces cerevisiae*, a number of vectors containing constitutive or inducible promoters such as alpha factor, alcohol oxidase, and PGH may be used. For reviews, see Ausubel et al. (supra) and Grant et al. (1987) *Methods*  
25 *Enzymol.* 153:516-544.

In cases where plant expression vectors are used, the expression of sequences encoding polypeptides may be driven by any of a number of promoters. For example, viral promoters such as the 35S and 19S promoters of CaMV may be used alone or in combination with the omega leader sequence from TMV (Takamatsu, N.  
30 (1987) *EMBO J.* 6:307-311. Alternatively, plant promoters such as the small subunit of RUBISCO or heat shock promoters may be used (Coruzzi, G. et al. (1984) *EMBO J.*

3:1671-1680; Broglie, R. et al. (1984) *Science* 224:838-843; and Winter, J. et al. (1991) *Results Probl. Cell Differ.* 17:85-105). These constructs can be introduced into plant cells by direct DNA transformation or pathogen-mediated transfection. Such techniques are described in a number of generally available reviews (see, for example, Hobbs, S. or  
5 Murry, L. E. in McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York, N.Y.; pp. 191-196).

An insect system may also be used to express a polypeptide of interest. For example, in one such system, *Autographa californica* nuclear polyhedrosis virus (AcNPV) is used as a vector to express foreign genes in *Spodoptera frugiperda* cells or  
10 in *Trichoplusia* larvae. The sequences encoding the polypeptide may be cloned into a non-essential region of the virus, such as the polyhedrin gene, and placed under control of the polyhedrin promoter. Successful insertion of the polypeptide-encoding sequence will render the polyhedrin gene inactive and produce recombinant virus lacking coat protein. The recombinant viruses may then be used to infect, for example, *S. frugiperda*  
15 cells or *Trichoplusia* larvae in which the polypeptide of interest may be expressed (Engelhard, E. K. et al. (1994) *Proc. Natl. Acad. Sci.* 91 :3224-3227).

In mammalian host cells, a number of viral-based expression systems are generally available. For example, in cases where an adenovirus is used as an expression vector, sequences encoding a polypeptide of interest may be ligated into an adenovirus  
20 transcription/translation complex consisting of the late promoter and tripartite leader sequence. Insertion in a non-essential E1 or E3 region of the viral genome may be used to obtain a viable virus which is capable of expressing the polypeptide in infected host cells (Logan, J. and Shenk, T. (1984) *Proc. Natl. Acad. Sci.* 81:3655-3659). In addition, transcription enhancers, such as the Rous sarcoma virus (RSV) enhancer, may be used  
25 to increase expression in mammalian host cells.

Specific initiation signals may also be used to achieve more efficient translation of sequences encoding a polypeptide of interest. Such signals include the ATG initiation codon and adjacent sequences. In cases where sequences encoding the polypeptide, its initiation codon, and upstream sequences are inserted into the  
30 appropriate expression vector, no additional transcriptional or translational control signals may be needed. However, in cases where only coding sequence, or a portion



thereof, is inserted, exogenous translational control signals including the ATG initiation codon should be provided. Furthermore, the initiation codon should be in the correct reading frame to ensure translation of the entire insert. Exogenous translational elements and initiation codons may be of various origins, both natural and synthetic.

- 5 The efficiency of expression may be enhanced by the inclusion of enhancers which are appropriate for the particular cell system which is used, such as those described in the literature (Scharf, D. et al. (1994) *Results Probl. Cell Differ.* 20:125-162).

In addition, a host cell strain may be chosen for its ability to modulate the expression of the inserted sequences or to process the expressed protein in the  
10 desired fashion. Such modifications of the polypeptide include, but are not limited to, acetylation, carboxylation, glycosylation, phosphorylation, lipidation, and acylation. Post-translational processing which cleaves a "prepro" form of the protein may also be used to facilitate correct insertion, folding and/or function. Different host cells such as CHO, HeLa, MDCK, HEK293, and WI38, which have specific cellular machinery and  
15 characteristic mechanisms for such post-translational activities, may be chosen to ensure the correct modification and processing of the foreign protein.

For long-term, high-yield production of recombinant proteins, stable expression is generally preferred. For example, cell lines which stably express a polynucleotide of interest may be transformed using expression vectors which may  
20 contain viral origins of replication and/or endogenous expression elements and a selectable marker gene on the same or on a separate vector. Following the introduction of the vector, cells may be allowed to grow for 1-2 days in an enriched media before they are switched to selective media. The purpose of the selectable marker is to confer resistance to selection, and its presence allows growth and recovery of cells which  
25 successfully express the introduced sequences. Resistant clones of stably transformed cells may be proliferated using tissue culture techniques appropriate to the cell type.

Any number of selection systems may be used to recover transformed cell lines. These include, but are not limited to, the herpes simplex virus thymidine kinase (Wigler, M. et al. (1977) *Cell* 11:223-32) and adenine phosphoribosyltransferase  
30 (Lowy, I. et al. (1990) *Cell* 22:817-23) genes which can be employed in tk.sup.- or aprt.sup.- cells, respectively. Also, antimetabolite, antibiotic or herbicide resistance can

be used as the basis for selection; for example, dhfr which confers resistance to methotrexate (Wigler, M. et al. (1980) *Proc. Natl. Acad. Sci.* 77:3567-70); npt, which confers resistance to the aminoglycosides, neomycin and G-418 (Colbere-Garapin, F. et al (1981) *J. Mol. Biol.* 150:1-14); and als or pat, which confer resistance to  
5 chlorsulfuron and phosphinotricin acetyltransferase, respectively (Murry, *supra*). Additional selectable genes have been described, for example, trpB, which allows cells to utilize indole in place of tryptophan, or hisD, which allows cells to utilize histinol in place of histidine (Hartman, S. C. and R. C. Mulligan (1988) *Proc. Natl. Acad. Sci.* 85:8047-51). Recently, the use of visible markers has gained popularity with such  
10 markers as anthocyanins, beta-glucuronidase and its substrate GUS, and luciferase and its substrate luciferin, being widely used not only to identify transformants, but also to quantify the amount of transient or stable protein expression attributable to a specific vector system (Rhodes, C. A. et al. (1995) *Methods Mol. Biol.* 55:121-131).

Although the presence/absence of marker gene expression suggests that  
15 the gene of interest is also present, its presence and expression may need to be confirmed. For example, if the sequence encoding a polypeptide is inserted within a marker gene sequence, recombinant cells containing sequences can be identified by the absence of marker gene function. Alternatively, a marker gene can be placed in tandem with a polypeptide-encoding sequence under the control of a single promoter.  
20 Expression of the marker gene in response to induction or selection usually indicates expression of the tandem gene as well.

Alternatively, host cells which contain and express a desired polynucleotide sequence may be identified by a variety of procedures known to those of skill in the art. These procedures include, but are not limited to, DNA-DNA or DNA-  
25 RNA hybridizations and protein bioassay or immunoassay techniques which include membrane, solution, or chip based technologies for the detection and/or quantification of nucleic acid or protein.

A variety of protocols for detecting and measuring the expression of polynucleotide-encoded products, using either polyclonal or monoclonal antibodies  
30 specific for the product are known in the art. Examples include enzyme-linked immunosorbent assay (ELISA), radioimmunoassay (RIA), and fluorescence activated

cell sorting (FACS). A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering epitopes on a given polypeptide may be preferred for some applications, but a competitive binding assay may also be employed. These and other assays are described, among other places, in Hampton, R. et al. (1990; 5 Serological Methods, a Laboratory Manual, APS Press, St Paul. Minn.) and Maddox, D. E. et al. (1983; *J. Exp. Med.* 158:1211-1216).

A wide variety of labels and conjugation techniques are known by those skilled in the art and may be used in various nucleic acid and amino acid assays. Means for producing labeled hybridization or PCR probes for detecting sequences related to 10 polynucleotides include oligolabeling, nick translation, end-labeling or PCR amplification using a labeled nucleotide. Alternatively, the sequences, or any portions thereof may be cloned into a vector for the production of an mRNA probe. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes in vitro by addition of an appropriate RNA polymerase such as T7, T3, or SP6 15 and labeled nucleotides. These procedures may be conducted using a variety of commercially available kits. Suitable reporter molecules or labels, which may be used include radionuclides, enzymes, fluorescent, chemiluminescent, or chromogenic agents as well as substrates, cofactors, inhibitors, magnetic particles, and the like.

Host cells transformed with a polynucleotide sequence of interest may be 20 cultured under conditions suitable for the expression and recovery of the protein from cell culture. The protein produced by a recombinant cell may be secreted or contained intracellularly depending on the sequence and/or the vector used. As will be understood by those of skill in the art, expression vectors containing polynucleotides of the invention may be designed to contain signal sequences which direct secretion of the 25 encoded polypeptide through a prokaryotic or eukaryotic cell membrane. Other recombinant constructions may be used to join sequences encoding a polypeptide of interest to nucleotide sequence encoding a polypeptide domain which will facilitate purification of soluble proteins. Such purification facilitating domains include, but are not limited to, metal chelating peptides such as histidine-tryptophan modules that allow 30 purification on immobilized metals, protein A domains that allow purification on immobilized immunoglobulin, and the domain utilized in the FLAGS extension/affinity

purification system (Immunex Corp., Seattle, Wash.). The inclusion of cleavable linker sequences such as those specific for Factor XA or enterokinase (Invitrogen, San Diego, Calif.) between the purification domain and the encoded polypeptide may be used to facilitate purification. One such expression vector provides for expression of a fusion  
5 protein containing a polypeptide of interest and a nucleic acid encoding 6 histidine residues preceding a thioredoxin or an enterokinase cleavage site. The histidine residues facilitate purification on IMIAC (immobilized metal ion affinity chromatography) as described in Porath, J. et al. (1992, *Prot. Exp. Purif.* 3:263-281) while the enterokinase cleavage site provides a means for purifying the desired polypeptide from the fusion  
10 protein. A discussion of vectors which contain fusion proteins is provided in Kroll, D. J. et al. (1993; *DNA Cell Biol.* 12:441-453).

In addition to recombinant production methods, polypeptides of the invention, and fragments thereof, may be produced by direct peptide synthesis using solid-phase techniques (Merrifield J. (1963) *J. Am. Chem. Soc.* 85:2149-2154). Protein  
15 synthesis may be performed using manual techniques or by automation. Automated synthesis may be achieved, for example, using Applied Biosystems 431A Peptide Synthesizer (Perkin Elmer). Alternatively, various fragments may be chemically synthesized separately and combined using chemical methods to produce the full length molecule.

## 20 SITE-SPECIFIC MUTAGENESIS

Site-specific mutagenesis is a technique useful in the preparation of individual peptides, or biologically functional equivalent polypeptides, through specific mutagenesis of the underlying polynucleotides that encode them. The technique, well-known to those of skill in the art, further provides a ready ability to prepare and  
25 test sequence variants, for example, incorporating one or more of the foregoing considerations, by introducing one or more nucleotide sequence changes into the DNA. Site-specific mutagenesis allows the production of mutants through the use of specific oligonucleotide sequences which encode the DNA sequence of the desired mutation, as well as a sufficient number of adjacent nucleotides, to provide a primer sequence of  
30 sufficient size and sequence complexity to form a stable duplex on both sides of the

deletion junction being traversed. Mutations may be employed in a selected polynucleotide sequence to improve, alter, decrease, modify, or otherwise change the properties of the polynucleotide itself, and/or alter the properties, activity, composition, stability, or primary sequence of the encoded polypeptide.

5 In certain embodiments of the present invention, the inventors contemplate the mutagenesis of the disclosed polynucleotide sequences to alter one or more properties of the encoded polypeptide, such as the antigenicity of a polypeptide vaccine. The techniques of site-specific mutagenesis are well-known in the art, and are widely used to create variants of both polypeptides and polynucleotides. For example,  
10 site-specific mutagenesis is often used to alter a specific portion of a DNA molecule. In such embodiments, a primer comprising typically about 14 to about 25 nucleotides or so in length is employed, with about 5 to about 10 residues on both sides of the junction of the sequence being altered.

As will be appreciated by those of skill in the art, site-specific  
15 mutagenesis techniques have often employed a phage vector that exists in both a single stranded and double stranded form. Typical vectors useful in site-directed mutagenesis include vectors such as the M13 phage. These phage are readily commercially-available and their use is generally well-known to those skilled in the art. Double-stranded plasmids are also routinely employed in site directed mutagenesis that  
20 eliminates the step of transferring the gene of interest from a plasmid to a phage.

In general, site-directed mutagenesis in accordance herewith is performed by first obtaining a single-stranded vector or melting apart of two strands of a double-stranded vector that includes within its sequence a DNA sequence that encodes the desired peptide. An oligonucleotide primer bearing the desired mutated sequence is  
25 prepared, generally synthetically. This primer is then annealed with the single-stranded vector, and subjected to DNA polymerizing enzymes such as *E. coli* polymerase I Klenow fragment, in order to complete the synthesis of the mutation-bearing strand. Thus, a heteroduplex is formed wherein one strand encodes the original non-mutated sequence and the second strand bears the desired mutation. This heteroduplex vector is  
30 then used to transform appropriate cells, such as *E. coli* cells, and clones are selected which include recombinant vectors bearing the mutated sequence arrangement.

The preparation of sequence variants of the selected peptide-encoding DNA segments using site-directed mutagenesis provides a means of producing potentially useful species and is not meant to be limiting as there are other ways in which sequence variants of peptides and the DNA sequences encoding them may be  
5 obtained. For example, recombinant vectors encoding the desired peptide sequence may be treated with mutagenic agents, such as hydroxylamine, to obtain sequence variants. Specific details regarding these methods and protocols are found in the teachings of Maloy *et al.*, 1994; Segal, 1976; Prokop and Bajpai, 1991; Kuby, 1994; and Maniatis *et al.*, 1982, each incorporated herein by reference, for that purpose.

10 As used herein, the term "oligonucleotide directed mutagenesis procedure" refers to template-dependent processes and vector-mediated propagation which result in an increase in the concentration of a specific nucleic acid molecule relative to its initial concentration, or in an increase in the concentration of a detectable signal, such as amplification. As used herein, the term "oligonucleotide directed  
15 mutagenesis procedure" is intended to refer to a process that involves the template-dependent extension of a primer molecule. The term template dependent process refers to nucleic acid synthesis of an RNA or a DNA molecule wherein the sequence of the newly synthesized strand of nucleic acid is dictated by the well-known rules of complementary base pairing (see, for example, Watson, 1987). Typically,  
20 vector mediated methodologies involve the introduction of the nucleic acid fragment into a DNA or RNA vector, the clonal amplification of the vector, and the recovery of the amplified nucleic acid fragment. Examples of such methodologies are provided by U. S. Patent No. 4,237,224, specifically incorporated herein by reference in its entirety.

#### **POLYNUCLEOTIDE AMPLIFICATION TECHNIQUES**

25 A number of template dependent processes are available to amplify the target sequences of interest present in a sample. One of the best known amplification methods is the polymerase chain reaction (PCR™) which is described in detail in U.S. Patent Nos. 4,683,195, 4,683,202 and 4,800,159, each of which is incorporated herein by reference in its entirety. Briefly, in PCR™, two primer sequences are prepared  
30 which are complementary to regions on opposite complementary strands of the target

sequence. An excess of deoxynucleoside triphosphates is added to a reaction mixture along with a DNA polymerase (e.g., *Taq* polymerase). If the target sequence is present in a sample, the primers will bind to the target and the polymerase will cause the primers to be extended along the target sequence by adding on nucleotides. By raising  
5 and lowering the temperature of the reaction mixture, the extended primers will dissociate from the target to form reaction products, excess primers will bind to the target and to the reaction product and the process is repeated. Preferably reverse transcription and PCR<sup>TM</sup> amplification procedure may be performed in order to quantify the amount of mRNA amplified. Polymerase chain reaction methodologies are well  
10 known in the art.

Another method for amplification is the ligase chain reaction (referred to as LCR), disclosed in Eur. Pat. Appl. Publ. No. 320,308 (specifically incorporated herein by reference in its entirety). In LCR, two complementary probe pairs are prepared, and in the presence of the target sequence, each pair will bind to opposite  
15 complementary strands of the target such that they abut. In the presence of a ligase, the two probe pairs will link to form a single unit. By temperature cycling, as in PCR<sup>TM</sup>, bound ligated units dissociate from the target and then serve as "target sequences" for ligation of excess probe pairs. U.S. Patent No. 4,883,750, incorporated herein by reference in its entirety, describes an alternative method of amplification similar to LCR  
20 for binding probe pairs to a target sequence.

Qbeta Replicase, described in PCT Intl. Pat. Appl. Publ. No. PCT/US87/00880, incorporated herein by reference in its entirety, may also be used as still another amplification method in the present invention. In this method, a replicative sequence of RNA that has a region complementary to that of a target is added to a  
25 sample in the presence of an RNA polymerase. The polymerase will copy the replicative sequence that can then be detected.

An isothermal amplification method, in which restriction endonucleases and ligases are used to achieve the amplification of target molecules that contain nucleotide 5'-[ $\alpha$ -thio]triphosphates in one strand of a restriction site (Walker *et al.*,  
30 1992, incorporated herein by reference in its entirety), may also be useful in the amplification of nucleic acids in the present invention.

Strand Displacement Amplification (SDA) is another method of carrying out isothermal amplification of nucleic acids which involves multiple rounds of strand displacement and synthesis, *i.e.* nick translation. A similar method, called Repair Chain Reaction (RCR) is another method of amplification which may be useful in the present invention and is involves annealing several probes throughout a region targeted for amplification, followed by a repair reaction in which only two of the four bases are present. The other two bases can be added as biotinylated derivatives for easy detection. A similar approach is used in SDA.

Sequences can also be detected using a cyclic probe reaction (CPR). In CPR, a probe having a 3' and 5' sequences of non-target DNA and an internal or "middle" sequence of the target protein specific RNA is hybridized to DNA which is present in a sample. Upon hybridization, the reaction is treated with RNaseH, and the products of the probe are identified as distinctive products by generating a signal that is released after digestion. The original template is annealed to another cycling probe and the reaction is repeated. Thus, CPR involves amplifying a signal generated by hybridization of a probe to a target gene specific expressed nucleic acid.

Still other amplification methods described in Great Britain Pat. Appl. No. 2 202 328, and in PCT Intl. Pat. Appl. Publ. No. PCT/US89/01025, each of which is incorporated herein by reference in its entirety, may be used in accordance with the present invention. In the former application, "modified" primers are used in a PCR-like, template and enzyme dependent synthesis. The primers may be modified by labeling with a capture moiety (*e.g.*, biotin) and/or a detector moiety (*e.g.*, enzyme). In the latter application, an excess of labeled probes is added to a sample. In the presence of the target sequence, the probe binds and is cleaved catalytically. After cleavage, the target sequence is released intact to be bound by excess probe. Cleavage of the labeled probe signals the presence of the target sequence.

Other nucleic acid amplification procedures include transcription-based amplification systems (TAS) (Kwoh *et al.*, 1989; PCT Intl. Pat. Appl. Publ. No. WO 88/10315, incorporated herein by reference in its entirety), including nucleic acid sequence based amplification (NASBA) and 3SR. In NASBA, the nucleic acids can be prepared for amplification by standard phenol/chloroform extraction, heat denaturation



of a sample, treatment with lysis buffer and minispin columns for isolation of DNA and RNA or guanidinium chloride extraction of RNA. These amplification techniques involve annealing a primer that has sequences specific to the target sequence. Following polymerization, DNA/RNA hybrids are digested with RNase H while double stranded DNA molecules are heat-denatured again. In either case the single stranded DNA is made fully double stranded by addition of second target-specific primer, followed by polymerization. The double stranded DNA molecules are then multiply transcribed by a polymerase such as T7 or SP6. In an isothermal cyclic reaction, the RNAs are reverse transcribed into DNA, and transcribed once again with a polymerase such as T7 or SP6. The resulting products, whether truncated or complete, indicate target-specific sequences.

Eur. Pat. Appl. Publ. No. 329,822, incorporated herein by reference in its entirety, disclose a nucleic acid amplification process involving cyclically synthesizing single-stranded RNA ("ssRNA"), ssDNA, and double-stranded DNA (dsDNA), which may be used in accordance with the present invention. The ssRNA is a first template for a first primer oligonucleotide, which is elongated by reverse transcriptase (RNA-dependent DNA polymerase). The RNA is then removed from resulting DNA:RNA duplex by the action of ribonuclease H (RNase H, an RNase specific for RNA in a duplex with either DNA or RNA). The resultant ssDNA is a second template for a second primer, which also includes the sequences of an RNA polymerase promoter (exemplified by T7 RNA polymerase) 5' to its homology to its template. This primer is then extended by DNA polymerase (exemplified by the large "Klenow" fragment of *E. coli* DNA polymerase I), resulting as a double-stranded DNA ("dsDNA") molecule, having a sequence identical to that of the original RNA between the primers and having additionally, at one end, a promoter sequence. This promoter sequence can be used by the appropriate RNA polymerase to make many RNA copies of the DNA. These copies can then re-enter the cycle leading to very swift amplification. With proper choice of enzymes, this amplification can be done isothermally without addition of enzymes at each cycle. Because of the cyclical nature of this process, the starting sequence can be chosen to be in the form of either DNA or RNA.

PCT Intl. Pat. Appl. Publ. No. WO 89/06700, incorporated herein by reference in its entirety, disclose a nucleic acid sequence amplification scheme based on the hybridization of a promoter/primer sequence to a target single-stranded DNA ("ssDNA") followed by transcription of many RNA copies of the sequence. This  
5 scheme is not cyclic; i.e. new templates are not produced from the resultant RNA transcripts. Other amplification methods include "RACE" (Frohman, 1990), and "one-sided PCR" (Ohara, 1989) which are well-known to those of skill in the art.

Methods based on ligation of two (or more) oligonucleotides in the presence of nucleic acid having the sequence of the resulting "di-oligonucleotide",  
10 thereby amplifying the di-oligonucleotide (Wu and Dean, 1996, incorporated herein by reference in its entirety), may also be used in the amplification of DNA sequences of the present invention.

#### BIOLOGICAL FUNCTIONAL EQUIVALENTS

Modification and changes may be made in the structure of the  
15 polynucleotides and polypeptides of the present invention and still obtain a functional molecule that encodes a polypeptide with desirable characteristics. As mentioned above, it is often desirable to introduce one or more mutations into a specific polynucleotide sequence. In certain circumstances, the resulting encoded polypeptide sequence is altered by this mutation, or in other cases, the sequence of the polypeptide  
20 is unchanged by one or more mutations in the encoding polynucleotide.

When it is desirable to alter the amino acid sequence of a polypeptide to create an equivalent, or even an improved, second-generation molecule, the amino acid changes may be achieved by changing one or more of the codons of the encoding DNA sequence, according to Table 1.

25 For example, certain amino acids may be substituted for other amino acids in a protein structure without appreciable loss of interactive binding capacity with structures such as, for example, antigen-binding regions of antibodies or binding sites on substrate molecules. Since it is the interactive capacity and nature of a protein that defines that protein's biological functional activity, certain amino acid sequence  
30 substitutions can be made in a protein sequence, and, of course, its underlying DNA

coding sequence, and nevertheless obtain a protein with like properties. It is thus contemplated by the inventors that various changes may be made in the peptide sequences of the disclosed compositions, or corresponding DNA sequences which encode said peptides without appreciable loss of their biological utility or activity.

5

TABLE 1

Amino Acids			Codons						
Alanine	Ala	A	GCA	GCC	GCG	GCU			
Cysteine	Cys	C	UGC	UGU					
Aspartic acid	Asp	D	GAC	GAU					
Glutamic acid	Glu	E	GAA	GAG					
Phenylalanine	Phe	F	UUC	UUU					
Glycine	Gly	G	GGA	GGC	GGG	GGU			
Histidine	His	H	CAC	CAU					
Isoleucine	Ile	I	AUA	AUC	AUU				
Lysine	Lys	K	AAA	AAG					
Leucine	Leu	L	UUA	UUG	CUA	CUC	CUG	CUU	
Methionine	Met	M	AUG						
Asparagine	Asn	N	AAC	AAU					
Proline	Pro	P	CCA	CCC	CCG	CCU			
Glutamine	Gln	Q	CAA	CAG					
Arginine	Arg	R	AGA	AGG	CGA	CGC	CGG	CGU	
Serine	Ser	S	AGC	AGU	UCA	UCC	UCG	UCU	
Threonine	Thr	T	ACA	ACC	ACG	ACU			
Valine	Val	V	GUA	GUC	GUG	GUU			
Tryptophan	Trp	W	UGG						
Tyrosine	Tyr	Y	UAC	UAU					

In making such changes, the hydropathic index of amino acids may be considered. The importance of the hydropathic amino acid index in conferring interactive biologic function on a protein is generally understood in the art (Kyte and Doolittle, 1982, incorporated herein by reference). It is accepted that the relative

Doolittle, 1982, incorporated herein by reference). It is accepted that the relative hydrophobic character of the amino acid contributes to the secondary structure of the resultant protein, which in turn defines the interaction of the protein with other molecules, for example, enzymes, substrates, receptors, DNA, antibodies, antigens, and the like. Each amino acid has been assigned a hydrophobic index on the basis of its hydrophobicity and charge characteristics (Kyte and Doolittle, 1982). These values are: isoleucine (+4.5); valine (+4.2); leucine (+3.8); phenylalanine (+2.8); cysteine/cystine (+2.5); methionine (+1.9); alanine (+1.8); glycine (-0.4); threonine (-0.7); serine (-0.8); tryptophan (-0.9); tyrosine (-1.3); proline (-1.6); histidine (-3.2); glutamate (-3.5); glutamine (-3.5); aspartate (-3.5); asparagine (-3.5); lysine (-3.9); and arginine (-4.5).

It is known in the art that certain amino acids may be substituted by other amino acids having a similar hydrophobic index or score and still result in a protein with similar biological activity, *i.e.* still obtain a biological functionally equivalent protein. In making such changes, the substitution of amino acids whose hydrophobic indices are within  $\pm 2$  is preferred, those within  $\pm 1$  are particularly preferred, and those within  $\pm 0.5$  are even more particularly preferred. It is also understood in the art that the substitution of like amino acids can be made effectively on the basis of hydrophilicity. U. S. Patent 4,554,101 (specifically incorporated herein by reference in its entirety), states that the greatest local average hydrophilicity of a protein, as governed by the hydrophilicity of its adjacent amino acids, correlates with a biological property of the protein.

As detailed in U. S. Patent 4,554,101, the following hydrophilicity values have been assigned to amino acid residues: arginine (+3.0); lysine (+3.0); aspartate (+3.0  $\pm$  1); glutamate (+3.0  $\pm$  1); serine (+0.3); asparagine (+0.2); glutamine (+0.2); glycine (0); threonine (-0.4); proline (-0.5  $\pm$  1); alanine (-0.5); histidine (-0.5); cysteine (-1.0); methionine (-1.3); valine (-1.5); leucine (-1.8); isoleucine (-1.8); tyrosine (-2.3); phenylalanine (-2.5); tryptophan (-3.4). It is understood that an amino acid can be substituted for another having a similar hydrophilicity value and still obtain a biologically equivalent, and in particular, an immunologically equivalent protein. In such changes, the substitution of amino acids whose hydrophilicity values are within  $\pm 2$

is preferred, those within  $\pm 1$  are particularly preferred, and those within  $\pm 0.5$  are even more particularly preferred.

As outlined above, amino acid substitutions are generally therefore based on the relative similarity of the amino acid side-chain substituents, for example, their hydrophobicity, hydrophilicity, charge, size, and the like. Exemplary substitutions that take various of the foregoing characteristics into consideration are well known to those of skill in the art and include: arginine and lysine; glutamate and aspartate; serine and threonine; glutamine and asparagine; and valine, leucine and isoleucine.

In addition, any polynucleotide may be further modified to increase stability *in vivo*. Possible modifications include, but are not limited to, the addition of flanking sequences at the 5' and/or 3' ends; the use of phosphorothioate or 2' O-methyl rather than phosphodiesterase linkages in the backbone; and/or the inclusion of nontraditional bases such as inosine, queosine and wybutosine, as well as acetyl-methyl-, thio- and other modified forms of adenine, cytidine, guanine, thymine and uridine.

#### IN VIVO POLYNUCLEOTIDE DELIVERY TECHNIQUES

In additional embodiments, genetic constructs comprising one or more of the polynucleotides of the invention are introduced into cells *in vivo*. This may be achieved using any of a variety of well known approaches, several of which are outlined below for the purpose of illustration.

##### 1. ADENOVIRUS

One of the preferred methods for *in vivo* delivery of one or more nucleic acid sequences involves the use of an adenovirus expression vector. "Adenovirus expression vector" is meant to include those constructs containing adenovirus sequences sufficient to (a) support packaging of the construct and (b) to express a polynucleotide that has been cloned therein in a sense or antisense orientation. Of course, in the context of an antisense construct, expression does not require that the gene product be synthesized.

The expression vector comprises a genetically engineered form of an adenovirus. Knowledge of the genetic organization of adenovirus, a 36 kb, linear, double-stranded DNA virus, allows substitution of large pieces of adenoviral DNA with foreign sequences up to 7 kb (Grunhaus and Horwitz, 1992). In contrast to retrovirus, the adenoviral infection of host cells does not result in chromosomal integration because adenoviral DNA can replicate in an episomal manner without potential genotoxicity. Also, adenoviruses are structurally stable, and no genome rearrangement has been detected after extensive amplification. Adenovirus can infect virtually all epithelial cells regardless of their cell cycle stage. So far, adenoviral infection appears to be linked only to mild disease such as acute respiratory disease in humans.

Adenovirus is particularly suitable for use as a gene transfer vector because of its mid-sized genome, ease of manipulation, high titer, wide target-cell range and high infectivity. Both ends of the viral genome contain 100-200 base pair inverted repeats (ITRs), which are *cis* elements necessary for viral DNA replication and packaging. The early (E) and late (L) regions of the genome contain different transcription units that are divided by the onset of viral DNA replication. The E1 region (E1A and E1B) encodes proteins responsible for the regulation of transcription of the viral genome and a few cellular genes. The expression of the E2 region (E2A and E2B) results in the synthesis of the proteins for viral DNA replication. These proteins are involved in DNA replication, late gene expression and host cell shut-off (Renan, 1990). The products of the late genes, including the majority of the viral capsid proteins, are expressed only after significant processing of a single primary transcript issued by the major late promoter (MLP). The MLP, (located at 16.8 m.u.) is particularly efficient during the late phase of infection, and all the mRNA's issued from this promoter possess a 5'-tripartite leader (TPL) sequence which makes them preferred mRNA's for translation.

In a current system, recombinant adenovirus is generated from homologous recombination between shuttle vector and provirus vector. Due to the possible recombination between two proviral vectors, wild-type adenovirus may be generated from this process. Therefore, it is critical to isolate a single clone of virus from an individual plaque and examine its genomic structure.

Generation and propagation of the current adenovirus vectors, which are replication deficient, depend on a unique helper cell line, designated 293, which was transformed from human embryonic kidney cells by Ad5 DNA fragments and constitutively expresses E1 proteins (Graham *et al.*, 1977). Since the E3 region is  
5 dispensable from the adenovirus genome (Jones and Shenk, 1978), the current adenovirus vectors, with the help of 293 cells, carry foreign DNA in either the E1, the D3 or both regions (Graham and Prevec, 1991). In nature, adenovirus can package approximately 105% of the wild-type genome (Ghosh-Choudhury *et al.*, 1987), providing capacity for about 2 extra kB of DNA. Combined with the approximately 5.5  
10 kB of DNA that is replaceable in the E1 and E3 regions, the maximum capacity of the current adenovirus vector is under 7.5 kB, or about 15% of the total length of the vector. More than 80% of the adenovirus viral genome remains in the vector backbone and is the source of vector-borne cytotoxicity. Also, the replication deficiency of the E1-deleted virus is incomplete. For example, leakage of viral gene expression has been  
15 observed with the currently available vectors at high multiplicities of infection (MOI) (Mulligan, 1993).

Helper cell lines may be derived from human cells such as human embryonic kidney cells, muscle cells, hematopoietic cells or other human embryonic mesenchymal or epithelial cells. Alternatively, the helper cells may be derived from the  
20 cells of other mammalian species that are permissive for human adenovirus. Such cells include, *e.g.*, Vero cells or other monkey embryonic mesenchymal or epithelial cells. As stated above, the currently preferred helper cell line is 293.

Recently, Racher *et al.* (1995) disclosed improved methods for culturing 293 cells and propagating adenovirus. In one format, natural cell aggregates are grown  
25 by inoculating individual cells into 1 liter siliconized spinner flasks (Techne, Cambridge, UK) containing 100-200 ml of medium. Following stirring at 40 rpm, the cell viability is estimated with trypan blue. In another format, Fibra-Cel microcarriers (Bibby Sterlin, Stone, UK) (5 g/l) is employed as follows. A cell inoculum, resuspended in 5 ml of medium, is added to the carrier (50 ml) in a 250 ml Erlenmeyer  
30 flask and left stationary, with occasional agitation, for 1 to 4 h. The medium is then replaced with 50 ml of fresh medium and shaking initiated. For virus production, cells

are allowed to grow to about 80% confluence, after which time the medium is replaced (to 25% of the final volume) and adenovirus added at an MOI of 0.05. Cultures are left stationary overnight, following which the volume is increased to 100% and shaking commenced for another 72 h.

5           Other than the requirement that the adenovirus vector be replication defective, or at least conditionally defective, the nature of the adenovirus vector is not believed to be crucial to the successful practice of the invention. The adenovirus may be of any of the 42 different known serotypes or subgroups A-F. Adenovirus type 5 of subgroup C is the preferred starting material in order to obtain a conditional replication-  
10   defective adenovirus vector for use in the present invention, since Adenovirus type 5 is a human adenovirus about which a great deal of biochemical and genetic information is known, and it has historically been used for most constructions employing adenovirus as a vector.

          As stated above, the typical vector according to the present invention is  
15   replication defective and will not have an adenovirus E1 region. Thus, it will be most convenient to introduce the polynucleotide encoding the gene of interest at the position from which the E1-coding sequences have been removed. However, the position of insertion of the construct within the adenovirus sequences is not critical to the invention. The polynucleotide encoding the gene of interest may also be inserted in lieu  
20   of the deleted E3 region in E3 replacement vectors as described by Karlsson *et al.* (1986) or in the E4 region where a helper cell line or helper virus complements the E4 defect.

          Adenovirus is easy to grow and manipulate and exhibits broad host range *in vitro* and *in vivo*. This group of viruses can be obtained in high titers, *e.g.*,  $10^9$ - $10^{11}$   
25   plaque-forming units per ml, and they are highly infective. The life cycle of adenovirus does not require integration into the host cell genome. The foreign genes delivered by adenovirus vectors are episomal and, therefore, have low genotoxicity to host cells. No side effects have been reported in studies of vaccination with wild-type adenovirus (Couch *et al.*, 1963; Top *et al.*, 1971), demonstrating their safety and therapeutic  
30   potential as *in vivo* gene transfer vectors.



Adenovirus vectors have been used in eukaryotic gene expression (Levrero *et al.*, 1991; Gomez-Foix *et al.*, 1992) and vaccine development (Grunhaus and Horwitz, 1992; Graham and Prevec, 1992). Recently, animal studies suggested that recombinant adenovirus could be used for gene therapy (Stratford-Perricaudet and Perricaudet, 1991; Stratford-Perricaudet *et al.*, 1990; Rich *et al.*, 1993). Studies in administering recombinant adenovirus to different tissues include trachea instillation (Rosenfeld *et al.*, 1991; Rosenfeld *et al.*, 1992), muscle injection (Ragot *et al.*, 1993), peripheral intravenous injections (Herz and Gerard, 1993) and stereotactic inoculation into the brain (Le Gal La Salle *et al.*, 1993).

## 10 2. RETROVIRUSES

The retroviruses are a group of single-stranded RNA viruses characterized by an ability to convert their RNA to double-stranded DNA in infected cells by a process of reverse-transcription (Coffin, 1990). The resulting DNA then stably integrates into cellular chromosomes as a provirus and directs synthesis of viral proteins. The integration results in the retention of the viral gene sequences in the recipient cell and its descendants. The retroviral genome contains three genes, gag, pol, and env that code for capsid proteins, polymerase enzyme, and envelope components, respectively. A sequence found upstream from the gag gene contains a signal for packaging of the genome into virions. Two long terminal repeat (LTR) sequences are present at the 5' and 3' ends of the viral genome. These contain strong promoter and enhancer sequences and are also required for integration in the host cell genome (Coffin, 1990).

In order to construct a retroviral vector, a nucleic acid encoding one or more oligonucleotide or polynucleotide sequences of interest is inserted into the viral genome in the place of certain viral sequences to produce a virus that is replication-defective. In order to produce virions, a packaging cell line containing the gag, pol, and env genes but without the LTR and packaging components is constructed (Mann *et al.*, 1983). When a recombinant plasmid containing a cDNA, together with the retroviral LTR and packaging sequences is introduced into this cell line (by calcium phosphate precipitation for example), the packaging sequence allows the RNA transcript of the

recombinant plasmid to be packaged into viral particles, which are then secreted into the culture media (Nicolas and Rubenstein, 1988; Temin, 1986; Mann *et al.*, 1983). The media containing the recombinant retroviruses is then collected, optionally concentrated, and used for gene transfer. Retroviral vectors are able to infect a broad  
5 variety of cell types. However, integration and stable expression require the division of host cells (Paskind *et al.*, 1975).

A novel approach designed to allow specific targeting of retrovirus vectors was recently developed based on the chemical modification of a retrovirus by the chemical addition of lactose residues to the viral envelope. This modification could  
10 permit the specific infection of hepatocytes *via* sialoglycoprotein receptors.

A different approach to targeting of recombinant retroviruses was designed in which biotinylated antibodies against a retroviral envelope protein and against a specific cell receptor were used. The antibodies were coupled *via* the biotin components by using streptavidin (Roux *et al.*, 1989). Using antibodies against major  
15 histocompatibility complex class I and class II antigens, they demonstrated the infection of a variety of human cells that bore those surface antigens with an ecotropic virus *in vitro* (Roux *et al.*, 1989).

### 3. ADENO-ASSOCIATED VIRUSES

AAV (Ridgeway, 1988; Hermonat and Muzycka, 1984) is a parovirus,  
20 discovered as a contamination of adenoviral stocks. It is a ubiquitous virus (antibodies are present in 85% of the US human population) that has not been linked to any disease. It is also classified as a dependovirus, because its replication is dependent on the presence of a helper virus, such as adenovirus. Five serotypes have been isolated, of which AAV-2 is the best characterized. AAV has a single-stranded linear DNA that is  
25 encapsidated into capsid proteins VP1, VP2 and VP3 to form an icosahedral virion of 20 to 24 nm in diameter (Muzycka and McLaughlin, 1988).

The AAV DNA is approximately 4.7 kilobases long. It contains two open reading frames and is flanked by two ITRs. There are two major genes in the AAV genome: *rep* and *cap*. The *rep* gene codes for proteins responsible for viral  
30 replications, whereas *cap* codes for capsid protein VP1-3. Each ITR forms a T-shaped

hairpin structure. These terminal repeats are the only essential *cis* components of the AAV for chromosomal integration. Therefore, the AAV can be used as a vector with all viral coding sequences removed and replaced by the cassette of genes for delivery. Three viral promoters have been identified and named p5, p19, and p40, according to  
5 their map position. Transcription from p5 and p19 results in production of rep proteins, and transcription from p40 produces the capsid proteins (Hermonat and Muzyczka, 1984).

There are several factors that prompted researchers to study the possibility of using rAAV as an expression vector. One is that the requirements for  
10 delivering a gene to integrate into the host chromosome are surprisingly few. It is necessary to have the 145-bp ITRs, which are only 6% of the AAV genome. This leaves room in the vector to assemble a 4.5-kb DNA insertion. While this carrying capacity may prevent the AAV from delivering large genes, it is amply suited for delivering the antisense constructs of the present invention.

15 AAV is also a good choice of delivery vehicles due to its safety. There is a relatively complicated rescue mechanism: not only wild type adenovirus but also AAV genes are required to mobilize rAAV. Likewise, AAV is not pathogenic and not associated with any disease. The removal of viral coding sequences minimizes immune reactions to viral gene expression, and therefore, rAAV does not evoke an inflammatory  
20 response.

#### 4. OTHER VIRAL VECTORS AS EXPRESSION CONSTRUCTS

Other viral vectors may be employed as expression constructs in the present invention for the delivery of oligonucleotide or polynucleotide sequences to a host cell. Vectors derived from viruses such as vaccinia virus (Ridgeway, 1988; Coupar  
25 *et al.*, 1988), lentiviruses, polio viruses and herpes viruses may be employed. They offer several attractive features for various mammalian cells (Friedmann, 1989; Ridgeway, 1988; Coupar *et al.*, 1988; Horwich *et al.*, 1990).

With the recent recognition of defective hepatitis B viruses, new insight was gained into the structure-function relationship of different viral sequences. *In vitro*  
30 studies showed that the virus could retain the ability for helper-dependent packaging

and reverse transcription despite the deletion of up to 80% of its genome (Horwich *et al.*, 1990). This suggested that large portions of the genome could be replaced with foreign genetic material. The hepatotropism and persistence (integration) were particularly attractive properties for liver-directed gene transfer. Chang *et al.* (1991) introduced the chloramphenicol acetyltransferase (CAT) gene into duck hepatitis B virus genome in the place of the polymerase, surface, and pre-surface coding sequences. It was cotransfected with wild-type virus into an avian hepatoma cell line. Culture media containing high titers of the recombinant virus were used to infect primary duckling hepatocytes. Stable CAT gene expression was detected for at least 24 days after transfection (Chang *et al.*, 1991).

## 5. NON-VIRAL VECTORS

In order to effect expression of the oligonucleotide or polynucleotide sequences of the present invention, the expression construct must be delivered into a cell. This delivery may be accomplished *in vitro*, as in laboratory procedures for transforming cells lines, or *in vivo* or *ex vivo*, as in the treatment of certain disease states. As described above, one preferred mechanism for delivery is *via* viral infection where the expression construct is encapsulated in an infectious viral particle.

Once the expression construct has been delivered into the cell the nucleic acid encoding the desired oligonucleotide or polynucleotide sequences may be positioned and expressed at different sites. In certain embodiments, the nucleic acid encoding the construct may be stably integrated into the genome of the cell. This integration may be in the specific location and orientation *via* homologous recombination (gene replacement) or it may be integrated in a random, non-specific location (gene augmentation). In yet further embodiments, the nucleic acid may be stably maintained in the cell as a separate, episomal segment of DNA. Such nucleic acid segments or "episomes" encode sequences sufficient to permit maintenance and replication independent of or in synchronization with the host cell cycle. How the expression construct is delivered to a cell and where in the cell the nucleic acid remains is dependent on the type of expression construct employed.

In certain embodiments of the invention, the expression construct comprising one or more oligonucleotide or polynucleotide sequences may simply consist of naked recombinant DNA or plasmids. Transfer of the construct may be performed by any of the methods mentioned above which physically or chemically permeabilize the cell membrane. This is particularly applicable for transfer *in vitro* but it may be applied to *in vivo* use as well. Dubensky *et al.* (1984) successfully injected polyomavirus DNA in the form of calcium phosphate precipitates into liver and spleen of adult and newborn mice demonstrating active viral replication and acute infection. Benvenisty and Reshef (1986) also demonstrated that direct intraperitoneal injection of calcium phosphate-precipitated plasmids results in expression of the transfected genes. It is envisioned that DNA encoding a gene of interest may also be transferred in a similar manner *in vivo* and express the gene product.

Another embodiment of the invention for transferring a naked DNA expression construct into cells may involve particle bombardment. This method depends on the ability to accelerate DNA-coated microprojectiles to a high velocity allowing them to pierce cell membranes and enter cells without killing them (Klein *et al.*, 1987). Several devices for accelerating small particles have been developed. One such device relies on a high voltage discharge to generate an electrical current, which in turn provides the motive force (Yang *et al.*, 1990). The microprojectiles used have consisted of biologically inert substances such as tungsten or gold beads.

Selected organs including the liver, skin, and muscle tissue of rats and mice have been bombarded *in vivo* (Yang *et al.*, 1990; Zelenin *et al.*, 1991). This may require surgical exposure of the tissue or cells, to eliminate any intervening tissue between the gun and the target organ, *i.e. ex vivo* treatment. Again, DNA encoding a particular gene may be delivered *via* this method and still be incorporated by the present invention.

#### ANTISENSE OLIGONUCLEOTIDES

The end result of the flow of genetic information is the synthesis of protein. DNA is transcribed by polymerases into messenger RNA and translated on the ribosome to yield a folded, functional protein. Thus there are several steps along the

route where protein synthesis can be inhibited. The native DNA segment coding for a polypeptide described herein, as all such mammalian DNA strands, has two strands: a sense strand and an antisense strand held together by hydrogen bonding. The messenger RNA coding for polypeptide has the same nucleotide sequence as the sense  
5 DNA strand except that the DNA thymidine is replaced by uridine. Thus, synthetic antisense nucleotide sequences will bind to a mRNA and inhibit expression of the protein encoded by that mRNA.

The targeting of antisense oligonucleotides to mRNA is thus one mechanism to shut down protein synthesis, and, consequently, represents a powerful  
10 and targeted therapeutic approach. For example, the synthesis of polygalacturonase and the muscarine type 2 acetylcholine receptor are inhibited by antisense oligonucleotides directed to their respective mRNA sequences (U. S. Patent 5,739,119 and U. S. Patent 5,759,829, each specifically incorporated herein by reference in its entirety). Further, examples of antisense inhibition have been demonstrated with the  
15 nuclear protein cyclin, the multiple drug resistance gene (MDG1), ICAM-1, E-selectin, STK-1, striatal GABA<sub>A</sub> receptor and human EGF (Jaskulski *et al.*, 1988; Vasanthakumar and Ahmed, 1989; Peris *et al.*, 1998; U. S. Patent 5,801,154; U. S. Patent 5,789,573; U. S. Patent 5,718,709 and U. S. Patent 5,610,288, each specifically incorporated herein by reference in its entirety). Antisense constructs have also been  
20 described that inhibit and can be used to treat a variety of abnormal cellular proliferations, *e.g.* cancer (U. S. Patent 5,747,470; U. S. Patent 5,591,317 and U. S. Patent 5,783,683, each specifically incorporated herein by reference in its entirety).

Therefore, in exemplary embodiments, the invention provides oligonucleotide sequences that comprise all, or a portion of, any sequence that is  
25 capable of specifically binding to polynucleotide sequence described herein, or a complement thereof. In one embodiment, the antisense oligonucleotides comprise DNA or derivatives thereof. In another embodiment, the oligonucleotides comprise RNA or derivatives thereof. In a third embodiment, the oligonucleotides are modified DNAs comprising a phosphorothioated modified backbone. In a fourth embodiment, the  
30 oligonucleotide sequences comprise peptide nucleic acids or derivatives thereof. In each case, preferred compositions comprise a sequence region that is complementary,

and more preferably substantially-complementary, and even more preferably, completely complementary to one or more portions of polynucleotides disclosed herein.

Selection of antisense compositions specific for a given gene sequence is based upon analysis of the chosen target sequence (*i.e.* in these illustrative examples the  
5 rat and human sequences) and determination of secondary structure,  $T_m$ , binding energy, relative stability, and antisense compositions were selected based upon their relative inability to form dimers, hairpins, or other secondary structures that would reduce or prohibit specific binding to the target mRNA in a host cell.

Highly preferred target regions of the mRNA, are those which are at or  
10 near the AUG translation initiation codon, and those sequences which were substantially complementary to 5' regions of the mRNA. These secondary structure analyses and target site selection considerations were performed using v.4 of the OLIGO primer analysis software (Rychlik, 1997) and the BLASTN 2.0.5 algorithm software (Altschul *et al.*, 1997).

15 The use of an antisense delivery method employing a short peptide vector, termed MPG (27 residues), is also contemplated. The MPG peptide contains a hydrophobic domain derived from the fusion sequence of HIV gp41 and a hydrophilic domain from the nuclear localization sequence of SV40 T-antigen (Morris *et al.*, 1997). It has been demonstrated that several molecules of the MPG peptide coat the antisense  
20 oligonucleotides and can be delivered into cultured mammalian cells in less than 1 hour with relatively high efficiency (90%). Further, the interaction with MPG strongly increases both the stability of the oligonucleotide to nuclease and the ability to cross the plasma membrane (Morris *et al.*, 1997).

#### **RIBOZYMES**

25 Although proteins traditionally have been used for catalysis of nucleic acids, another class of macromolecules has emerged as useful in this endeavor. Ribozymes are RNA-protein complexes that cleave nucleic acids in a site-specific fashion. Ribozymes have specific catalytic domains that possess endonuclease activity (Kim and Cech, 1987; Gerlach *et al.*, 1987; Forster and Symons, 1987). For example, a  
30 large number of ribozymes accelerate phosphoester transfer reactions with a high degree

of specificity, often cleaving only one of several phosphoesters in an oligonucleotide substrate (Cech *et al.*, 1981; Michel and Westhof, 1990; Reinhold-Hurek and Shub, 1992). This specificity has been attributed to the requirement that the substrate bind via specific base-pairing interactions to the internal guide sequence ("IGS") of the ribozyme  
5 prior to chemical reaction.

Ribozyme catalysis has primarily been observed as part of sequence-specific cleavage/ligation reactions involving nucleic acids (Joyce, 1989; Cech *et al.*, 1981). For example, U. S. Patent No. 5,354,855 (specifically incorporated herein by reference) reports that certain ribozymes can act as endonucleases with a sequence  
10 specificity greater than that of known ribonucleases and approaching that of the DNA restriction enzymes. Thus, sequence-specific ribozyme-mediated inhibition of gene expression may be particularly suited to therapeutic applications (Scanlon *et al.*, 1991; Sarver *et al.*, 1990). Recently, it was reported that ribozymes elicited genetic changes in some cells lines to which they were applied; the altered genes included the oncogenes  
15 H-*ras*, c-*fos* and genes of HIV. Most of this work involved the modification of a target mRNA, based on a specific mutant codon that is cleaved by a specific ribozyme.

Six basic varieties of naturally-occurring enzymatic RNAs are known presently. Each can catalyze the hydrolysis of RNA phosphodiester bonds *in trans* (and thus can cleave other RNA molecules) under physiological conditions. In general,  
20 enzymatic nucleic acids act by first binding to a target RNA. Such binding occurs through the target binding portion of a enzymatic nucleic acid which is held in close proximity to an enzymatic portion of the molecule that acts to cleave the target RNA. Thus, the enzymatic nucleic acid first recognizes and then binds a target RNA through complementary base-pairing, and once bound to the correct site, acts enzymatically to  
25 cut the target RNA. Strategic cleavage of such a target RNA will destroy its ability to direct synthesis of an encoded protein. After an enzymatic nucleic acid has bound and cleaved its RNA target, it is released from that RNA to search for another target and can repeatedly bind and cleave new targets.

The enzymatic nature of a ribozyme is advantageous over many  
30 technologies, such as antisense technology (where a nucleic acid molecule simply binds to a nucleic acid target to block its translation) since the concentration of ribozyme



necessary to affect a therapeutic treatment is lower than that of an antisense oligonucleotide. This advantage reflects the ability of the ribozyme to act enzymatically. Thus, a single ribozyme molecule is able to cleave many molecules of target RNA. In addition, the ribozyme is a highly specific inhibitor, with the specificity  
5 of inhibition depending not only on the base pairing mechanism of binding to the target RNA, but also on the mechanism of target RNA cleavage. Single mismatches, or base-substitutions, near the site of cleavage can completely eliminate catalytic activity of a ribozyme. Similar mismatches in antisense molecules do not prevent their action (Woolf *et al.*, 1992). Thus, the specificity of action of a ribozyme is greater than that of  
10 an antisense oligonucleotide binding the same RNA site.

The enzymatic nucleic acid molecule may be formed in a hammerhead, hairpin, a hepatitis  $\delta$  virus, group I intron or RNaseP RNA (in association with an RNA guide sequence) or Neurospora VS RNA motif. Examples of hammerhead motifs are described by Rossi *et al.* (1992). Examples of hairpin motifs are described by Hampel  
15 *et al.* (Eur. Pat. Appl. Publ. No. EP 0360257), Hampel and Tritz (1989), Hampel *et al.* (1990) and U. S. Patent 5,631,359 (specifically incorporated herein by reference). An example of the hepatitis  $\delta$  virus motif is described by Perrotta and Been (1992); an example of the RNaseP motif is described by Guerrier-Takada *et al.* (1983); Neurospora VS RNA ribozyme motif is described by Collins (Saville and Collins, 1990; Saville and  
20 Collins, 1991; Collins and Olive, 1993); and an example of the Group I intron is described in (U. S. Patent 4,987,071, specifically incorporated herein by reference). All that is important in an enzymatic nucleic acid molecule of this invention is that it has a specific substrate binding site which is complementary to one or more of the target gene RNA regions, and that it have nucleotide sequences within or surrounding that substrate  
25 binding site which impart an RNA cleaving activity to the molecule. Thus the ribozyme constructs need not be limited to specific motifs mentioned herein.

In certain embodiments, it may be important to produce enzymatic cleaving agents which exhibit a high degree of specificity for the RNA of a desired target, such as one of the sequences disclosed herein. The enzymatic nucleic acid  
30 molecule is preferably targeted to a highly conserved sequence region of a target mRNA. Such enzymatic nucleic acid molecules can be delivered exogenously to

specific cells as required. Alternatively, the ribozymes can be expressed from DNA or RNA vectors that are delivered to specific cells.

Small enzymatic nucleic acid motifs (*e.g.*, of the hammerhead or the hairpin structure) may also be used for exogenous delivery. The simple structure of these molecules increases the ability of the enzymatic nucleic acid to invade targeted regions of the mRNA structure. Alternatively, catalytic RNA molecules can be expressed within cells from eukaryotic promoters (*e.g.*, Scanlon *et al.*, 1991; Kashani-Sabet *et al.*, 1992; Dropulic *et al.*, 1992; Weerasinghe *et al.*, 1991; Ojwang *et al.*, 1992; Chen *et al.*, 1992; Sarver *et al.*, 1990). Those skilled in the art realize that any ribozyme can be expressed in eukaryotic cells from the appropriate DNA vector. The activity of such ribozymes can be augmented by their release from the primary transcript by a second ribozyme (Int. Pat. Appl. Publ. No. WO 93/23569, and Int. Pat. Appl. Publ. No. WO 94/02595, both hereby incorporated by reference; Ohkawa *et al.*, 1992; Taira *et al.*, 1991; and Ventura *et al.*, 1993).

Ribozymes may be added directly, or can be complexed with cationic lipids, lipid complexes, packaged within liposomes, or otherwise delivered to target cells. The RNA or RNA complexes can be locally administered to relevant tissues *ex vivo*, or *in vivo* through injection, aerosol inhalation, infusion pump or stent, with or without their incorporation in biopolymers.

Ribozymes may be designed as described in Int. Pat. Appl. Publ. No. WO 93/23569 and Int. Pat. Appl. Publ. No. WO 94/02595, each specifically incorporated herein by reference) and synthesized to be tested *in vitro* and *in vivo*, as described. Such ribozymes can also be optimized for delivery. While specific examples are provided, those in the art will recognize that equivalent RNA targets in other species can be utilized when necessary.

Hammerhead or hairpin ribozymes may be individually analyzed by computer folding (Jaeger *et al.*, 1989) to assess whether the ribozyme sequences fold into the appropriate secondary structure. Those ribozymes with unfavorable intramolecular interactions between the binding arms and the catalytic core are eliminated from consideration. Varying binding arm lengths can be chosen to optimize

activity. Generally, at least 5 or so bases on each arm are able to bind to, or otherwise interact with, the target RNA.

Ribozymes of the hammerhead or hairpin motif may be designed to anneal to various sites in the mRNA message, and can be chemically synthesized. The method of synthesis used follows the procedure for normal RNA synthesis as described in Usman *et al.* (1987) and in Scaringe *et al.* (1990) and makes use of common nucleic acid protecting and coupling groups, such as dimethoxytrityl at the 5'-end, and phosphoramidites at the 3'-end. Average stepwise coupling yields are typically >98%. Hairpin ribozymes may be synthesized in two parts and annealed to reconstruct an active ribozyme (Chowrira and Burke, 1992). Ribozymes may be modified extensively to enhance stability by modification with nuclease resistant groups, for example, 2'-amino, 2'-C-allyl, 2'-fluoro, 2'-O-methyl, 2'-H (for a review see *e.g.*, Usman and Cedergren, 1992). Ribozymes may be purified by gel electrophoresis using general methods or by high pressure liquid chromatography and resuspended in water.

Ribozyme activity can be optimized by altering the length of the ribozyme binding arms, or chemically synthesizing ribozymes with modifications that prevent their degradation by serum ribonucleases (see *e.g.*, Int. Pat. Appl. Publ. No. WO 92/07065; Perrault *et al.*, 1990; Pieken *et al.*, 1991; Usman and Cedergren, 1992; Int. Pat. Appl. Publ. No. WO 93/15187; Int. Pat. Appl. Publ. No. WO 91/03162; Eur. Pat. Appl. Publ. No. 92110298.4; U. S. Patent 5,334,711; and Int. Pat. Appl. Publ. No. WO 94/13688, which describe various chemical modifications that can be made to the sugar moieties of enzymatic RNA molecules), modifications which enhance their efficacy in cells, and removal of stem II bases to shorten RNA synthesis times and reduce chemical requirements.

Sullivan *et al.* (Int. Pat. Appl. Publ. No. WO 94/02595) describes the general methods for delivery of enzymatic RNA molecules. Ribozymes may be administered to cells by a variety of methods known to those familiar to the art, including, but not restricted to, encapsulation in liposomes, by iontophoresis, or by incorporation into other vehicles, such as hydrogels, cyclodextrins, biodegradable nanocapsules, and bioadhesive microspheres. For some indications, ribozymes may be directly delivered *ex vivo* to cells or tissues with or without the aforementioned vehicles.

Alternatively, the RNA/vehicle combination may be locally delivered by direct inhalation, by direct injection or by use of a catheter, infusion pump or stent. Other routes of delivery include, but are not limited to, intravascular, intramuscular, subcutaneous or joint injection, aerosol inhalation, oral (tablet or pill form), topical, 5 systemic, ocular, intraperitoneal and/or intrathecal delivery. More detailed descriptions of ribozyme delivery and administration are provided in Int. Pat. Appl. Publ. No. WO 94/02595 and Int. Pat. Appl. Publ. No. WO 93/23569, each specifically incorporated herein by reference.

Another means of accumulating high concentrations of a ribozyme(s) 10 within cells is to incorporate the ribozyme-encoding sequences into a DNA expression vector. Transcription of the ribozyme sequences are driven from a promoter for eukaryotic RNA polymerase I (pol I), RNA polymerase II (pol II), or RNA polymerase III (pol III). Transcripts from pol II or pol III promoters will be expressed at high levels in all cells; the levels of a given pol II promoter in a given cell type will depend on the 15 nature of the gene regulatory sequences (enhancers, silencers, *etc.*) present nearby. Prokaryotic RNA polymerase promoters may also be used, providing that the prokaryotic RNA polymerase enzyme is expressed in the appropriate cells (Elroy-Stein and Moss, 1990; Gao and Huang, 1993; Lieber *et al.*, 1993; Zhou *et al.*, 1990). Ribozymes expressed from such promoters can function in mammalian cells (*e.g.* 20 Kashani-Saber *et al.*, 1992; Ojwang *et al.*, 1992; Chen *et al.*, 1992; Yu *et al.*, 1993; L'Huillier *et al.*, 1992; Lisiewicz *et al.*, 1993). Such transcription units can be incorporated into a variety of vectors for introduction into mammalian cells, including but not restricted to, plasmid DNA vectors, viral DNA vectors (such as adenovirus or adeno-associated vectors), or viral RNA vectors (such as retroviral, semliki forest virus, 25 sindbis virus vectors).

Ribozymes may be used as diagnostic tools to examine genetic drift and mutations within diseased cells. They can also be used to assess levels of the target RNA molecule. The close relationship between ribozyme activity and the structure of the target RNA allows the detection of mutations in any region of the molecule which 30 alters the base-pairing and three-dimensional structure of the target RNA. By using multiple ribozymes, one may map nucleotide changes which are important to RNA

structure and function *in vitro*, as well as in cells and tissues. Cleavage of target RNAs with ribozymes may be used to inhibit gene expression and define the role (essentially) of specified gene products in the progression of disease. In this manner, other genetic targets may be defined as important mediators of the disease. These studies will lead to  
5 better treatment of the disease progression by affording the possibility of combinational therapies (*e.g.*, multiple ribozymes targeted to different genes, ribozymes coupled with known small molecule inhibitors, or intermittent treatment with combinations of ribozymes and/or other chemical or biological molecules). Other *in vitro* uses of ribozymes are well known in the art, and include detection of the presence of mRNA  
10 associated with an IL-5 related condition. Such RNA is detected by determining the presence of a cleavage product after treatment with a ribozyme using standard methodology.

#### PEPTIDE NUCLEIC ACIDS

In certain embodiments, the inventors contemplate the use of peptide  
15 nucleic acids (PNAs) in the practice of the methods of the invention. PNA is a DNA mimic in which the nucleobases are attached to a pseudopeptide backbone (Good and Nielsen, 1997). PNA is able to be utilized in a number methods that traditionally have used RNA or DNA. Often PNA sequences perform better in techniques than the corresponding RNA or DNA sequences and have utilities that are not inherent to RNA  
20 or DNA. A review of PNA including methods of making, characteristics of, and methods of using, is provided by Corey (1997) and is incorporated herein by reference. As such, in certain embodiments, one may prepare PNA sequences that are complementary to one or more portions of the ACE mRNA sequence, and such PNA compositions may be used to regulate, alter, decrease, or reduce the translation of ACE-  
25 specific mRNA, and thereby alter the level of ACE activity in a host cell to which such PNA compositions have been administered.

PNAs have 2-aminoethyl-glycine linkages replacing the normal phosphodiester backbone of DNA (Nielsen *et al.*, 1991; Hanvey *et al.*, 1992; Hyrup and Nielsen, 1996; Neilsen, 1996). This chemistry has three important consequences:  
30 firstly, in contrast to DNA or phosphorothioate oligonucleotides, PNAs are neutral

molecules; secondly, PNAs are achiral, which avoids the need to develop a stereoselective synthesis; and thirdly, PNA synthesis uses standard Boc (Dueholm *et al.*, 1994) or Fmoc (Thomson *et al.*, 1995) protocols for solid-phase peptide synthesis, although other methods, including a modified Merrifield method, have been used  
5 (Christensen *et al.*, 1995).

PNA monomers or ready-made oligomers are commercially available from PerSeptive Biosystems (Framingham, MA). PNA syntheses by either Boc or Fmoc protocols are straightforward using manual or automated protocols (Norton *et al.*, 1995). The manual protocol lends itself to the production of chemically modified PNAs  
10 or the simultaneous synthesis of families of closely related PNAs.

As with peptide synthesis, the success of a particular PNA synthesis will depend on the properties of the chosen sequence. For example, while in theory PNAs can incorporate any combination of nucleotide bases, the presence of adjacent purines can lead to deletions of one or more residues in the product. In expectation of this  
15 difficulty, it is suggested that, in producing PNAs with adjacent purines, one should repeat the coupling of residues likely to be added inefficiently. This should be followed by the purification of PNAs by reverse-phase high-pressure liquid chromatography (Norton *et al.*, 1995) providing yields and purity of product similar to those observed during the synthesis of peptides.

20 Modifications of PNAs for a given application may be accomplished by coupling amino acids during solid-phase synthesis or by attaching compounds that contain a carboxylic acid group to the exposed N-terminal amine. Alternatively, PNAs can be modified after synthesis by coupling to an introduced lysine or cysteine. The ease with which PNAs can be modified facilitates optimization for better solubility or  
25 for specific functional requirements. Once synthesized, the identity of PNAs and their derivatives can be confirmed by mass spectrometry. Several studies have made and utilized modifications of PNAs (Norton *et al.*, 1995; Haaime *et al.*, 1996; Stetsenko *et al.*, 1996; Petersen *et al.*, 1995; Ulmann *et al.*, 1996; Koch *et al.*, 1995; Orum *et al.*, 1995; Footer *et al.*, 1996; Griffith *et al.*, 1995; Kremsky *et al.*, 1996; Pardridge *et al.*,  
30 1995; Boffa *et al.*, 1995; Landsdorp *et al.*, 1996; Gambacorti-Passerini *et al.*, 1996; Armitage *et al.*, 1997; Seeger *et al.*, 1997; Ruskowski *et al.*, 1997). U.S. Patent No.

5,700,922 discusses PNA-DNA-PNA chimeric molecules and their uses in diagnostics, modulating protein in organisms, and treatment of conditions susceptible to therapeutics.

In contrast to DNA and RNA, which contain negatively charged linkages, the PNA backbone is neutral. In spite of this dramatic alteration, PNAs recognize complementary DNA and RNA by Watson-Crick pairing (Egholm *et al.*, 1993), validating the initial modeling by Nielsen *et al.* (1991). PNAs lack 3' to 5' polarity and can bind in either parallel or antiparallel fashion, with the antiparallel mode being preferred (Egholm *et al.*, 1993).

Hybridization of DNA oligonucleotides to DNA and RNA is destabilized by electrostatic repulsion between the negatively charged phosphate backbones of the complementary strands. By contrast, the absence of charge repulsion in PNA-DNA or PNA-RNA duplexes increases the melting temperature ( $T_m$ ) and reduces the dependence of  $T_m$  on the concentration of mono- or divalent cations (Nielsen *et al.*, 1991). The enhanced rate and affinity of hybridization are significant because they are responsible for the surprising ability of PNAs to perform strand invasion of complementary sequences within relaxed double-stranded DNA. In addition, the efficient hybridization at inverted repeats suggests that PNAs can recognize secondary structure effectively within double-stranded DNA. Enhanced recognition also occurs with PNAs immobilized on surfaces, and Wang *et al.* have shown that support-bound PNAs can be used to detect hybridization events (Wang *et al.*, 1996).

One might expect that tight binding of PNAs to complementary sequences would also increase binding to similar (but not identical) sequences, reducing the sequence specificity of PNA recognition. As with DNA hybridization, however, selective recognition can be achieved by balancing oligomer length and incubation temperature. Moreover, selective hybridization of PNAs is encouraged by PNA-DNA hybridization being less tolerant of base mismatches than DNA-DNA hybridization. For example, a single mismatch within a 16 bp PNA-DNA duplex can reduce the  $T_m$  by up to 15°C (Egholm *et al.*, 1993). This high level of discrimination has allowed the development of several PNA-based strategies for the analysis of point mutations (Wang

*et al.*, 1996; Carlsson *et al.*, 1996; Thiede *et al.*, 1996; Webb and Hurskainen, 1996; Perry-O'Keefe *et al.*, 1996).

High-affinity binding provides clear advantages for molecular recognition and the development of new applications for PNAs. For example, 11-13  
5 nucleotide PNAs inhibit the activity of telomerase, a ribonucleo-protein that extends telomere ends using an essential RNA template, while the analogous DNA oligomers do not (Norton *et al.*, 1996).

Neutral PNAs are more hydrophobic than analogous DNA oligomers, and this can lead to difficulty solubilizing them at neutral pH, especially if the PNAs  
10 have a high purine content or if they have the potential to form secondary structures. Their solubility can be enhanced by attaching one or more positive charges to the PNA termini (Nielsen *et al.*, 1991).

Findings by Allfrey and colleagues suggest that strand invasion will occur spontaneously at sequences within chromosomal DNA (Boffa *et al.*, 1995; Boffa  
15 *et al.*, 1996). These studies targeted PNAs to triplet repeats of the nucleotides CAG and used this recognition to purify transcriptionally active DNA (Boffa *et al.*, 1995) and to inhibit transcription (Boffa *et al.*, 1996). This result suggests that if PNAs can be delivered within cells then they will have the potential to be general sequence-specific regulators of gene expression. Studies and reviews concerning the use of PNAs as  
20 antisense and anti-gene agents include Nielsen *et al.* (1993b), Hanvey *et al.* (1992), and Good and Nielsen (1997). Koppelhus *et al.* (1997) have used PNAs to inhibit HIV-1 inverse transcription, showing that PNAs may be used for antiviral therapies.

Methods of characterizing the antisense binding properties of PNAs are discussed in Rose (1993) and Jensen *et al.* (1997). Rose uses capillary gel  
25 electrophoresis to determine binding of PNAs to their complementary oligonucleotide, measuring the relative binding kinetics and stoichiometry. Similar types of measurements were made by Jensen *et al.* using BIAcore™ technology.

Other applications of PNAs include use in DNA strand invasion (Nielsen *et al.*, 1991), antisense inhibition (Hanvey *et al.*, 1992), mutational analysis (Orum *et al.*, 1993), enhancers of transcription (Mollegaard *et al.*, 1994), nucleic acid purification  
30 (Orum *et al.*, 1995), isolation of transcriptionally active genes (Boffa *et al.*, 1995),



blocking of transcription factor binding (Vickers *et al.*, 1995), genome cleavage (Veselkov *et al.*, 1996), biosensors (Wang *et al.*, 1996), *in situ* hybridization (Thisted *et al.*, 1996), and in a alternative to Southern blotting (Perry-O'Keefe, 1996).

#### POLYPEPTIDE COMPOSITIONS

5                   The present invention, in other aspects, provides polypeptide compositions. Generally, a polypeptide of the invention will be an isolated polypeptide (or an epitope, variant, or active fragment thereof) derived from a mammalian species. Preferably, the polypeptide is encoded by a polynucleotide sequence disclosed herein or a sequence which hybridizes under moderately stringent conditions to a polynucleotide  
10                   sequence disclosed herein. Alternatively, the polypeptide may be defined as a polypeptide which comprises a contiguous amino acid sequence from an amino acid sequence disclosed herein, or which polypeptide comprises an entire amino acid sequence disclosed herein.

                  In the present invention, a polypeptide composition is also understood to  
15                   comprise one or more polypeptides that are immunologically reactive with antibodies generated against a polypeptide of the invention, or to active fragments, or to variants or biological functional equivalents thereof.

                  Likewise, a polypeptide composition of the present invention is understood to comprise one or more polypeptides that are capable of eliciting antibodies  
20                   that are immunologically reactive with one or more polypeptides encoded by one or more contiguous nucleic acid sequences contained in SEQ ID NO: 1-38, 42-204, 205, 207 and 210-290, or to active fragments, or to variants thereof, or to one or more nucleic acid sequences which hybridize to one or more of these sequences under conditions of moderate to high stringency.

25                   As used herein, an active fragment of a polypeptide includes a whole or a portion of a polypeptide which is modified by conventional techniques, *e.g.*, mutagenesis, or by addition, deletion, or substitution, but which active fragment exhibits substantially the same structure function, antigenicity, etc., as a polypeptide as described herein.

In certain illustrative embodiments, the polypeptides of the invention will comprise at least an immunogenic portion of a breast tumor protein or a variant thereof, as described herein. As noted above, a "breast tumor protein" is a protein that is expressed by breast tumor cells. Proteins that are breast tumor proteins react  
5 detectably within an immunoassay (such as an ELISA) with antisera from a patient with breast cancer. Polypeptides as described herein may be of any length. Additional sequences derived from the native protein and/or heterologous sequences may be present, and such sequences may (but need not) possess further immunogenic or antigenic properties.

10 An "immunogenic portion," as used herein is a portion of a protein that is recognized (*i.e.*, specifically bound) by a B-cell and/or T-cell surface antigen receptor. Such immunogenic portions generally comprise at least 5 amino acid residues, more preferably at least 10, and still more preferably at least 20 amino acid residues of a breast tumor protein or a variant thereof. Certain preferred immunogenic  
15 portions include peptides in which an N-terminal leader sequence and/or transmembrane domain have been deleted. Other preferred immunogenic portions may contain a small N- and/or C-terminal deletion (*e.g.*, 1-30 amino acids, preferably 5-15 amino acids), relative to the mature protein.

Immunogenic portions may generally be identified using well known  
20 techniques, such as those summarized in Paul, *Fundamental Immunology*, 3rd ed., 243-247 (Raven Press, 1993) and references cited therein. Such techniques include screening polypeptides for the ability to react with antigen-specific antibodies, antisera and/or T-cell lines or clones. As used herein, antisera and antibodies are "antigen-specific" if they specifically bind to an antigen (*i.e.*, they react with the protein in an  
25 ELISA or other immunoassay, and do not react detectably with unrelated proteins). Such antisera and antibodies may be prepared as described herein, and using well known techniques. An immunogenic portion of a native breast tumor protein is a portion that reacts with such antisera and/or T-cells at a level that is not substantially less than the reactivity of the full length polypeptide (*e.g.*, in an ELISA and/or T-cell  
30 reactivity assay). Such immunogenic portions may react within such assays at a level that is similar to or greater than the reactivity of the full length polypeptide. Such

5 screens may generally be performed using methods well known to those of ordinary skill in the art, such as those described in Harlow and Lane, *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory, 1988. For example, a polypeptide may be immobilized on a solid support and contacted with patient sera to allow binding of antibodies within the sera to the immobilized polypeptide. Unbound sera may then be removed and bound antibodies detected using, for example, <sup>125</sup>I-labeled Protein A.

As noted above, a composition may comprise a variant of a native breast tumor protein. A polypeptide "variant," as used herein, is a polypeptide that differs from a native breast tumor protein in one or more substitutions, deletions, additions and/or insertions, such that the immunogenicity of the polypeptide is not substantially diminished. In other words, the ability of a variant to react with antigen-specific antisera may be enhanced or unchanged, relative to the native protein, or may be diminished by less than 50%, and preferably less than 20%, relative to the native protein. Such variants may generally be identified by modifying one of the above polypeptide sequences and evaluating the reactivity of the modified polypeptide with antigen-specific antibodies or antisera as described herein. Preferred variants include those in which one or more portions, such as an N-terminal leader sequence or transmembrane domain, have been removed. Other preferred variants include variants in which a small portion (*e.g.*, 1-30 amino acids, preferably 5-15 amino acids) has been removed from the N- and/or C-terminal of the mature protein.

Polypeptide variants encompassed by the present invention include those exhibiting at least about 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% or more identity (determined as described above) to the polypeptides disclosed herein.

25 Preferably, a variant contains conservative substitutions. A "conservative substitution" is one in which an amino acid is substituted for another amino acid that has similar properties, such that one skilled in the art of peptide chemistry would expect the secondary structure and hydrophobic nature of the polypeptide to be substantially unchanged. Amino acid substitutions may generally be made on the basis of similarity in polarity, charge, solubility, hydrophobicity, hydrophilicity and/or the amphipathic nature of the residues. For example, negatively

- charged amino acids include aspartic acid and glutamic acid; positively charged amino acids include lysine and arginine; and amino acids with uncharged polar head groups having similar hydrophilicity values include leucine, isoleucine and valine; glycine and alanine; asparagine and glutamine; and serine, threonine, phenylalanine and tyrosine.
- 5 Other groups of amino acids that may represent conservative changes include: (1) ala, pro, gly, glu, asp, gln, asn, ser, thr; (2) cys, ser, tyr, thr; (3) val, ile, leu, met, ala, phe; (4) lys, arg, his; and (5) phe, tyr, trp, his. A variant may also, or alternatively, contain nonconservative changes. In a preferred embodiment, variant polypeptides differ from a native sequence by substitution, deletion or addition of five amino acids or fewer.
- 10 Variants may also (or alternatively) be modified by, for example, the deletion or addition of amino acids that have minimal influence on the immunogenicity, secondary structure and hydropathic nature of the polypeptide.

As noted above, polypeptides may comprise a signal (or leader) sequence at the N-terminal end of the protein, which co-translationally or post-translationally directs transfer of the protein. The polypeptide may also be conjugated to a linker or other sequence for ease of synthesis, purification or identification of the polypeptide (*e.g.*, poly-His), or to enhance binding of the polypeptide to a solid support. For example, a polypeptide may be conjugated to an immunoglobulin Fc region.

- Polypeptides may be prepared using any of a variety of well known techniques. Recombinant polypeptides encoded by DNA sequences as described above may be readily prepared from the DNA sequences using any of a variety of expression vectors known to those of ordinary skill in the art. Expression may be achieved in any appropriate host cell that has been transformed or transfected with an expression vector containing a DNA molecule that encodes a recombinant polypeptide. Suitable host cells include prokaryotes, yeast, and higher eukaryotic cells, such as mammalian cells and plant cells. Preferably, the host cells employed are *E. coli*, yeast or a mammalian cell line such as COS or CHO. Supernatants from suitable host/vector systems which secrete recombinant protein or polypeptide into culture media may be first concentrated using a commercially available filter. Following concentration, the concentrate may be applied to a suitable purification matrix such as an affinity matrix or an ion exchange

resin. Finally, one or more reverse phase HPLC steps can be employed to further purify a recombinant polypeptide.

Portions and other variants having less than about 100 amino acids, and generally less than about 50 amino acids, may also be generated by synthetic means, using techniques well known to those of ordinary skill in the art. For example, such polypeptides may be synthesized using any of the commercially available solid-phase techniques, such as the Merrifield solid-phase synthesis method, where amino acids are sequentially added to a growing amino acid chain. See Merrifield, *J. Am. Chem. Soc.* 85:2149-2146, 1963. Equipment for automated synthesis of polypeptides is commercially available from suppliers such as Perkin Elmer/Applied BioSystems Division (Foster City, CA), and may be operated according to the manufacturer's instructions.

Within certain specific embodiments, a polypeptide may be a fusion protein that comprises multiple polypeptides as described herein, or that comprises at least one polypeptide as described herein and an unrelated sequence, such as a known tumor protein. A fusion partner may, for example, assist in providing T helper epitopes (an immunological fusion partner), preferably T helper epitopes recognized by humans, or may assist in expressing the protein (an expression enhancer) at higher yields than the native recombinant protein. Certain preferred fusion partners are both immunological and expression enhancing fusion partners. Other fusion partners may be selected so as to increase the solubility of the protein or to enable the protein to be targeted to desired intracellular compartments. Still further fusion partners include affinity tags, which facilitate purification of the protein.

Fusion proteins may generally be prepared using standard techniques, including chemical conjugation. Preferably, a fusion protein is expressed as a recombinant protein, allowing the production of increased levels, relative to a non-fused protein, in an expression system. Briefly, DNA sequences encoding the polypeptide components may be assembled separately, and ligated into an appropriate expression vector. The 3' end of the DNA sequence encoding one polypeptide component is ligated, with or without a peptide linker, to the 5' end of a DNA sequence encoding the second polypeptide component so that the reading frames of the sequences are in phase.

This permits translation into a single fusion protein that retains the biological activity of both component polypeptides.

A peptide linker sequence may be employed to separate the first and second polypeptide components by a distance sufficient to ensure that each polypeptide  
5 folds into its secondary and tertiary structures. Such a peptide linker sequence is incorporated into the fusion protein using standard techniques well known in the art. Suitable peptide linker sequences may be chosen based on the following factors: (1) their ability to adopt a flexible extended conformation; (2) their inability to adopt a secondary structure that could interact with functional epitopes on the first and second  
10 polypeptides; and (3) the lack of hydrophobic or charged residues that might react with the polypeptide functional epitopes. Preferred peptide linker sequences contain Gly, Asn and Ser residues. Other near neutral amino acids, such as Thr and Ala may also be used in the linker sequence. Amino acid sequences which may be usefully employed as linkers include those disclosed in Maratea et al., *Gene* 40:39-46, 1985; Murphy et al.,  
15 *Proc. Natl. Acad. Sci. USA* 83:8258-8262, 1986; U.S. Patent No. 4,935,233 and U.S. Patent No. 4,751,180. The linker sequence may generally be from 1 to about 50 amino acids in length. Linker sequences are not required when the first and second polypeptides have non-essential N-terminal amino acid regions that can be used to separate the functional domains and prevent steric interference.

20 The ligated DNA sequences are operably linked to suitable transcriptional or translational regulatory elements. The regulatory elements responsible for expression of DNA are located only 5' to the DNA sequence encoding the first polypeptides. Similarly, stop codons required to end translation and transcription termination signals are only present 3' to the DNA sequence encoding the  
25 second polypeptide.

Fusion proteins are also provided. Such proteins comprise a polypeptide as described herein together with an unrelated immunogenic protein. Preferably the immunogenic protein is capable of eliciting a recall response. Examples of such proteins include tetanus, tuberculosis and hepatitis proteins (*see*, for example, Stoute  
30 et al. *New Engl. J. Med.*, 336:86-91, 1997).

Within preferred embodiments, an immunological fusion partner is derived from protein D, a surface protein of the gram-negative bacterium *Haemophilus influenza B* (WO 91/18926). Preferably, a protein D derivative comprises approximately the first third of the protein (e.g., the first N-terminal 100-110 amino acids), and a protein D derivative may be lipidated. Within certain preferred  
5       embodiments, the first 109 residues of a Lipoprotein D fusion partner is included on the N-terminus to provide the polypeptide with additional exogenous T-cell epitopes and to increase the expression level in *E. coli* (thus functioning as an expression enhancer). The lipid tail ensures optimal presentation of the antigen to antigen presenting cells.  
10       Other fusion partners include the non-structural protein from influenzae virus, NS1 (hemagglutinin). Typically, the N-terminal 81 amino acids are used, although different fragments that include T-helper epitopes may be used.

In another embodiment, the immunological fusion partner is the protein known as LYTA, or a portion thereof (preferably a C-terminal portion). LYTA is  
15       derived from *Streptococcus pneumoniae*, which synthesizes an N-acetyl-L-alanine amidase known as amidase LYTA (encoded by the *LytA* gene; *Gene* 43:265-292, 1986). LYTA is an autolysin that specifically degrades certain bonds in the peptidoglycan backbone. The C-terminal domain of the LYTA protein is responsible for the affinity to the choline or to some choline analogues such as DEAE. This  
20       property has been exploited for the development of *E. coli* C-LYTA expressing plasmids useful for expression of fusion proteins. Purification of hybrid proteins containing the C-LYTA fragment at the amino terminus has been described (see *Biotechnology* 10:795-798, 1992). Within a preferred embodiment, a repeat portion of LYTA may be incorporated into a fusion protein. A repeat portion is found in the C-  
25       terminal region starting at residue 178. A particularly preferred repeat portion incorporates residues 188-305.

In general, polypeptides (including fusion proteins) and polynucleotides as described herein are isolated. An "isolated" polypeptide or polynucleotide is one that is removed from its original environment. For example, a naturally-occurring protein is  
30       isolated if it is separated from some or all of the coexisting materials in the natural system. Preferably, such polypeptides are at least about 90% pure, more preferably at

least about 95% pure and most preferably at least about 99% pure. A polynucleotide is considered to be isolated if, for example, it is cloned into a vector that is not a part of the natural environment.

#### **BINDING AGENTS**

5                   The present invention further provides agents, such as antibodies and antigen-binding fragments thereof, that specifically bind to a breast tumor protein. As used herein, an antibody, or antigen-binding fragment thereof, is said to "specifically bind" to a breast tumor protein if it reacts at a detectable level (within, for example, an ELISA) with a breast tumor protein, and does not react detectably with unrelated  
10 proteins under similar conditions. As used herein, "binding" refers to a noncovalent association between two separate molecules such that a complex is formed. The ability to bind may be evaluated by, for example, determining a binding constant for the formation of the complex. The binding constant is the value obtained when the concentration of the complex is divided by the product of the component  
15 concentrations. In general, two compounds are said to "bind," in the context of the present invention, when the binding constant for complex formation exceeds about  $10^3$  L/mol. The binding constant may be determined using methods well known in the art.

                  Binding agents may be further capable of differentiating between patients with and without a cancer, such as breast cancer, using the representative assays  
20 provided herein. In other words, antibodies or other binding agents that bind to a breast tumor protein will generate a signal indicating the presence of a cancer in at least about 20% of patients with the disease, and will generate a negative signal indicating the absence of the disease in at least about 90% of individuals without the cancer. To determine whether a binding agent satisfies this requirement, biological samples (*e.g.*,  
25 blood, sera, sputum, urine and/or tumor biopsies) from patients with and without a cancer (as determined using standard clinical tests) may be assayed as described herein for the presence of polypeptides that bind to the binding agent. It will be apparent that a statistically significant number of samples with and without the disease should be assayed. Each binding agent should satisfy the above criteria; however, those of



ordinary skill in the art will recognize that binding agents may be used in combination to improve sensitivity.

Any agent that satisfies the above requirements may be a binding agent. For example, a binding agent may be a ribosome, with or without a peptide component, an RNA molecule or a polypeptide. In a preferred embodiment, a binding agent is an antibody or an antigen-binding fragment thereof. Antibodies may be prepared by any of a variety of techniques known to those of ordinary skill in the art. *See, e.g.,* Harlow and Lane, *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory, 1988. In general, antibodies can be produced by cell culture techniques, including the generation of monoclonal antibodies as described herein, or via transfection of antibody genes into suitable bacterial or mammalian cell hosts, in order to allow for the production of recombinant antibodies. In one technique, an immunogen comprising the polypeptide is initially injected into any of a wide variety of mammals (*e.g.,* mice, rats, rabbits, sheep or goats). In this step, the polypeptides of this invention may serve as the immunogen without modification. Alternatively, particularly for relatively short polypeptides, a superior immune response may be elicited if the polypeptide is joined to a carrier protein, such as bovine serum albumin or keyhole limpet hemocyanin. The immunogen is injected into the animal host, preferably according to a predetermined schedule incorporating one or more booster immunizations, and the animals are bled periodically. Polyclonal antibodies specific for the polypeptide may then be purified from such antisera by, for example, affinity chromatography using the polypeptide coupled to a suitable solid support.

Monoclonal antibodies specific for an antigenic polypeptide of interest may be prepared, for example, using the technique of Kohler and Milstein, *Eur. J. Immunol.* 6:511-519, 1976, and improvements thereto. Briefly, these methods involve the preparation of immortal cell lines capable of producing antibodies having the desired specificity (*i.e.,* reactivity with the polypeptide of interest). Such cell lines may be produced, for example, from spleen cells obtained from an animal immunized as described above. The spleen cells are then immortalized by, for example, fusion with a myeloma cell fusion partner, preferably one that is syngeneic with the immunized animal. A variety of fusion techniques may be employed. For example, the spleen cells

and myeloma cells may be combined with a nonionic detergent for a few minutes and then plated at low density on a selective medium that supports the growth of hybrid cells, but not myeloma cells. A preferred selection technique uses HAT (hypoxanthine, aminopterin, thymidine) selection. After a sufficient time, usually about 1 to 2 weeks,  
5 colonies of hybrids are observed. Single colonies are selected and their culture supernatants tested for binding activity against the polypeptide. Hybridomas having high reactivity and specificity are preferred.

Monoclonal antibodies may be isolated from the supernatants of growing hybridoma colonies. In addition, various techniques may be employed to enhance the  
10 yield, such as injection of the hybridoma cell line into the peritoneal cavity of a suitable vertebrate host, such as a mouse. Monoclonal antibodies may then be harvested from the ascites fluid or the blood. Contaminants may be removed from the antibodies by conventional techniques, such as chromatography, gel filtration, precipitation, and extraction. The polypeptides of this invention may be used in the purification process  
15 in, for example, an affinity chromatography step.

Within certain embodiments, the use of antigen-binding fragments of antibodies may be preferred. Such fragments include Fab fragments, which may be prepared using standard techniques. Briefly, immunoglobulins may be purified from rabbit serum by affinity chromatography on Protein A bead columns (Harlow and Lane,  
20 *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory, 1988) and digested by papain to yield Fab and Fc fragments. The Fab and Fc fragments may be separated by affinity chromatography on protein A bead columns.

Monoclonal antibodies of the present invention may be coupled to one or more therapeutic agents. Suitable agents in this regard include radionuclides,  
25 differentiation inducers, drugs, toxins, and derivatives thereof. Preferred radionuclides include  $^{90}\text{Y}$ ,  $^{123}\text{I}$ ,  $^{125}\text{I}$ ,  $^{131}\text{I}$ ,  $^{186}\text{Re}$ ,  $^{188}\text{Re}$ ,  $^{211}\text{At}$ , and  $^{212}\text{Bi}$ . Preferred drugs include methotrexate, and pyrimidine and purine analogs. Preferred differentiation inducers include phorbol esters and butyric acid. Preferred toxins include ricin, abrin, diphtheria toxin, cholera toxin, gelonin, *Pseudomonas* exotoxin, *Shigella* toxin, and pokeweed  
30 antiviral protein.

A therapeutic agent may be coupled (*e.g.*, covalently bonded) to a suitable monoclonal antibody either directly or indirectly (*e.g.*, via a linker group). A direct reaction between an agent and an antibody is possible when each possesses a substituent capable of reacting with the other. For example, a nucleophilic group, such as an amino or sulfhydryl group, on one may be capable of reacting with a carbonyl-containing group, such as an anhydride or an acid halide, or with an alkyl group containing a good leaving group (*e.g.*, a halide) on the other.

Alternatively, it may be desirable to couple a therapeutic agent and an antibody via a linker group. A linker group can function as a spacer to distance an antibody from an agent in order to avoid interference with binding capabilities. A linker group can also serve to increase the chemical reactivity of a substituent on an agent or an antibody, and thus increase the coupling efficiency. An increase in chemical reactivity may also facilitate the use of agents, or functional groups on agents, which otherwise would not be possible.

It will be evident to those skilled in the art that a variety of bifunctional or polyfunctional reagents, both homo- and hetero-functional (such as those described in the catalog of the Pierce Chemical Co., Rockford, IL), may be employed as the linker group. Coupling may be effected, for example, through amino groups, carboxyl groups, sulfhydryl groups or oxidized carbohydrate residues. There are numerous references describing such methodology, *e.g.*, U.S. Patent No. 4,671,958, to Rodwell et al.

Where a therapeutic agent is more potent when free from the antibody portion of the immunoconjugates of the present invention, it may be desirable to use a linker group which is cleavable during or upon internalization into a cell. A number of different cleavable linker groups have been described. The mechanisms for the intracellular release of an agent from these linker groups include cleavage by reduction of a disulfide bond (*e.g.*, U.S. Patent No. 4,489,710, to Spitler), by irradiation of a photolabile bond (*e.g.*, U.S. Patent No. 4,625,014, to Senter et al.), by hydrolysis of derivatized amino acid side chains (*e.g.*, U.S. Patent No. 4,638,045, to Kohn et al.), by serum complement-mediated hydrolysis (*e.g.*, U.S. Patent No. 4,671,958, to Rodwell et al.), and acid-catalyzed hydrolysis (*e.g.*, U.S. Patent No. 4,569,789, to Blattler et al.).

It may be desirable to couple more than one agent to an antibody. In one embodiment, multiple molecules of an agent are coupled to one antibody molecule. In another embodiment, more than one type of agent may be coupled to one antibody. Regardless of the particular embodiment, immunoconjugates with more than one agent  
5 may be prepared in a variety of ways. For example, more than one agent may be coupled directly to an antibody molecule, or linkers that provide multiple sites for attachment can be used. Alternatively, a carrier can be used.

A carrier may bear the agents in a variety of ways, including covalent bonding either directly or via a linker group. Suitable carriers include proteins such as  
10 albumins (*e.g.*, U.S. Patent No. 4,507,234, to Kato et al.), peptides and polysaccharides such as aminodextran (*e.g.*, U.S. Patent No. 4,699,784, to Shih et al.). A carrier may also bear an agent by noncovalent bonding or by encapsulation, such as within a liposome vesicle (*e.g.*, U.S. Patent Nos. 4,429,008 and 4,873,088). Carriers specific for radionuclide agents include radiohalogenated small molecules and chelating  
15 compounds. For example, U.S. Patent No. 4,735,792 discloses representative radiohalogenated small molecules and their synthesis. A radionuclide chelate may be formed from chelating compounds that include those containing nitrogen and sulfur atoms as the donor atoms for binding the metal, or metal oxide, radionuclide. For example, U.S. Patent No. 4,673,562, to Davison et al. discloses representative chelating  
20 compounds and their synthesis.

A variety of routes of administration for the antibodies and immunoconjugates may be used. Typically, administration will be intravenous, intramuscular, subcutaneous or in the bed of a resected tumor. It will be evident that the precise dose of the antibody/immunoconjugate will vary depending upon the antibody  
25 used, the antigen density on the tumor, and the rate of clearance of the antibody.

## T CELLS

Immunotherapeutic compositions may also, or alternatively, comprise T cells specific for a breast tumor protein. Such cells may generally be prepared *in vitro* or *ex vivo*, using standard procedures. For example, T cells may be isolated from bone  
30 marrow, peripheral blood, or a fraction of bone marrow or peripheral blood of a patient,

using a commercially available cell separation system, such as the Isolex™ System, available from Nexell Therapeutics, Inc. (Irvine, CA; see also U.S. Patent No. 5,240,856; U.S. Patent No. 5,215,926; WO 89/06280; WO 91/16116 and WO 92/07243). Alternatively, T cells may be derived from related or unrelated humans,  
5 non-human mammals, cell lines or cultures.

T cells may be stimulated with a breast tumor polypeptide, polynucleotide encoding a breast tumor polypeptide and/or an antigen presenting cell (APC) that expresses such a polypeptide. Such stimulation is performed under conditions and for a time sufficient to permit the generation of T cells that are specific  
10 for the polypeptide. Preferably, a breast tumor polypeptide or polynucleotide is present within a delivery vehicle, such as a microsphere, to facilitate the generation of specific T cells.

T cells are considered to be specific for a breast tumor polypeptide if the T cells specifically proliferate, secrete cytokines or kill target cells coated with the polypeptide or expressing a gene encoding the polypeptide. T cell specificity may be  
15 evaluated using any of a variety of standard techniques. For example, within a chromium release assay or proliferation assay, a stimulation index of more than two fold increase in lysis and/or proliferation, compared to negative controls, indicates T cell specificity. Such assays may be performed, for example, as described in Chen et al., *Cancer Res.* 54:1065-1070, 1994. Alternatively, detection of the proliferation of  
20 T cells may be accomplished by a variety of known techniques. For example, T cell proliferation can be detected by measuring an increased rate of DNA synthesis (e.g., by pulse-labeling cultures of T cells with tritiated thymidine and measuring the amount of tritiated thymidine incorporated into DNA). Contact with a breast tumor polypeptide  
25 (100 ng/ml - 100 µg/ml, preferably 200 ng/ml - 25 µg/ml) for 3 - 7 days should result in at least a two fold increase in proliferation of the T cells. Contact as described above for 2-3 hours should result in activation of the T cells, as measured using standard cytokine assays in which a two fold increase in the level of cytokine release (e.g., TNF or IFN-γ) is indicative of T cell activation (see Coligan et al., *Current Protocols in Immunology*, vol. 1, Wiley Interscience (Greene 1998)). T cells that have been  
30 activated in response to a breast tumor polypeptide, polynucleotide or polypeptide-

expressing APC may be CD4<sup>+</sup> and/or CD8<sup>+</sup>. Breast tumor protein-specific T cells may be expanded using standard techniques. Within preferred embodiments, the T cells are derived from a patient, a related donor or an unrelated donor, and are administered to the patient following stimulation and expansion.

5                   For therapeutic purposes, CD4<sup>+</sup> or CD8<sup>+</sup> T cells that proliferate in response to a breast tumor polypeptide, polynucleotide or APC can be expanded in number either *in vitro* or *in vivo*. Proliferation of such T cells *in vitro* may be accomplished in a variety of ways. For example, the T cells can be re-exposed to a breast tumor polypeptide, or a short peptide corresponding to an immunogenic portion  
10 of such a polypeptide, with or without the addition of T cell growth factors, such as interleukin-2, and/or stimulator cells that synthesize a breast tumor polypeptide. Alternatively, one or more T cells that proliferate in the presence of a breast tumor protein can be expanded in number by cloning. Methods for cloning cells are well known in the art, and include limiting dilution.

#### 15   PHARMACEUTICAL COMPOSITIONS

In additional embodiments, the present invention concerns formulation of one or more of the polynucleotide, polypeptide, T-cell and/or antibody compositions disclosed herein in pharmaceutically-acceptable solutions for administration to a cell or an animal, either alone, or in combination with one or more other modalities of therapy.

20                   It will also be understood that, if desired, the nucleic acid segment, RNA, DNA or PNA compositions that express a polypeptide as disclosed herein may be administered in combination with other agents as well, such as, *e.g.*, other proteins or polypeptides or various pharmaceutically-active agents. In fact, there is virtually no limit to other components that may also be included, given that the additional agents do  
25 not cause a significant adverse effect upon contact with the target cells or host tissues. The compositions may thus be delivered along with various other agents as required in the particular instance. Such compositions may be purified from host cells or other biological sources, or alternatively may be chemically synthesized as described herein. Likewise, such compositions may further comprise substituted or derivatized RNA or  
30 DNA compositions.

Formulation of pharmaceutically-acceptable excipients and carrier solutions is well-known to those of skill in the art, as is the development of suitable dosing and treatment regimens for using the particular compositions described herein in a variety of treatment regimens, including *e.g.*, oral, parenteral, intravenous, intranasal, and intramuscular administration and formulation.

#### 1. ORAL DELIVERY

In certain applications, the pharmaceutical compositions disclosed herein may be delivered *via* oral administration to an animal. As such, these compositions may be formulated with an inert diluent or with an assimilable edible carrier, or they may be enclosed in hard- or soft-shell gelatin capsule, or they may be compressed into tablets, or they may be incorporated directly with the food of the diet.

The active compounds may even be incorporated with excipients and used in the form of ingestible tablets, buccal tables, troches, capsules, elixirs, suspensions, syrups, wafers, and the like (Mathiowitz *et al.*, 1997; Hwang *et al.*, 1998; U. S. Patent 5,641,515; U. S. Patent 5,580,579 and U. S. Patent 5,792,451, each specifically incorporated herein by reference in its entirety). The tablets, troches, pills, capsules and the like may also contain the following: a binder, as gum tragacanth, acacia, cornstarch, or gelatin; excipients, such as dicalcium phosphate; a disintegrating agent, such as corn starch, potato starch, alginic acid and the like; a lubricant, such as magnesium stearate; and a sweetening agent, such as sucrose, lactose or saccharin may be added or a flavoring agent, such as peppermint, oil of wintergreen, or cherry flavoring. When the dosage unit form is a capsule, it may contain, in addition to materials of the above type, a liquid carrier. Various other materials may be present as coatings or to otherwise modify the physical form of the dosage unit. For instance, tablets, pills, or capsules may be coated with shellac, sugar, or both. A syrup or elixir may contain the active compound sucrose as a sweetening agent methyl and propylparabens as preservatives, a dye and flavoring, such as cherry or orange flavor. Of course, any material used in preparing any dosage unit form should be pharmaceutically pure and substantially non-toxic in the amounts employed. In

addition, the active compounds may be incorporated into sustained-release preparation and formulations.

Typically, these formulations may contain at least about 0.1% of the active compound or more, although the percentage of the active ingredient(s) may, of course, be varied and may conveniently be between about 1 or 2% and about 60% or 70% or more of the weight or volume of the total formulation. Naturally, the amount of active compound(s) in each therapeutically useful composition may be prepared in such a way that a suitable dosage will be obtained in any given unit dose of the compound. Factors such as solubility, bioavailability, biological half-life, route of administration, product shelf life, as well as other pharmacological considerations will be contemplated by one skilled in the art of preparing such pharmaceutical formulations, and as such, a variety of dosages and treatment regimens may be desirable.

For oral administration the compositions of the present invention may alternatively be incorporated with one or more excipients in the form of a mouthwash, dentifrice, buccal tablet, oral spray, or sublingual orally-administered formulation. For example, a mouthwash may be prepared incorporating the active ingredient in the required amount in an appropriate solvent, such as a sodium borate solution (Dobell's Solution). Alternatively, the active ingredient may be incorporated into an oral solution such as one containing sodium borate, glycerin and potassium bicarbonate, or dispersed in a dentifrice, or added in a therapeutically-effective amount to a composition that may include water, binders, abrasives, flavoring agents, foaming agents, and humectants. Alternatively the compositions may be fashioned into a tablet or solution form that may be placed under the tongue or otherwise dissolved in the mouth.

## 2. INJECTABLE DELIVERY

In certain circumstances it will be desirable to deliver the pharmaceutical compositions disclosed herein parenterally, intravenously, intramuscularly, or even intraperitoneally as described in U. S. Patent 5,543,158; U. S. Patent 5,641,515 and U. S. Patent 5,399,363 (each specifically incorporated herein by reference in its entirety). Solutions of the active compounds as free base or pharmacologically acceptable salts may be prepared in water suitably mixed with a surfactant, such as



hydroxypropylcellulose. Dispersions may also be prepared in glycerol, liquid polyethylene glycols, and mixtures thereof and in oils. Under ordinary conditions of storage and use, these preparations contain a preservative to prevent the growth of microorganisms.

5                   The pharmaceutical forms suitable for injectable use include sterile aqueous solutions or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersions (U. S. Patent 5,466,468, specifically incorporated herein by reference in its entirety). In all cases the form must be sterile and must be fluid to the extent that easy syringability exists. It must be stable  
10 under the conditions of manufacture and storage and must be preserved against the contaminating action of microorganisms, such as bacteria and fungi. The carrier can be a solvent or dispersion medium containing, for example, water, ethanol, polyol (*e.g.*, glycerol, propylene glycol, and liquid polyethylene glycol, and the like), suitable mixtures thereof, and/or vegetable oils. Proper fluidity may be maintained, for  
15 example, by the use of a coating, such as lecithin, by the maintenance of the required particle size in the case of dispersion and by the use of surfactants. The prevention of the action of microorganisms can be facilitated by various antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, sorbic acid, thimerosal, and the like. In many cases, it will be preferable to include isotonic agents, for example, sugars  
20 or sodium chloride. Prolonged absorption of the injectable compositions can be brought about by the use in the compositions of agents delaying absorption, for example, aluminum monostearate and gelatin.

For parenteral administration in an aqueous solution, for example, the solution should be suitably buffered if necessary and the liquid diluent first rendered  
25 isotonic with sufficient saline or glucose. These particular aqueous solutions are especially suitable for intravenous, intramuscular, subcutaneous and intraperitoneal administration. In this connection, a sterile aqueous medium that can be employed will be known to those of skill in the art in light of the present disclosure. For example, one dosage may be dissolved in 1 ml of isotonic NaCl solution and either added to 1000 ml  
30 of hypodermoclysis fluid or injected at the proposed site of infusion, (see for example, "Remington's Pharmaceutical Sciences" 15th Edition, pages 1035-1038 and 1570-

1580). Some variation in dosage will necessarily occur depending on the condition of the subject being treated. The person responsible for administration will, in any event, determine the appropriate dose for the individual subject. Moreover, for human administration, preparations should meet sterility, pyrogenicity, and the general safety  
5 and purity standards as required by FDA Office of Biologics standards.

Sterile injectable solutions are prepared by incorporating the active compounds in the required amount in the appropriate solvent with various of the other ingredients enumerated above, as required, followed by filtered sterilization. Generally, dispersions are prepared by incorporating the various sterilized active ingredients into a  
10 sterile vehicle which contains the basic dispersion medium and the required other ingredients from those enumerated above. In the case of sterile powders for the preparation of sterile injectable solutions, the preferred methods of preparation are vacuum-drying and freeze-drying techniques which yield a powder of the active ingredient plus any additional desired ingredient from a previously sterile-filtered  
15 solution thereof.

The compositions disclosed herein may be formulated in a neutral or salt form. Pharmaceutically-acceptable salts, include the acid addition salts (formed with the free amino groups of the protein) and which are formed with inorganic acids such as, for example, hydrochloric or phosphoric acids, or such organic acids as acetic,  
20 oxalic, tartaric, mandelic, and the like. Salts formed with the free carboxyl groups can also be derived from inorganic bases such as, for example, sodium, potassium, ammonium, calcium, or ferric hydroxides, and such organic bases as isopropylamine, trimethylamine, histidine, procaine and the like. Upon formulation, solutions will be administered in a manner compatible with the dosage formulation and in such amount  
25 as is therapeutically effective. The formulations are easily administered in a variety of dosage forms such as injectable solutions, drug-release capsules, and the like.

As used herein, "carrier" includes any and all solvents, dispersion media, vehicles, coatings, diluents, antibacterial and antifungal agents, isotonic and absorption delaying agents, buffers, carrier solutions, suspensions, colloids, and the like. The use  
30 of such media and agents for pharmaceutical active substances is well known in the art. Except insofar as any conventional media or agent is incompatible with the active

ingredient, its use in the therapeutic compositions is contemplated. Supplementary active ingredients can also be incorporated into the compositions.

The phrase "pharmaceutically-acceptable" refers to molecular entities and compositions that do not produce an allergic or similar untoward reaction when  
5 administered to a human. The preparation of an aqueous composition that contains a protein as an active ingredient is well understood in the art. Typically, such compositions are prepared as injectables, either as liquid solutions or suspensions; solid forms suitable for solution in, or suspension in, liquid prior to injection can also be prepared. The preparation can also be emulsified.

### 10 3. NASAL DELIVERY

In certain embodiments, the pharmaceutical compositions may be delivered by intranasal sprays, inhalation, and/or other aerosol delivery vehicles. Methods for delivering genes, nucleic acids, and peptide compositions directly to the lungs *via* nasal aerosol sprays has been described *e.g.*, in U. S. Patent 5,756,353 and U.  
15 S. Patent 5,804,212 (each specifically incorporated herein by reference in its entirety). Likewise, the delivery of drugs using intranasal microparticle resins (Takenaga *et al.*, 1998) and lysophosphatidyl-glycerol compounds (U. S. Patent 5,725,871, specifically incorporated herein by reference in its entirety) are also well-known in the pharmaceutical arts. Likewise, transmucosal drug delivery in the form of a  
20 polytetrafluoroethylene support matrix is described in U. S. Patent 5,780,045 (specifically incorporated herein by reference in its entirety).

### 4. LIPOSOME-, NANOCAPSULE-, AND MICROPARTICLE-MEDIATED DELIVERY

In certain embodiments, the inventors contemplate the use of liposomes, nanocapsules, microparticles, microspheres, lipid particles, vesicles, and the like, for the  
25 introduction of the compositions of the present invention into suitable host cells. In particular, the compositions of the present invention may be formulated for delivery either encapsulated in a lipid particle, a liposome, a vesicle, a nanosphere, or a nanoparticle or the like.

Such formulations may be preferred for the introduction of pharmaceutically-acceptable formulations of the nucleic acids or constructs disclosed herein. The formation and use of liposomes is generally known to those of skill in the art (see for example, Couvreur *et al.*, 1977; Couvreur, 1988; Lasic, 1998; which  
5 describes the use of liposomes and nanocapsules in the targeted antibiotic therapy for intracellular bacterial infections and diseases). Recently, liposomes were developed with improved serum stability and circulation half-times (Gabizon and Papahadjopoulos, 1988; Allen and Choun, 1987; U. S. Patent 5,741,516, specifically incorporated herein by reference in its entirety). Further, various methods of liposome  
10 and liposome like preparations as potential drug carriers have been reviewed (Takakura, 1998; Chandran *et al.*, 1997; Margalit, 1995; U. S. Patent 5,567,434; U. S. Patent 5,552,157; U. S. Patent 5,565,213; U. S. Patent 5,738,868 and U. S. Patent 5,795,587, each specifically incorporated herein by reference in its entirety).

Liposomes have been used successfully with a number of cell types that  
15 are normally resistant to transfection by other procedures including T cell suspensions, primary hepatocyte cultures and PC 12 cells (Renneisen *et al.*, 1990; Muller *et al.*, 1990). In addition, liposomes are free of the DNA length constraints that are typical of viral-based delivery systems. Liposomes have been used effectively to introduce genes, drugs (Heath and Martin, 1986; Heath *et al.*, 1986; Balazsovits *et al.*, 1989; Fresta and  
20 Puglisi, 1996), radiotherapeutic agents (Pikul *et al.*, 1987), enzymes (Imaizumi *et al.*, 1990a; Imaizumi *et al.*, 1990b), viruses (Faller and Baltimore, 1984), transcription factors and allosteric effectors (Nicolau and Gersonde, 1979) into a variety of cultured cell lines and animals. In addition, several successful clinical trials examining the effectiveness of liposome-mediated drug delivery have been completed (Lopez-  
25 Berestein *et al.*, 1985a; 1985b; Coune, 1988; Sculier *et al.*, 1988). Furthermore, several studies suggest that the use of liposomes is not associated with autoimmune responses, toxicity or gonadal localization after systemic delivery (Mori and Fukatsu, 1992).

Liposomes are formed from phospholipids that are dispersed in an aqueous medium and spontaneously form multilamellar concentric bilayer vesicles (also  
30 termed multilamellar vesicles (MLVs). MLVs generally have diameters of from 25 nm to 4  $\mu$ m. Sonication of MLVs results in the formation of small unilamellar vesicles

(SUVs) with diameters in the range of 200 to 500 Å, containing an aqueous solution in the core.

Liposomes bear resemblance to cellular membranes and are contemplated for use in connection with the present invention as carriers for the peptide compositions. They are widely suitable as both water- and lipid-soluble substances can be entrapped, *i.e.* in the aqueous spaces and within the bilayer itself, respectively. It is possible that the drug-bearing liposomes may even be employed for site-specific delivery of active agents by selectively modifying the liposomal formulation.

In addition to the teachings of Couvreur *et al.* (1977; 1988), the following information may be utilized in generating liposomal formulations. Phospholipids can form a variety of structures other than liposomes when dispersed in water, depending on the molar ratio of lipid to water. At low ratios the liposome is the preferred structure. The physical characteristics of liposomes depend on pH, ionic strength and the presence of divalent cations. Liposomes can show low permeability to ionic and polar substances, but at elevated temperatures undergo a phase transition which markedly alters their permeability. The phase transition involves a change from a closely packed, ordered structure, known as the gel state, to a loosely packed, less-ordered structure, known as the fluid state. This occurs at a characteristic phase-transition temperature and results in an increase in permeability to ions, sugars and drugs.

In addition to temperature, exposure to proteins can alter the permeability of liposomes. Certain soluble proteins, such as cytochrome c, bind, deform and penetrate the bilayer, thereby causing changes in permeability. Cholesterol inhibits this penetration of proteins, apparently by packing the phospholipids more tightly. It is contemplated that the most useful liposome formations for antibiotic and inhibitor delivery will contain cholesterol.

The ability to trap solutes varies between different types of liposomes. For example, MLVs are moderately efficient at trapping solutes, but SUVs are extremely inefficient. SUVs offer the advantage of homogeneity and reproducibility in size distribution, however, and a compromise between size and trapping efficiency is

offered by large unilamellar vesicles (LUVs). These are prepared by ether evaporation and are three to four times more efficient at solute entrapment than MLVs.

In addition to liposome characteristics, an important determinant in entrapping compounds is the physicochemical properties of the compound itself. Polar compounds are trapped in the aqueous spaces and nonpolar compounds bind to the lipid bilayer of the vesicle. Polar compounds are released through permeation or when the bilayer is broken, but nonpolar compounds remain affiliated with the bilayer unless it is disrupted by temperature or exposure to lipoproteins. Both types show maximum efflux rates at the phase transition temperature.

Liposomes interact with cells *via* four different mechanisms: endocytosis by phagocytic cells of the reticuloendothelial system such as macrophages and neutrophils; adsorption to the cell surface, either by nonspecific weak hydrophobic or electrostatic forces, or by specific interactions with cell-surface components; fusion with the plasma cell membrane by insertion of the lipid bilayer of the liposome into the plasma membrane, with simultaneous release of liposomal contents into the cytoplasm; and by transfer of liposomal lipids to cellular or subcellular membranes, or vice versa, without any association of the liposome contents. It often is difficult to determine which mechanism is operative and more than one may operate at the same time.

The fate and disposition of intravenously injected liposomes depend on their physical properties, such as size, fluidity, and surface charge. They may persist in tissues for h or days, depending on their composition, and half lives in the blood range from min to several h. Larger liposomes, such as MLVs and LUVs, are taken up rapidly by phagocytic cells of the reticuloendothelial system, but physiology of the circulatory system restrains the exit of such large species at most sites. They can exit only in places where large openings or pores exist in the capillary endothelium, such as the sinusoids of the liver or spleen. Thus, these organs are the predominate site of uptake. On the other hand, SUVs show a broader tissue distribution but still are sequestered highly in the liver and spleen. In general, this *in vivo* behavior limits the potential targeting of liposomes to only those organs and tissues accessible to their large size. These include the blood, liver, spleen, bone marrow, and lymphoid organs.

Targeting is generally not a limitation in terms of the present invention. However, should specific targeting be desired, methods are available for this to be accomplished. Antibodies may be used to bind to the liposome surface and to direct the antibody and its drug contents to specific antigenic receptors located on a particular cell-type surface. Carbohydrate determinants (glycoprotein or glycolipid cell-surface components that play a role in cell-cell recognition, interaction and adhesion) may also be used as recognition sites as they have potential in directing liposomes to particular cell types. Mostly, it is contemplated that intravenous injection of liposomal preparations would be used, but other routes of administration are also conceivable.

Alternatively, the invention provides for pharmaceutically-acceptable nanocapsule formulations of the compositions of the present invention. Nanocapsules can generally entrap compounds in a stable and reproducible way (Henry-Michelland *et al.*, 1987; Quintanar-Guerrero *et al.*, 1998; Douglas *et al.*, 1987). To avoid side effects due to intracellular polymeric overloading, such ultrafine particles (sized around 0.1  $\mu\text{m}$ ) should be designed using polymers able to be degraded *in vivo*. Biodegradable polyalkyl-cyanoacrylate nanoparticles that meet these requirements are contemplated for use in the present invention. Such particles may be easily made, as described (Couvreur *et al.*, 1980; 1988; zur Muhlen *et al.*, 1998; Zambaux *et al.* 1998; Pinto-Alphandry *et al.*, 1995 and U. S. Patent 5,145,684, specifically incorporated herein by reference in its entirety).

## VACCINES

In certain preferred embodiments of the present invention, vaccines are provided. The vaccines will generally comprise one or more pharmaceutical compositions, such as those discussed above, in combination with an immunostimulant. An immunostimulant may be any substance that enhances or potentiates an immune response (antibody and/or cell-mediated) to an exogenous antigen. Examples of immunostimulants include adjuvants, biodegradable microspheres (*e.g.*, polylactic galactide) and liposomes (into which the compound is incorporated; *see e.g.*, Fullerton, U.S. Patent No. 4,235,877). Vaccine preparation is generally described in, for example, M.F. Powell and M.J. Newman, eds., "Vaccine Design (the subunit and adjuvant

approach)," Plenum Press (NY, 1995). Pharmaceutical compositions and vaccines within the scope of the present invention may also contain other compounds, which may be biologically active or inactive. For example, one or more immunogenic portions of other tumor antigens may be present, either incorporated into a fusion  
5 polypeptide or as a separate compound, within the composition or vaccine.

Illustrative vaccines may contain DNA encoding one or more of the polypeptides as described above, such that the polypeptide is generated *in situ*. As noted above, the DNA may be present within any of a variety of delivery systems known to those of ordinary skill in the art, including nucleic acid expression systems,  
10 bacteria and viral expression systems. Numerous gene delivery techniques are well known in the art, such as those described by Rolland, *Crit. Rev. Therap. Drug Carrier Systems* 15:143-198, 1998, and references cited therein. Appropriate nucleic acid expression systems contain the necessary DNA sequences for expression in the patient (such as a suitable promoter and terminating signal). Bacterial delivery systems involve  
15 the administration of a bacterium (such as *Bacillus-Calmette-Guerrin*) that expresses an immunogenic portion of the polypeptide on its cell surface or secretes such an epitope. In a preferred embodiment, the DNA may be introduced using a viral expression system (*e.g.*, vaccinia or other pox virus, retrovirus, or adenovirus), which may involve the use of a non-pathogenic (defective), replication competent virus. Suitable systems are  
20 disclosed, for example, in Fisher-Hoch et al., *Proc. Natl. Acad. Sci. USA* 86:317-321, 1989; Flexner et al., *Ann. N.Y. Acad. Sci.* 569:86-103, 1989; Flexner et al., *Vaccine* 8:17-21, 1990; U.S. Patent Nos. 4,603,112, 4,769,330, and 5,017,487; WO 89/01973; U.S. Patent No. 4,777,127; GB 2,200,651; EP 0,345,242; WO 91/02805; Berkner, *Biotechniques* 6:616-627, 1988; Rosenfeld et al., *Science* 252:431-434, 1991; Kolls et al., *Proc. Natl. Acad. Sci. USA* 91:215-219, 1994; Kass-Eisler et al., *Proc. Natl. Acad. Sci. USA* 90:11498-11502, 1993; Guzman et al., *Circulation* 88:2838-2848, 1993; and Guzman et al., *Cir. Res.* 73:1202-1207, 1993. Techniques for incorporating DNA into such expression systems are well known to those of ordinary skill in the art. The DNA may also be "naked," as described, for example, in Ulmer et al., *Science* 259:1745-1749,  
25 1993 and reviewed by Cohen, *Science* 259:1691-1692, 1993. The uptake of naked DNA may be increased by coating the DNA onto biodegradable beads, which are



efficiently transported into the cells. It will be apparent that a vaccine may comprise both a polynucleotide and a polypeptide component. Such vaccines may provide for an enhanced immune response.

It will be apparent that a vaccine may contain pharmaceutically acceptable salts of the polynucleotides and polypeptides provided herein. Such salts may be prepared from pharmaceutically acceptable non-toxic bases, including organic bases (*e.g.*, salts of primary, secondary and tertiary amines and basic amino acids) and inorganic bases (*e.g.*, sodium, potassium, lithium, ammonium, calcium and magnesium salts).

While any suitable carrier known to those of ordinary skill in the art may be employed in the vaccine compositions of this invention, the type of carrier will vary depending on the mode of administration. Compositions of the present invention may be formulated for any appropriate manner of administration, including for example, topical, oral, nasal, intravenous, intracranial, intraperitoneal, subcutaneous or intramuscular administration. For parenteral administration, such as subcutaneous injection, the carrier preferably comprises water, saline, alcohol, a fat, a wax or a buffer. For oral administration, any of the above carriers or a solid carrier, such as mannitol, lactose, starch, magnesium stearate, sodium saccharine, talcum, cellulose, glucose, sucrose, and magnesium carbonate, may be employed. Biodegradable microspheres (*e.g.*, polylactate polyglycolate) may also be employed as carriers for the pharmaceutical compositions of this invention. Suitable biodegradable microspheres are disclosed, for example, in U.S. Patent Nos. 4,897,268; 5,075,109; 5,928,647; 5,811,128; 5,820,883; 5,853,763; 5,814,344 and 5,942,252. One may also employ a carrier comprising the particulate-protein complexes described in U.S. Patent No. 5,928,647, which are capable of inducing a class I-restricted cytotoxic T lymphocyte responses in a host.

Such compositions may also comprise buffers (*e.g.*, neutral buffered saline or phosphate buffered saline), carbohydrates (*e.g.*, glucose, mannose, sucrose or dextrans), mannitol, proteins, polypeptides or amino acids such as glycine, antioxidants, bacteriostats, chelating agents such as EDTA or glutathione, adjuvants (*e.g.*, aluminum hydroxide), solutes that render the formulation isotonic, hypotonic or weakly hypertonic

with the blood of a recipient, suspending agents, thickening agents and/or preservatives. Alternatively, compositions of the present invention may be formulated as a lyophilizate. Compounds may also be encapsulated within liposomes using well known technology.

5                   Any of a variety of immunostimulants may be employed in the vaccines of this invention. For example, an adjuvant may be included. Most adjuvants contain a substance designed to protect the antigen from rapid catabolism, such as aluminum hydroxide or mineral oil, and a stimulator of immune responses, such as lipid A, *Bordetella pertussis* or *Mycobacterium tuberculosis* derived proteins. Suitable  
10 adjuvants are commercially available as, for example, Freund's Incomplete Adjuvant and Complete Adjuvant (Difco Laboratories, Detroit, MI); Merck Adjuvant 65 (Merck and Company, Inc., Rahway, NJ); AS-2 (SmithKline Beecham, Philadelphia, PA); aluminum salts such as aluminum hydroxide gel (alum) or aluminum phosphate; salts of calcium, iron or zinc; an insoluble suspension of acylated tyrosine; acylated sugars;  
15 cationically or anionically derivatized polysaccharides; polyphosphazenes; biodegradable microspheres; monophosphoryl lipid A and quil A. Cytokines, such as GM-CSF or interleukin-2, -7, or -12, may also be used as adjuvants.

                  Within the vaccines provided herein, the adjuvant composition is preferably designed to induce an immune response predominantly of the Th1 type.  
20 High levels of Th1-type cytokines (*e.g.*, IFN- $\gamma$ , TNF $\alpha$ , IL-2 and IL-12) tend to favor the induction of cell mediated immune responses to an administered antigen. In contrast, high levels of Th2-type cytokines (*e.g.*, IL-4, IL-5, IL-6 and IL-10) tend to favor the induction of humoral immune responses. Following application of a vaccine as provided herein, a patient will support an immune response that includes Th1- and Th2-  
25 type responses. Within a preferred embodiment, in which a response is predominantly Th1-type, the level of Th1-type cytokines will increase to a greater extent than the level of Th2-type cytokines. The levels of these cytokines may be readily assessed using standard assays. For a review of the families of cytokines, see Mosmann and Coffman, *Ann. Rev. Immunol.* 7:145-173, 1989.

30                   Preferred adjuvants for use in eliciting a predominantly Th1-type response include, for example, a combination of monophosphoryl lipid A, preferably 3-

de-O-acylated monophosphoryl lipid A (3D-MPL), together with an aluminum salt. MPL adjuvants are available from Corixa Corporation (Seattle, WA; *see* US Patent Nos. 4,436,727; 4,877,611; 4,866,034 and 4,912,094). CpG-containing oligonucleotides (in which the CpG dinucleotide is unmethylated) also induce a predominantly Th1  
5 response. Such oligonucleotides are well known and are described, for example, in WO 96/02555, WO 99/33488 and U.S. Patent Nos. 6,008,200 and 5,856,462. Immunostimulatory DNA sequences are also described, for example, by Sato et al., *Science* 273:352, 1996. Another preferred adjuvant is a saponin, preferably QS21 (Aquila Biopharmaceuticals Inc., Framingham, MA), which may be used alone or in  
10 combination with other adjuvants. For example, an enhanced system involves the combination of a monophosphoryl lipid A and saponin derivative, such as the combination of QS21 and 3D-MPL as described in WO 94/00153, or a less reactogenic composition where the QS21 is quenched with cholesterol, as described in WO 96/33739. Other preferred formulations comprise an oil-in-water emulsion and  
15 tocopherol. A particularly potent adjuvant formulation involving QS21, 3D-MPL and tocopherol in an oil-in-water emulsion is described in WO 95/17210.

Other preferred adjuvants include Montanide ISA 720 (Seppic, France), SAF (Chiron, California, United States), ISCOMS (CSL), MF-59 (Chiron), the SBAS series of adjuvants (*e.g.*, SBAS-2 or SBAS-4, available from SmithKline Beecham,  
20 Rixensart, Belgium), Detox (Corixa, Hamilton, MT), RC-529 (Corixa, Hamilton, MT) and other aminoalkyl glucosaminide 4-phosphates (AGPs), such as those described in pending U.S. Patent Application Serial Nos. 08/853,826 and 09/074,720, the disclosures of which are incorporated herein by reference in their entireties.

Any vaccine provided herein may be prepared using well known  
25 methods that result in a combination of antigen, immune response enhancer and a suitable carrier or excipient. The compositions described herein may be administered as part of a sustained release formulation (*i.e.*, a formulation such as a capsule, sponge or gel (composed of polysaccharides, for example) that effects a slow release of compound following administration). Such formulations may generally be prepared using well  
30 known technology (*see, e.g.*, Coombes et al., *Vaccine* 14:1429-1438, 1996) and administered by, for example, oral, rectal or subcutaneous implantation, or by

implantation at the desired target site. Sustained-release formulations may contain a polypeptide, polynucleotide or antibody dispersed in a carrier matrix and/or contained within a reservoir surrounded by a rate controlling membrane.

Carriers for use within such formulations are biocompatible, and may also be biodegradable; preferably the formulation provides a relatively constant level of active component release. Such carriers include microparticles of poly(lactide-co-glycolide), polyacrylate, latex, starch, cellulose, dextran and the like. Other delayed-release carriers include supramolecular biovectors, which comprise a non-liquid hydrophilic core (*e.g.*, a cross-linked polysaccharide or oligosaccharide) and, optionally, an external layer comprising an amphiphilic compound, such as a phospholipid (*see e.g.*, U.S. Patent No. 5,151,254 and PCT applications WO 94/20078, WO/94/23701 and WO 96/06638). The amount of active compound contained within a sustained release formulation depends upon the site of implantation, the rate and expected duration of release and the nature of the condition to be treated or prevented.

Any of a variety of delivery vehicles may be employed within pharmaceutical compositions and vaccines to facilitate production of an antigen-specific immune response that targets tumor cells. Delivery vehicles include antigen presenting cells (APCs), such as dendritic cells, macrophages, B cells, monocytes and other cells that may be engineered to be efficient APCs. Such cells may, but need not, be genetically modified to increase the capacity for presenting the antigen, to improve activation and/or maintenance of the T cell response, to have anti-tumor effects *per se* and/or to be immunologically compatible with the receiver (*i.e.*, matched HLA haplotype). APCs may generally be isolated from any of a variety of biological fluids and organs, including tumor and peritumoral tissues, and may be autologous, allogeneic, syngeneic or xenogeneic cells.

Certain preferred embodiments of the present invention use dendritic cells or progenitors thereof as antigen-presenting cells. Dendritic cells are highly potent APCs (Banchereau and Steinman, *Nature* 392:245-251, 1998) and have been shown to be effective as a physiological adjuvant for eliciting prophylactic or therapeutic antitumor immunity (*see* Timmerman and Levy, *Ann. Rev. Med.* 50:507-529, 1999). In general, dendritic cells may be identified based on their typical shape (*stellate in situ*,

with marked cytoplasmic processes (dendrites) visible *in vitro*), their ability to take up, process and present antigens with high efficiency and their ability to activate naïve T cell responses. Dendritic cells may, of course, be engineered to express specific cell-surface receptors or ligands that are not commonly found on dendritic cells *in vivo* or *ex vivo*, and such modified dendritic cells are contemplated by the present invention. As an alternative to dendritic cells, secreted vesicles antigen-loaded dendritic cells (called exosomes) may be used within a vaccine (see Zitvogel et al., *Nature Med.* 4:594-600, 1998).

Dendritic cells and progenitors may be obtained from peripheral blood, bone marrow, tumor-infiltrating cells, peritumoral tissues-infiltrating cells, lymph nodes, spleen, skin, umbilical cord blood or any other suitable tissue or fluid. For example, dendritic cells may be differentiated *ex vivo* by adding a combination of cytokines such as GM-CSF, IL-4, IL-13 and/or TNF $\alpha$  to cultures of monocytes harvested from peripheral blood. Alternatively, CD34 positive cells harvested from peripheral blood, umbilical cord blood or bone marrow may be differentiated into dendritic cells by adding to the culture medium combinations of GM-CSF, IL-3, TNF $\alpha$ , CD40 ligand, LPS, flt3 ligand and/or other compound(s) that induce differentiation, maturation and proliferation of dendritic cells.

Dendritic cells are conveniently categorized as "immature" and "mature" cells, which allows a simple way to discriminate between two well characterized phenotypes. However, this nomenclature should not be construed to exclude all possible intermediate stages of differentiation. Immature dendritic cells are characterized as APC with a high capacity for antigen uptake and processing, which correlates with the high expression of Fc $\gamma$  receptor and mannose receptor. The mature phenotype is typically characterized by a lower expression of these markers, but a high expression of cell surface molecules responsible for T cell activation such as class I and class II MHC, adhesion molecules (*e.g.*, CD54 and CD11) and costimulatory molecules (*e.g.*, CD40, CD80, CD86 and 4-1BB).

APCs may generally be transfected with a polynucleotide encoding a breast tumor protein (or portion or other variant thereof) such that the breast tumor polypeptide, or an immunogenic portion thereof, is expressed on the cell surface. Such

transfection may take place *ex vivo*, and a composition or vaccine comprising such transfected cells may then be used for therapeutic purposes, as described herein. Alternatively, a gene delivery vehicle that targets a dendritic or other antigen presenting cell may be administered to a patient, resulting in transfection that occurs *in vivo*. *In vivo* and *ex vivo* transfection of dendritic cells, for example, may generally be performed using any methods known in the art, such as those described in WO 97/24447, or the gene gun approach described by Mahvi et al., *Immunology and cell Biology* 75:456-460, 1997. Antigen loading of dendritic cells may be achieved by incubating dendritic cells or progenitor cells with the breast tumor polypeptide, DNA (naked or within a plasmid vector) or RNA; or with antigen-expressing recombinant bacterium or viruses (*e.g.*, vaccinia, fowlpox, adenovirus or lentivirus vectors). Prior to loading, the polypeptide may be covalently conjugated to an immunological partner that provides T cell help (*e.g.*, a carrier molecule). Alternatively, a dendritic cell may be pulsed with a non-conjugated immunological partner, separately or in the presence of the polypeptide.

Vaccines and pharmaceutical compositions may be presented in unit-dose or multi-dose containers, such as sealed ampoules or vials. Such containers are preferably hermetically sealed to preserve sterility of the formulation until use. In general, formulations may be stored as suspensions, solutions or emulsions in oily or aqueous vehicles. Alternatively, a vaccine or pharmaceutical composition may be stored in a freeze-dried condition requiring only the addition of a sterile liquid carrier immediately prior to use.

#### CANCER THERAPY

In further aspects of the present invention, the compositions described herein may be used for immunotherapy of cancer, such as breast cancer. Within such methods, pharmaceutical compositions and vaccines are typically administered to a patient. As used herein, a "patient" refers to any warm-blooded animal, preferably a human. A patient may or may not be afflicted with cancer. Accordingly, the above pharmaceutical compositions and vaccines may be used to prevent the development of a cancer or to treat a patient afflicted with a cancer. A cancer may be diagnosed using

criteria generally accepted in the art, including the presence of a malignant tumor. Pharmaceutical compositions and vaccines may be administered either prior to or following surgical removal of primary tumors and/or treatment such as administration of radiotherapy or conventional chemotherapeutic drugs. Administration may be by any  
5 suitable method, including administration by intravenous, intraperitoneal, intramuscular, subcutaneous, intranasal, intradermal, anal, vaginal, topical and oral routes.

Within certain embodiments, immunotherapy may be active immunotherapy, in which treatment relies on the *in vivo* stimulation of the endogenous  
10 host immune system to react against tumors with the administration of immune response-modifying agents (such as polypeptides and polynucleotides as provided herein).

Within other embodiments, immunotherapy may be passive immunotherapy, in which treatment involves the delivery of agents with established  
15 tumor-immune reactivity (such as effector cells or antibodies) that can directly or indirectly mediate antitumor effects and does not necessarily depend on an intact host immune system. Examples of effector cells include T cells as discussed above, T lymphocytes (such as CD8<sup>+</sup> cytotoxic T lymphocytes and CD4<sup>+</sup> T-helper tumor-infiltrating lymphocytes), killer cells (such as Natural Killer cells and lymphokine-  
20 activated killer cells), B cells and antigen-presenting cells (such as dendritic cells and macrophages) expressing a polypeptide provided herein. T cell receptors and antibody receptors specific for the polypeptides recited herein may be cloned, expressed and transferred into other vectors or effector cells for adoptive immunotherapy. The polypeptides provided herein may also be used to generate antibodies or anti-idiotypic  
25 antibodies (as described above and in U.S. Patent No. 4,918,164) for passive immunotherapy.

Effector cells may generally be obtained in sufficient quantities for adoptive immunotherapy by growth *in vitro*, as described herein. Culture conditions for expanding single antigen-specific effector cells to several billion in number with  
30 retention of antigen recognition *in vivo* are well known in the art. Such *in vitro* culture conditions typically use intermittent stimulation with antigen, often in the presence of

cytokines (such as IL-2) and non-dividing feeder cells. As noted above, immunoreactive polypeptides as provided herein may be used to rapidly expand antigen-specific T cell cultures in order to generate a sufficient number of cells for immunotherapy. In particular, antigen-presenting cells, such as dendritic, macrophage, monocyte, fibroblast and/or B cells, may be pulsed with immunoreactive polypeptides or transfected with one or more polynucleotides using standard techniques well known in the art. For example, antigen-presenting cells can be transfected with a polynucleotide having a promoter appropriate for increasing expression in a recombinant virus or other expression system. Cultured effector cells for use in therapy must be able to grow and distribute widely, and to survive long term *in vivo*. Studies have shown that cultured effector cells can be induced to grow *in vivo* and to survive long term in substantial numbers by repeated stimulation with antigen supplemented with IL-2 (*see, for example, Cheever et al., Immunological Reviews 157:177, 1997*).

Alternatively, a vector expressing a polypeptide recited herein may be introduced into antigen presenting cells taken from a patient and clonally propagated *ex vivo* for transplant back into the same patient. Transfected cells may be reintroduced into the patient using any means known in the art, preferably in sterile form by intravenous, intracavitary, intraperitoneal or intratumor administration.

Routes and frequency of administration of the therapeutic compositions described herein, as well as dosage, will vary from individual to individual, and may be readily established using standard techniques. In general, the pharmaceutical compositions and vaccines may be administered by injection (*e.g.*, intracutaneous, intramuscular, intravenous or subcutaneous), intranasally (*e.g.*, by aspiration) or orally. Preferably, between 1 and 10 doses may be administered over a 52 week period. Preferably, 6 doses are administered, at intervals of 1 month, and booster vaccinations may be given periodically thereafter. Alternate protocols may be appropriate for individual patients. A suitable dose is an amount of a compound that, when administered as described above, is capable of promoting an anti-tumor immune response, and is at least 10-50% above the basal (*i.e.*, untreated) level. Such response can be monitored by measuring the anti-tumor antibodies in a patient or by vaccine-dependent generation of cytolytic effector cells capable of killing the patient's tumor



cells *in vitro*. Such vaccines should also be capable of causing an immune response that leads to an improved clinical outcome (e.g., more frequent remissions, complete or partial or longer disease-free survival) in vaccinated patients as compared to non-vaccinated patients. In general, for pharmaceutical compositions and vaccines  
5 comprising one or more polypeptides, the amount of each polypeptide present in a dose ranges from about 25 µg to 5 mg per kg of host. Suitable dose sizes will vary with the size of the patient, but will typically range from about 0.1 mL to about 5 mL.

In general, an appropriate dosage and treatment regimen provides the active compound(s) in an amount sufficient to provide therapeutic and/or prophylactic  
10 benefit. Such a response can be monitored by establishing an improved clinical outcome (e.g., more frequent remissions, complete or partial, or longer disease-free survival) in treated patients as compared to non-treated patients. Increases in preexisting immune responses to a breast tumor protein generally correlate with an improved clinical outcome. Such immune responses may generally be evaluated using  
15 standard proliferation, cytotoxicity or cytokine assays, which may be performed using samples obtained from a patient before and after treatment.

#### CANCER DETECTION AND DIAGNOSIS

In general, a cancer may be detected in a patient based on the presence of one or more breast tumor proteins and/or polynucleotides encoding such proteins in a  
20 biological sample (for example, blood, sera, sputum urine and/or tumor biopsies) obtained from the patient. In other words, such proteins may be used as markers to indicate the presence or absence of a cancer such as breast cancer. In addition, such proteins may be useful for the detection of other cancers. The binding agents provided herein generally permit detection of the level of antigen that binds to the agent in the  
25 biological sample. Polynucleotide primers and probes may be used to detect the level of mRNA encoding a tumor protein, which is also indicative of the presence or absence of a cancer. In general, a breast tumor sequence should be present at a level that is at least three fold higher in tumor tissue than in normal tissue

There are a variety of assay formats known to those of ordinary skill in  
30 the art for using a binding agent to detect polypeptide markers in a sample. See, e.g.,

Harlow and Lane, *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory, 1988. In general, the presence or absence of a cancer in a patient may be determined by (a) contacting a biological sample obtained from a patient with a binding agent; (b) detecting in the sample a level of polypeptide that binds to the binding agent; and (c) 5 comparing the level of polypeptide with a predetermined cut-off value.

In a preferred embodiment, the assay involves the use of binding agent immobilized on a solid support to bind to and remove the polypeptide from the remainder of the sample. The bound polypeptide may then be detected using a detection reagent that contains a reporter group and specifically binds to the binding 10 agent/polypeptide complex. Such detection reagents may comprise, for example, a binding agent that specifically binds to the polypeptide or an antibody or other agent that specifically binds to the binding agent, such as an anti-immunoglobulin, protein G, protein A or a lectin. Alternatively, a competitive assay may be utilized, in which a polypeptide is labeled with a reporter group and allowed to bind to the immobilized 15 binding agent after incubation of the binding agent with the sample. The extent to which components of the sample inhibit the binding of the labeled polypeptide to the binding agent is indicative of the reactivity of the sample with the immobilized binding agent. Suitable polypeptides for use within such assays include full length breast tumor proteins and portions thereof to which the binding agent binds, as described above.

20 The solid support may be any material known to those of ordinary skill in the art to which the tumor protein may be attached. For example, the solid support may be a test well in a microtiter plate or a nitrocellulose or other suitable membrane. Alternatively, the support may be a bead or disc, such as glass, fiberglass, latex or a plastic material such as polystyrene or polyvinylchloride. The support may also be a 25 magnetic particle or a fiber optic sensor, such as those disclosed, for example, in U.S. Patent No. 5,359,681. The binding agent may be immobilized on the solid support using a variety of techniques known to those of skill in the art, which are amply described in the patent and scientific literature. In the context of the present invention, the term "immobilization" refers to both noncovalent association, such as adsorption, 30 and covalent attachment (which may be a direct linkage between the agent and functional groups on the support or may be a linkage by way of a cross-linking agent).

Immobilization by adsorption to a well in a microtiter plate or to a membrane is preferred. In such cases, adsorption may be achieved by contacting the binding agent, in a suitable buffer, with the solid support for a suitable amount of time. The contact time varies with temperature, but is typically between about 1 hour and about 1 day. In  
5 general, contacting a well of a plastic microtiter plate (such as polystyrene or polyvinylchloride) with an amount of binding agent ranging from about 10 ng to about 10 µg, and preferably about 100 ng to about 1 µg, is sufficient to immobilize an adequate amount of binding agent.

Covalent attachment of binding agent to a solid support may generally be  
10 achieved by first reacting the support with a bifunctional reagent that will react with both the support and a functional group, such as a hydroxyl or amino group, on the binding agent. For example, the binding agent may be covalently attached to supports having an appropriate polymer coating using benzoquinone or by condensation of an aldehyde group on the support with an amine and an active hydrogen on the binding  
15 partner (*see, e.g.,* Pierce Immunotechnology Catalog and Handbook, 1991, at A12-A13).

In certain embodiments, the assay is a two-antibody sandwich assay. This assay may be performed by first contacting an antibody that has been immobilized on a solid support, commonly the well of a microtiter plate, with the sample, such that  
20 polypeptides within the sample are allowed to bind to the immobilized antibody. Unbound sample is then removed from the immobilized polypeptide-antibody complexes and a detection reagent (preferably a second antibody capable of binding to a different site on the polypeptide) containing a reporter group is added. The amount of detection reagent that remains bound to the solid support is then determined using a  
25 method appropriate for the specific reporter group.

More specifically, once the antibody is immobilized on the support as described above, the remaining protein binding sites on the support are typically blocked. Any suitable blocking agent known to those of ordinary skill in the art, such as bovine serum albumin or Tween 20™ (Sigma Chemical Co., St. Louis, MO). The  
30 immobilized antibody is then incubated with the sample, and polypeptide is allowed to bind to the antibody. The sample may be diluted with a suitable diluent, such as

phosphate-buffered saline (PBS) prior to incubation. In general, an appropriate contact time (*i.e.*, incubation time) is a period of time that is sufficient to detect the presence of polypeptide within a sample obtained from an individual with breast cancer. Preferably, the contact time is sufficient to achieve a level of binding that is at least about 95% of that achieved at equilibrium between bound and unbound polypeptide. Those of ordinary skill in the art will recognize that the time necessary to achieve equilibrium may be readily determined by assaying the level of binding that occurs over a period of time. At room temperature, an incubation time of about 30 minutes is generally sufficient.

10                   Unbound sample may then be removed by washing the solid support with an appropriate buffer, such as PBS containing 0.1% Tween 20™. The second antibody, which contains a reporter group, may then be added to the solid support. Preferred reporter groups include those groups recited above.

                  The detection reagent is then incubated with the immobilized antibody-polypeptide complex for an amount of time sufficient to detect the bound polypeptide. An appropriate amount of time may generally be determined by assaying the level of binding that occurs over a period of time. Unbound detection reagent is then removed and bound detection reagent is detected using the reporter group. The method employed for detecting the reporter group depends upon the nature of the reporter group. For radioactive groups, scintillation counting or autoradiographic methods are generally appropriate. Spectroscopic methods may be used to detect dyes, luminescent groups and fluorescent groups. Biotin may be detected using avidin, coupled to a different reporter group (commonly a radioactive or fluorescent group or an enzyme). Enzyme reporter groups may generally be detected by the addition of substrate (generally for a specific period of time), followed by spectroscopic or other analysis of the reaction products.

                  To determine the presence or absence of a cancer, such as breast cancer, the signal detected from the reporter group that remains bound to the solid support is generally compared to a signal that corresponds to a predetermined cut-off value. In one preferred embodiment, the cut-off value for the detection of a cancer is the average mean signal obtained when the immobilized antibody is incubated with samples from

patients without the cancer. In general, a sample generating a signal that is three standard deviations above the predetermined cut-off value is considered positive for the cancer. In an alternate preferred embodiment, the cut-off value is determined using a Receiver Operator Curve, according to the method of Sackett et al., *Clinical*  
5 *Epidemiology: A Basic Science for Clinical Medicine*, Little Brown and Co., 1985, p. 106-7. Briefly, in this embodiment, the cut-off value may be determined from a plot of pairs of true positive rates (*i.e.*, sensitivity) and false positive rates (100%-specificity) that correspond to each possible cut-off value for the diagnostic test result. The cut-off value on the plot that is the closest to the upper left-hand corner (*i.e.*, the value that  
10 encloses the largest area) is the most accurate cut-off value, and a sample generating a signal that is higher than the cut-off value determined by this method may be considered positive. Alternatively, the cut-off value may be shifted to the left along the plot, to minimize the false positive rate, or to the right, to minimize the false negative rate. In general, a sample generating a signal that is higher than the cut-off value determined by  
15 this method is considered positive for a cancer.

In a related embodiment, the assay is performed in a flow-through or strip test format, wherein the binding agent is immobilized on a membrane, such as nitrocellulose. In the flow-through test, polypeptides within the sample bind to the immobilized binding agent as the sample passes through the membrane. A second,  
20 labeled binding agent then binds to the binding agent-polypeptide complex as a solution containing the second binding agent flows through the membrane. The detection of bound second binding agent may then be performed as described above. In the strip test format, one end of the membrane to which binding agent is bound is immersed in a solution containing the sample. The sample migrates along the membrane through a  
25 region containing second binding agent and to the area of immobilized binding agent. Concentration of second binding agent at the area of immobilized antibody indicates the presence of a cancer. Typically, the concentration of second binding agent at that site generates a pattern, such as a line, that can be read visually. The absence of such a pattern indicates a negative result. In general, the amount of binding agent immobilized  
30 on the membrane is selected to generate a visually discernible pattern when the biological sample contains a level of polypeptide that would be sufficient to generate a

positive signal in the two-antibody sandwich assay, in the format discussed above. Preferred binding agents for use in such assays are antibodies and antigen-binding fragments thereof. Preferably, the amount of antibody immobilized on the membrane ranges from about 25 ng to about 1 µg, and more preferably from about 50 ng to about 500 ng. Such tests can typically be performed with a very small amount of biological sample.

Of course, numerous other assay protocols exist that are suitable for use with the tumor proteins or binding agents of the present invention. The above descriptions are intended to be exemplary only. For example, it will be apparent to those of ordinary skill in the art that the above protocols may be readily modified to use breast tumor polypeptides to detect antibodies that bind to such polypeptides in a biological sample. The detection of such breast tumor protein specific antibodies may correlate with the presence of a cancer.

A cancer may also, or alternatively, be detected based on the presence of T cells that specifically react with a breast tumor protein in a biological sample. Within certain methods, a biological sample comprising CD4<sup>+</sup> and/or CD8<sup>+</sup> T cells isolated from a patient is incubated with a breast tumor polypeptide, a polynucleotide encoding such a polypeptide and/or an APC that expresses at least an immunogenic portion of such a polypeptide, and the presence or absence of specific activation of the T cells is detected. Suitable biological samples include, but are not limited to, isolated T cells. For example, T cells may be isolated from a patient by routine techniques (such as by Ficoll/Hypaque density gradient centrifugation of peripheral blood lymphocytes). T cells may be incubated *in vitro* for 2-9 days (typically 4 days) at 37°C with polypeptide (e.g., 5 - 25 µg/ml). It may be desirable to incubate another aliquot of a T cell sample in the absence of breast tumor polypeptide to serve as a control. For CD4<sup>+</sup> T cells, activation is preferably detected by evaluating proliferation of the T cells. For CD8<sup>+</sup> T cells, activation is preferably detected by evaluating cytolytic activity. A level of proliferation that is at least two fold greater and/or a level of cytolytic activity that is at least 20% greater than in disease-free patients indicates the presence of a cancer in the patient.

As noted above, a cancer may also, or alternatively, be detected based on the level of mRNA encoding a breast tumor protein in a biological sample. For example, at least two oligonucleotide primers may be employed in a polymerase chain reaction (PCR) based assay to amplify a portion of a breast tumor cDNA derived from a biological sample, wherein at least one of the oligonucleotide primers is specific for (i.e., hybridizes to) a polynucleotide encoding the breast tumor protein. The amplified cDNA is then separated and detected using techniques well known in the art, such as gel electrophoresis. Similarly, oligonucleotide probes that specifically hybridize to a polynucleotide encoding a breast tumor protein may be used in a hybridization assay to detect the presence of polynucleotide encoding the tumor protein in a biological sample.

To permit hybridization under assay conditions, oligonucleotide primers and probes should comprise an oligonucleotide sequence that has at least about 60%, preferably at least about 75% and more preferably at least about 90%, identity to a portion of a polynucleotide encoding a breast tumor protein that is at least 10 nucleotides, and preferably at least 20 nucleotides, in length. Preferably, oligonucleotide primers and/or probes hybridize to a polynucleotide encoding a polypeptide described herein under moderately stringent conditions, as defined above. Oligonucleotide primers and/or probes which may be usefully employed in the diagnostic methods described herein preferably are at least 10-40 nucleotides in length. In a preferred embodiment, the oligonucleotide primers comprise at least 10 contiguous nucleotides, more preferably at least 15 contiguous nucleotides, of a DNA molecule having a sequence recited in SEQ ID NO: 1-38, 42-204, 205, 207 and 210-290. Techniques for both PCR based assays and hybridization assays are well known in the art (see, for example, Mullis et al., *Cold Spring Harbor Symp. Quant. Biol.*, 51:263, 1987; Erlich ed., *PCR Technology*, Stockton Press, NY, 1989).

One preferred assay employs RT-PCR, in which PCR is applied in conjunction with reverse transcription. Typically, RNA is extracted from a biological sample, such as biopsy tissue, and is reverse transcribed to produce cDNA molecules. PCR amplification using at least one specific primer generates a cDNA molecule, which may be separated and visualized using, for example, gel electrophoresis. Amplification may be performed on biological samples taken from a test patient and from an

individual who is not afflicted with a cancer. The amplification reaction may be performed on several dilutions of cDNA spanning two orders of magnitude. A two-fold or greater increase in expression in several dilutions of the test patient sample as compared to the same dilutions of the non-cancerous sample is typically considered  
5 positive.

In another embodiment, the compositions described herein may be used as markers for the progression of cancer. In this embodiment, assays as described above for the diagnosis of a cancer may be performed over time, and the change in the level of reactive polypeptide(s) or polynucleotide(s) evaluated. For example, the assays  
10 may be performed every 24-72 hours for a period of 6 months to 1 year, and thereafter performed as needed. In general, a cancer is progressing in those patients in whom the level of polypeptide or polynucleotide detected increases over time. In contrast, the cancer is not progressing when the level of reactive polypeptide or polynucleotide either remains constant or decreases with time.

15 Certain *in vivo* diagnostic assays may be performed directly on a tumor. One such assay involves contacting tumor cells with a binding agent. The bound binding agent may then be detected directly or indirectly via a reporter group. Such binding agents may also be used in histological applications. Alternatively, polynucleotide probes may be used within such applications.

20 As noted above, to improve sensitivity, multiple breast tumor protein markers may be assayed within a given sample. It will be apparent that binding agents specific for different proteins provided herein may be combined within a single assay. Further, multiple primers or probes may be used concurrently. The selection of tumor protein markers may be based on routine experiments to determine combinations that  
25 results in optimal sensitivity. In addition, or alternatively, assays for tumor proteins provided herein may be combined with assays for other known tumor antigens.

#### **DIAGNOSTIC KITS**

The present invention further provides kits for use within any of the above diagnostic methods. Such kits typically comprise two or more components  
30 necessary for performing a diagnostic assay. Components may be compounds,



reagents, containers and/or equipment. For example, one container within a kit may contain a monoclonal antibody or fragment thereof that specifically binds to a breast tumor protein. Such antibodies or fragments may be provided attached to a support material, as described above. One or more additional containers may enclose elements,  
5 such as reagents or buffers, to be used in the assay. Such kits may also, or alternatively, contain a detection reagent as described above that contains a reporter group suitable for direct or indirect detection of antibody binding.

Alternatively, a kit may be designed to detect the level of mRNA encoding a breast tumor protein in a biological sample. Such kits generally comprise at  
10 least one oligonucleotide probe or primer, as described above, that hybridizes to a polynucleotide encoding a breast tumor protein. Such an oligonucleotide may be used, for example, within a PCR or hybridization assay. Additional components that may be present within such kits include a second oligonucleotide and/or a diagnostic reagent or container to facilitate the detection of a polynucleotide encoding a breast tumor protein.

15 The following Examples are offered by way of illustration and not by way of limitation.

## EXAMPLE 1

IDENTIFICATION OF BREAST TUMOR PROTEIN cDNAS USING  
SUBTRACTION METHODOLOGY

This Example illustrates the identification of cDNA molecules encoding  
5 breast tumor proteins.

A human metastatic breast tumor cDNA expression library was constructed from metastatic breast tumor poly A<sup>+</sup> RNA using a Superscript Plasmid System for cDNA Synthesis and Plasmid Cloning kit (BRL Life Technologies, Gaithersburg, MD 20897) following the manufacturer's protocol. Specifically, breast  
10 tumor tissues were homogenized with polytron (Kinematica, Switzerland) and total RNA was extracted using Trizol reagent (BRL Life Technologies) as directed by the manufacturer. The poly A<sup>+</sup> RNA was then purified using a Qiagen oligotex spin column mRNA purification kit (Qiagen, Santa Clarita, CA 91355) according to the manufacturer's protocol. First-strand cDNA was synthesized using the NotI/Oligo-  
15 dT18 primer. Double-stranded cDNA was synthesized, ligated with EcoRI/BstX I adaptors (Invitrogen, Carlsbad, CA) and digested with NotI. Following size fractionation with Chroma Spin-1000 columns (Clontech, Palo Alto, CA 94303), the cDNA was ligated into the EcoRI/NotI site of pCDNA3.1 (Invitrogen, Carlsbad, CA) and transformed into ElectroMax *E. coli* DH10B cells (BRL Life Technologies) by  
20 electroporation.

Using the same procedure, a normal human breast cDNA expression library was prepared from a pool of four normal breast tissue specimens. The cDNA libraries were characterized by determining the number of independent colonies, the percentage of clones that carried insert, the average insert size and by sequence analysis.  
25 Sequencing analysis showed both libraries to contain good complex cDNA clones that were synthesized from mRNA, with minimal rRNA and mitochondrial DNA contamination sequencing.

A cDNA subtracted library (referred to as BS3) was prepared using the above metastatic breast tumor and normal breast cDNA libraries, as described by Hara  
30 *et al.* (*Blood*, 84:189-199, 1994) with some modifications. Specifically, a breast tumor-

specific subtracted cDNA library was generated as follows. Normal breast cDNA library (70 µg) was digested with EcoRI, NotI, and SfuI, followed by a filling-in reaction with DNA polymerase Klenow fragment. After phenol-chloroform extraction and ethanol precipitation, the DNA was dissolved in 100 µl of H<sub>2</sub>O, heat-denatured and mixed with 100 µl (100 µg) of Photoprobe biotin (Vector Laboratories, Burlingame, CA), the resulting mixture was irradiated with a 270 W sunlamp on ice for 20 minutes. Additional Photoprobe biotin (50 µl) was added and the biotinylation reaction was repeated. After extraction with butanol five times, the DNA was ethanol-precipitated and dissolved in 23 µl H<sub>2</sub>O to form the driver DNA.

10 To form the tracer DNA, 10 µg breast tumor cDNA library was digested with BamHI and XhoI, phenol chloroform extracted and passed through Chroma spin-400 columns (Clontech). Following ethanol precipitation, the tracer DNA was dissolved in 5 µl H<sub>2</sub>O. Tracer DNA was mixed with 15 µl driver DNA and 20 µl of 2 x hybridization buffer (1.5 M NaCl/10 mM EDTA/50 mM HEPES pH 7.5/0.2% sodium dodecyl sulfate), overlaid with mineral oil, and heat-denatured completely. The sample was immediately transferred into a 68 °C water bath and incubated for 20 hours (long hybridization [LH]). The reaction mixture was then subjected to a streptavidin treatment followed by phenol/chloroform extraction. This process was repeated three more times. Subtracted DNA was precipitated, dissolved in 12 µl H<sub>2</sub>O, mixed with 8 µl driver DNA and 20 µl of 2 x hybridization buffer, and subjected to a hybridization at 68 °C for 2 hours (short hybridization [SH]). After removal of biotinylated double-stranded DNA, subtracted cDNA was ligated into BamHI/XhoI site of chloramphenicol resistant pBCSK<sup>+</sup> (Stratagene, La Jolla, CA 92037) and transformed into ElectroMax *E. coli* DH10B cells by electroporation to generate a breast tumor specific subtracted cDNA library.

To analyze the subtracted cDNA library, plasmid DNA was prepared from independent clones, randomly picked from the subtracted breast tumor specific library and characterized by DNA sequencing with a Perkin Elmer/Applied Biosystems Division Automated Sequencer Model 373A (Foster City, CA).

30 A second cDNA subtraction library containing cDNA from breast tumor subtracted with normal breast cDNA, and known as BT, was constructed as follows.

Total RNA was extracted from primary breast tumor tissues using Trizol reagent (Gibco BRL Life Technologies, Gaithersburg, MD) as described by the manufacturer. The polyA<sup>+</sup> RNA was purified using an oligo(dT) cellulose column according to standard protocols. First strand cDNA was synthesized using the primer supplied in a Clontech  
5 PCR-Select cDNA Subtraction Kit (Clontech, Palo Alto, CA). The driver DNA consisted of cDNAs from two normal breast tissues with the tester cDNA being from three primary breast tumors. Double-stranded cDNA was synthesized for both tester and driver, and digested with a combination of endonucleases (MluI, MscI, PvuII, SalI and StuI) which recognize six base pairs DNA. This modification increased the average  
10 cDNA size dramatically compared with cDNAs generated according to the protocol of Clontech. The digested tester cDNAs were ligated to two different adaptors and the subtraction was performed according to Clontech's protocol. The subtracted cDNAs were subjected to two rounds of PCR amplification, following the manufacturer's protocol. The resulting PCR products were subcloned into the TA cloning vector,  
15 pCRII (Invitrogen, San Diego, CA) and transformed into ElectroMax *E. coli* DH10B cells (Gibco BRL Life, Technologies) by electroporation. DNA was isolated from independent clones and sequenced using a Perkin Elmer/Applied Biosystems Division (Foster City, CA) Automated Sequencer Model 373A.

Two additional subtracted cDNA libraries were prepared from cDNA  
20 from breast tumors subtracted with a pool of cDNA from six normal tissues (liver, brain, stomach, small intestine, kidney and heart; referred to as 2BT and BC6) using the PCR-subtraction protocol of Clontech, described above. A fourth subtracted library (referred to as Bt-Met) was prepared using the protocol of Clontech from cDNA from metastatic breast tumors subtracted with cDNA from five normal tissues (brain, lung,  
25 PBMC, pancreas and normal breast).

cDNA clones isolated in the breast subtractions BS3, BT, 2BT, BC6 and BT-Met, described above, were colony PCR amplified and their mRNA expression levels in breast tumor, normal breast and various other normal tissues were determined using microarray technology. Briefly, the PCR amplification products were dotted onto  
30 slides in an array format, with each product occupying a unique location in the array. mRNA was extracted from the tissue sample to be tested, reverse transcribed, and

fluorescent-labeled cDNA probes were generated. The microarrays were probed with the labeled cDNA probes, the slides scanned and fluorescence intensity was measured. This intensity correlates with the hybridization intensity.

The determined cDNA sequences of 131 clones determined to be over-  
5 expressed in breast tumor tissue compared to other tissues tested by a visual analysis of the microarray data are provided in SEQ ID NO: 1-35 and 42-137. Comparison of these cDNA sequences with known sequences in the gene bank using the EMBL and GenBank databases revealed no significant homologies to the sequences provided in SEQ ID NO: 7, 10, 21, 26, 30, 63, 81 and 104. The sequences of SEQ ID NO: 2-5, 8, 9,  
10 13, 15, 16, 22, 25, 27, 28, 33, 35, 72, 73, 103, 107, 109, 118, 128, 129 134 and 136 showed some homology to previously isolated expressed sequences tags (ESTs), while the sequences of SEQ ID NO: 1, 6, 11, 12, 14, 17-20, 23, 24, 29, 31, 32, 34, 42-62, 64-71, 74-80, 82-102, 105, 106, 108, 110-117, 119-127, 130-133, 135 and 137 showed some homology to previously identified genes.

15 The determined cDNA sequences of an additional 45 clones isolated from the BT-Met library as described above and found to be over-expressed in breast tumors and metastatic breast tumors compared to other tissues tested, are provided in SEQ ID NO: 138-182. Comparison of the sequences of SEQ ID NO: 159-161, 164 and 181 revealed no significant homologies to previously identified sequences. The  
20 sequences of SEQ ID NO: 138-158, 162, 163, 165-180 and 182 showed some homology to previously identified genes.

In further studies, suppression subtractive hybridization (Clontech) was preformed using a pool of cDNA from 3 unique human breast tumors as the tester and a pool of cDNA from 6 other normal human tissues (liver, brain, stomach, small intestine,  
25 heart and kidney) as the driver. The isolated cDNA fragments were subcloned and characterized by DNA sequencing. The determined cDNA sequences of 22 isolated clones are provided in SEQ ID NO: 183-204. Comparison of these sequences with those in the public databases revealed no significant homologies to previously identified sequences.

30 The determined cDNA sequences of 71 additional breast-specific genes isolated during characterization of breast tumor cDNA libraries are provided in SEQ ID

NO: 210-290. Comparison of these sequences with those in the GenBank and Geneseq databases revealed no significant homologies.

## EXAMPLE 2

### 5     IDENTIFICATION OF BREAST TUMOR PROTEIN cDNAS BY RT-PCR

GABA<sub>A</sub> receptor clones were isolated from human breast cancer cDNA libraries by first preparing cDNA libraries from breast tumor samples from different patients as described above. PCR primers were designed based on the GABA<sub>A</sub> receptor subunit sequences described by Hedblom and Kirkness (*Jnl. Biol. Chem.* 272:15346-10 15350, 1997) and used to amplify sequences from the breast tumor cDNA libraries by RT-PCR. The determined cDNA sequences of three GABA<sub>A</sub> receptor clones are provided in SEQ ID NO: 36-38, with the corresponding amino acid sequences being provided in SEQ ID NO: 39-41.

The clone with the longest open reading frame (ORF; SEQ ID NO: 36) 15 showed homology to the GABA<sub>A</sub> receptor of Hedblom and Kirkness, with four potential transmembrane regions at the C-terminal part of the protein, while the clones of SEQ ID NO: 37 and 38 retained either no transmembrane region or only the first transmembrane region. Some patients were found to have only the clones with the shorter ORFs while others had both the clones with longer and shorter ORFs.

20

## EXAMPLE 3

### EXPRESSION OF OVARIAN TUMOR-DERIVED ANTIGENS IN BREAST

Isolation of the antigens O772P and O8E from ovarian tumor tissue is 25 described in US Patent Application No. 09/338,933, filed June 23, 1999. The determined cDNA sequence for O772P is provided in SEQ ID NO: 205, with the corresponding amino acid sequence being provided in SEQ ID NO: 206. The full-length cDNA sequence for O8E is provided in SEQ ID NO: 207. Two protein sequences may be translated from the full length O8E. Form "A" (SEQ ID NO: 208)

begins with a putative start methionine. A second form "B" (SEQ ID NO: 209) includes 27 additional upstream residues to the 5' end of the nucleotide sequence.

The expression levels of O772P and O8E in a variety of tumor and normal tissues, including metastatic breast tumors, were analyzed by real time PCR.

5 Both genes were found to have increased mRNA expression in 30-50% of breast tumors. For O772P, elevated expression was also observed in normal trachea, ureter, uterus and ovary. For O8E, elevated expression was also observed in normal trachea, kidney and ovary. Additional analysis employing a panel of tumor cell lines demonstrated increased expression of O8E in the breast tumor cell lines SKBR3, MDA-  
10 MB-415 and BT474, and increased expression of O772P in SKBR3. Collectively, the data indicate that O772P and O8E may be useful in the diagnosis and therapy of breast cancer.

#### EXAMPLE 4

#### 15 SYNTHESIS OF POLYPEPTIDES

Polypeptides may be synthesized on a Perkin Elmer/Applied Biosystems Division 430A peptide synthesizer using Fmoc chemistry with HPTU (O-Benzotriazole-N,N,N',N'-tetramethyluronium hexafluorophosphate) activation. A Gly-Cys-Gly sequence may be attached to the amino terminus of the peptide to provide a  
20 method of conjugation, binding to an immobilized surface, or labeling of the peptide. Cleavage of the peptides from the solid support may be carried out using the following cleavage mixture: trifluoroacetic acid:ethanedithiol:thioanisole:water:phenol (40:1:2:2:3). After cleaving for 2 hours, the peptides may be precipitated in cold methyl-t-butyl-ether. The peptide pellets may then be dissolved in water containing  
25 0.1% trifluoroacetic acid (TFA) and lyophilized prior to purification by C18 reverse phase HPLC. A gradient of 0%-60% acetonitrile (containing 0.1% TFA) in water (containing 0.1% TFA) may be used to elute the peptides. Following lyophilization of the pure fractions, the peptides may be characterized using electrospray or other types of mass spectrometry and by amino acid analysis.

30

From the foregoing it will be appreciated that, although specific embodiments of the invention have been described herein for purposes of illustration, various modifications may be made without deviating from the spirit and scope of the invention. Accordingly, the invention is not limited except as by the appended claims.



## CLAIMS

1. An isolated polypeptide, comprising at least an immunogenic portion of a breast tumor protein, or a variant thereof that differs in one or more substitutions, deletions, additions and/or insertions such that the ability of the variant to react with antigen-specific antisera is not substantially diminished, wherein the tumor protein comprises an amino acid sequence that is encoded by a polynucleotide sequence selected from the group consisting of:

(a) sequences recited in SEQ ID NOs: 2-5, 7, 10, 13, 15, 16, 21, 22, 25, 28, 30, 33, 35, 63, 72, 73, 81, 103, 104, 107, 109, 118, 128, 129, 134, 136, 159-161, 164, 181, 183-204 and 210-290;

(b) sequences that hybridize to a sequence recited in any one of SEQ ID NOs: 2-5, 7, 10, 13, 15, 16, 21, 22, 25, 28, 30, 33, 35, 63, 72, 73, 81, 103, 104, 107, 109, 118, 128, 129, 134, 136, 159-161, 164, 181, 183-204 and 210-290 under moderately stringent conditions; and

(c) complements of sequences of (a) or (b).

2. An isolated polypeptide according to claim 1, wherein the polypeptide comprises an amino acid sequence that is encoded by a polynucleotide sequence recited in any one of SEQ ID NOs: 2-5, 7, 10, 13, 15, 16, 21, 22, 25, 28, 30, 33, 35, 63, 72, 73, 81, 103, 104, 107, 109, 118, 128, 129, 134, 136, 159-161, 164, 181, 183-204 and 210-290 or a complement of any of the foregoing polynucleotide sequences.

3. An isolated polynucleotide encoding at least 15 amino acid residues of a breast tumor protein, or a variant thereof that differs in one or more substitutions, deletions, additions and/or insertions such that the ability of the variant to react with antigen-specific antisera is not substantially diminished, wherein the tumor protein comprises an amino acid sequence that is encoded by a polynucleotide comprising a sequence recited in any one of SEQ ID NOs: 2-5, 7, 10, 13, 15, 16, 21, 22,

25, 28, 30, 33, 35, 63, 72, 73, 81, 103, 104, 107, 109, 118, 128, 129, 134, 136, 159-161, 164, 181, 183-204 and 210-290 or a complement of any of the foregoing sequences.

4. An isolated polynucleotide encoding a breast tumor protein, or a variant thereof, wherein the tumor protein comprises an amino acid sequence that is encoded by a polynucleotide comprising a sequence recited in any one of SEQ ID NOs: 2-5, 7, 10, 13, 15, 16, 21, 22, 25, 28, 30, 33, 35, 63, 72, 73, 81, 103, 104, 107, 109, 118, 128, 129, 134, 136, 159-161, 164, 181, 183-204 and 210-290 or a complement of any of the foregoing sequences.

5. An isolated polynucleotide, comprising a sequence recited in any one of SEQ ID NOs: 2-5, 7, 10, 13, 15, 16, 21, 22, 25, 28, 30, 33, 35, 63, 72, 73, 81, 103, 104, 107, 109, 118, 128, 129, 134, 136, 159-161, 164, 181, 183-204 and 210-290.

6. An isolated polynucleotide, comprising a sequence that hybridizes to a sequence recited in any one of SEQ ID NOs: 2-5, 7, 10, 13, 15, 16, 21, 22, 25, 28, 30, 33, 35, 63, 72, 73, 81, 103, 104, 107, 109, 118, 128, 129, 134, 136, 159-161, 164, 181, 183-204 and 210-290 under moderately stringent conditions.

7. An isolated polynucleotide complementary to a polynucleotide according to any one of claims 3-6.

8. An expression vector, comprising a polynucleotide according to any one of claims 3-7.

9. A host cell transformed or transfected with an expression vector according to claim 8.

10. An isolated antibody, or antigen-binding fragment thereof, that specifically binds to a breast tumor protein that comprises an amino acid sequence that is encoded by a polynucleotide sequence recited in any one of SEQ ID NOs: 2-5, 7, 10, 13, 15, 16, 21, 22, 25, 28, 30, 33, 35, 63, 72, 73, 81, 103, 104, 107, 109, 118, 128, 129,

134, 136, 159-161, 164, 181, 183-204 and 210-290 or a complement of any of the foregoing polynucleotide sequences.

11. A fusion protein, comprising at least one polypeptide according to claim 1.

12. A fusion protein according to claim 11, wherein the fusion protein comprises an expression enhancer that increases expression of the fusion protein in a host cell transfected with a polynucleotide encoding the fusion protein.

13. A fusion protein according to claim 11, wherein the fusion protein comprises a T helper epitope that is not present within the polypeptide of claim 1.

14. A fusion protein according to claim 11, wherein the fusion protein comprises an affinity tag.

15. An isolated polynucleotide encoding a fusion protein according to claim 11.

16. A pharmaceutical composition, comprising a physiologically acceptable carrier and at least one component selected from the group consisting of:

- (a) a polypeptide according to claim 1;
- (b) a polynucleotide according to claim 3;
- (c) an antibody according to claim 10;
- (d) a fusion protein according to claim 11; and
- (e) a polynucleotide according to claim 15.

17. A vaccine comprising an immunostimulant and at least one component selected from the group consisting of:

- (a) a polypeptide according to claim 1;

- (b) a polynucleotide according to claim 3;
  - (c) an antibody according to claim 10;
  - (d) a fusion protein according to claim 11; and
  - (e) a polynucleotide according to claim 15.
18. A vaccine according to claim 17, wherein the immunostimulant is an adjuvant.
19. A vaccine according to any claim 17, wherein the immunostimulant induces a predominantly Type I response.
20. A method for inhibiting the development of a cancer in a patient, comprising administering to a patient an effective amount of a pharmaceutical composition according to claim 16.
21. A method for inhibiting the development of a cancer in a patient, comprising administering to a patient an effective amount of a vaccine according to claim 17.
22. A pharmaceutical composition comprising an antigen-presenting cell that expresses a polypeptide according to claim 1, in combination with a pharmaceutically acceptable carrier or excipient.
23. A pharmaceutical composition according to claim 22, wherein the antigen presenting cell is a dendritic cell or a macrophage.
24. A vaccine comprising an antigen-presenting cell that expresses a polypeptide comprising at least an immunogenic portion of a breast tumor protein, or a variant thereof, wherein the tumor protein comprises an amino acid sequence that is encoded by a polynucleotide sequence selected from the group consisting of:

- (a) sequences recited in SEQ ID NOs: 1-38, 42-204, 205, 207 and 210-290;
- (b) sequences that hybridize to a sequence recited in any one of SEQ ID NOs: 1-38, 42-204, 205, 207 and 210-290 under moderately stringent conditions; and
- (c) complements of sequences of (i) or (ii);  
in combination with an immunostimulant.

25. A vaccine according to claim 24, wherein the immunostimulant is an adjuvant.

26. A vaccine according to claim 24, wherein the immunostimulant induces a predominantly Type I response.

27. A vaccine according to claim 24, wherein the antigen-presenting cell is a dendritic cell.

28. A method for inhibiting the development of a cancer in a patient, comprising administering to a patient an effective amount of an antigen-presenting cell that expresses a polypeptide comprising at least an immunogenic portion of a breast tumor protein, or a variant thereof, wherein the tumor protein comprises an amino acid sequence that is encoded by a polynucleotide sequence selected from the group consisting of:

- (a) sequences recited in SEQ ID NOs: 1-38, 42-204, 205, 207 and 210-290;
- (b) sequences that hybridize to a sequence recited in any one of SEQ ID NOs: 1-38, 42-204, 205, 207 and 210-290 under moderately stringent conditions; and
- (c) complements of sequences of (i) or (ii) encoded by a polynucleotide recited in any one of SEQ ID NOs: 1-38, 42-204, 205, 207 and 210-290; and thereby inhibiting the development of a cancer in the patient.

29. A method according to claim 28, wherein the antigen-presenting cell is a dendritic cell.

30. A method according to any one of claims 20, 21 and 28, wherein the cancer is breast cancer.

31. A method for removing tumor cells from a biological sample, comprising contacting a biological sample with T cells that specifically react with a breast tumor protein, wherein the tumor protein comprises an amino acid sequence that is encoded by a polynucleotide sequence selected from the group consisting of:

(i) polynucleotides recited in any one of SEQ ID NOs: 1-38, 42-204, 205, 207 and 210-290; and

(ii) complements of the foregoing polynucleotides;

wherein the step of contacting is performed under conditions and for a time sufficient to permit the removal of cells expressing the antigen from the sample.

32. A method according to claim 31, wherein the biological sample is blood or a fraction thereof.

33. A method for inhibiting the development of a cancer in a patient, comprising administering to a patient a biological sample treated according to the method of claim 31.

34. A method for stimulating and/or expanding T cells specific for a breast tumor protein, comprising contacting T cells with at least one component selected from the group consisting of:

(a) polypeptides comprising at least an immunogenic portion of a breast tumor protein, or a variant thereof, wherein the tumor protein comprises an amino acid sequence that is encoded by a polynucleotide sequence selected from the group consisting of:

- (i) sequences recited in SEQ ID NOs: 1-38, 42-204, 205, 207 and 210-290;
  - (ii) sequences that hybridize to a sequence recited in any one of SEQ ID NOs: 1-38, 42-204, 205, 207 and 210-290 under moderately stringent conditions; and
  - (iii) complements of sequences of (i) or (ii);
- (b) polynucleotides encoding a polypeptide of (a); and
  - (c) antigen presenting cells that express a polypeptide of (a);
- under conditions and for a time sufficient to permit the stimulation and/or expansion of T cells.

35. An isolated T cell population, comprising T cells prepared according to the method of claim 34.

36. A method for inhibiting the development of a cancer in a patient, comprising administering to a patient an effective amount of a T cell population according to claim 35.

37. A method for inhibiting the development of a cancer in a patient, comprising the steps of:

(a) incubating CD4<sup>+</sup> and/or CD8<sup>+</sup> T cells isolated from a patient with at least one component selected from the group consisting of:

(i) polypeptides comprising at least an immunogenic portion of a breast tumor protein, or a variant thereof, wherein the tumor protein comprises an amino acid sequence that is encoded by a polynucleotide sequence selected from the group consisting of:

(1) sequences recited in SEQ ID NOs: 1-38, 42-204, 205, 207 and 210-290;

(2) sequences that hybridize to a sequence recited in any one of SEQ ID NOs: 1-38, 42-204, 205, 207 and 210-290 under moderately stringent conditions; and

- (3) complements of sequences of (1) or (2);
- (ii) polynucleotides encoding a polypeptide of (i); and
- (iii) antigen presenting cells that expresses a polypeptide of (i);

such that T cells proliferate; and

- (b) administering to the patient an effective amount of the proliferated T cells, and thereby inhibiting the development of a cancer in the patient.

38. A method for inhibiting the development of a cancer in a patient, comprising the steps of:

- (a) incubating CD4<sup>+</sup> and/or CD8<sup>+</sup> T cells isolated from a patient with at least one component selected from the group consisting of:

- (i) polypeptides comprising at least an immunogenic portion of a breast tumor protein, or a variant thereof, wherein the tumor protein comprises an amino acid sequence that is encoded by a polynucleotide sequence selected from the group consisting of:

- (1) sequences recited in SEQ ID NOs: 1-38, 42-204, 205, 207 and 210-290;

- (2) sequences that hybridize to a sequence recited in any one of SEQ ID NOs: 1-38, 42-204, 205, 207 and 210-290 under moderately stringent conditions; and

- (3) complements of sequences of (1) or (2);
  - (ii) polynucleotides encoding a polypeptide of (i); and
  - (iii) antigen presenting cells that express a polypeptide of (i);

such that T cells proliferate;

- (b) cloning at least one proliferated cell to provide cloned T cells;
- and

- (c) administering to the patient an effective amount of the cloned T cells, and thereby inhibiting the development of a cancer in the patient.



39. A method for determining the presence or absence of a cancer in a patient, comprising the steps of:

(a) contacting a biological sample obtained from a patient with a binding agent that binds to a breast tumor protein, wherein the tumor protein comprises an amino acid sequence that is encoded by a polynucleotide sequence recited in any one of SEQ ID NOs: 1-38, 42-204, 205, 207 and 210-290 or a complement of any of the foregoing polynucleotide sequences;

(b) detecting in the sample an amount of polypeptide that binds to the binding agent; and

(c) comparing the amount of polypeptide to a predetermined cut-off value, and therefrom determining the presence or absence of a cancer in the patient.

40. A method according to claim 39, wherein the binding agent is an antibody.

41. A method according to claim 40, wherein the antibody is a monoclonal antibody.

42. A method according to claim 40, wherein the cancer is breast cancer.

43. A method for monitoring the progression of a cancer in a patient, comprising the steps of:

(a) contacting a biological sample obtained from a patient at a first point in time with a binding agent that binds to a breast tumor protein, wherein the tumor protein comprises an amino acid sequence that is encoded by a polynucleotide sequence recited in any one of SEQ ID NOs: 1-38, 42-204, 205, 207 and 210-290 or a complement of any of the foregoing polynucleotide sequences;

(b) detecting in the sample an amount of polypeptide that binds to the binding agent;

(c) repeating steps (a) and (b) using a biological sample obtained from the patient at a subsequent point in time; and

(d) comparing the amount of polypeptide detected in step (c) to the amount detected in step (b) and therefrom monitoring the progression of the cancer in the patient.

44. A method according to claim 43, wherein the binding agent is an antibody.

45. A method according to claim 44, wherein the antibody is a monoclonal antibody.

46. A method according to claim 43, wherein the cancer is a breast cancer.

47. A method for determining the presence or absence of a cancer in a patient, comprising the steps of:

(a) contacting a biological sample obtained from a patient with an oligonucleotide that hybridizes to a polynucleotide that encodes a breast tumor protein, wherein the tumor protein comprises an amino acid sequence that is encoded by a polynucleotide sequence recited in any one of SEQ ID NO: 1-38, 42-204, 205, 207 and 210-290 or a complement of any of the foregoing polynucleotide sequences;

(b) detecting in the sample an amount of a polynucleotide that hybridizes to the oligonucleotide; and

(c) comparing the amount of polynucleotide that hybridizes to the oligonucleotide to a predetermined cut-off value, and therefrom determining the presence or absence of a cancer in the patient.

48. A method according to claim 47, wherein the amount of polynucleotide that hybridizes to the oligonucleotide is determined using a polymerase chain reaction.

49. A method according to claim 47, wherein the amount of polynucleotide that hybridizes to the oligonucleotide is determined using a hybridization assay.

50. A method for monitoring the progression of a cancer in a patient, comprising the steps of:

(a) contacting a biological sample obtained from a patient with an oligonucleotide that hybridizes to a polynucleotide that encodes a breast tumor protein, wherein the tumor protein comprises an amino acid sequence that is encoded by a polynucleotide sequence recited in any one of SEQ ID NO: 1-38, 42-204, 205, 207 and 210-290 or a complement of any of the foregoing polynucleotide sequences;

(b) detecting in the sample an amount of a polynucleotide that hybridizes to the oligonucleotide;

(c) repeating steps (a) and (b) using a biological sample obtained from the patient at a subsequent point in time; and

(d) comparing the amount of polynucleotide detected in step (c) to the amount detected in step (b) and therefrom monitoring the progression of the cancer in the patient.

51. A method according to claim 50, wherein the amount of polynucleotide that hybridizes to the oligonucleotide is determined using a polymerase chain reaction.

52. A method according to claim 50, wherein the amount of polynucleotide that hybridizes to the oligonucleotide is determined using a hybridization assay.

53. A diagnostic kit, comprising:

(a) one or more antibodies according to claim 10; and

(b) a detection reagent comprising a reporter group.

54. A kit according to claim 53, wherein the antibodies are immobilized on a solid support.

55. A kit according to claim 53, wherein the detection reagent comprises an anti-immunoglobulin, protein G, protein A or lectin.

56. A kit according to claim 53, wherein the reporter group is selected from the group consisting of radioisotopes, fluorescent groups, luminescent groups, enzymes, biotin and dye particles.

57. An oligonucleotide comprising 10 to 40 contiguous nucleotides that hybridize under moderately stringent conditions to a polynucleotide that encodes a breast tumor protein, wherein the tumor protein comprises an amino acid sequence that is encoded by a polynucleotide sequence recited in any one of SEQ ID NOs: 2-5, 7, 10, 13, 15, 16, 21, 22, 25, 28, 30, 33, 35, 63, 72, 73, 81, 103, 104, 107, 109, 118, 128, 129, 134, 136, 159-161, 164, 181, 183-204 and 210-290 or a complement of any of the foregoing polynucleotides.

58. A oligonucleotide according to claim 57, wherein the oligonucleotide comprises 10-40 contiguous nucleotides recited in any one of SEQ ID NOs: 2-5, 7, 10, 13, 15, 16, 21, 22, 25, 28, 30, 33, 35, 63, 72, 73, 81, 103, 104, 107, 109, 118, 128, 129, 134, 136, 159-161, 164, 181, 183-204 and 210-290.

59. A diagnostic kit, comprising:  
(a) an oligonucleotide according to claim 58; and  
(b) a diagnostic reagent for use in a polymerase chain reaction or hybridization assay.

## SEQUENCE LISTING

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<120> COMPOSITIONS AND METHODS FOR THERAPY AND  
 DIAGNOSIS OF BREAST CANCER

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 agacccatct ttggaaatga ttcccaaatt aganaacccat caggtctcaa aaaaggaagg 240  
 gtcacaaag tccatccagc ccagccaccc tgaggngcct gtatctctc aacaagccca 300  
 acacaatg 308

<210> 11  
 <211> 510  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(510)  
 <223> n = A,T,C or G

<400> 11  
 attatatgaa tatttttaatg caaaatgctt aacacttaaa attagcaaag cgtcatttaa 60  
 attaaaattc catttaacta agatgggtta accccaanaa attgtacagt agttgatttc 120  
 tgctatataa tgccagtcct atgccatata ataagaactg caacattagc tgctacttcc 180  
 tccattgctc ttctggacc ctaaggatga gggaggggac tcagacacaa aacacaaccc 240  
 aaataaactg tgcagtgtt cctaatagtt ataaaccaa tctaagttgt ccaaacagct 300  
 gaagaataac tgcaggtatt gttccanagc tgatacgagg ttttgctttt acagcctggt 360  
 aaaagtcttg cactaggtga gaagtcacag tttaaggatg catgttctgt aaatagttac 420  
 tacatatata catttactgt ctgtaaacac tagaaatata cattagacag agtaccctca 480  
 caagttgggt acagtttaaa aaagaagatg 510

<210> 12  
 <211> 611  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(611)  
 <223> n = A,T,C or G

<400> 12  
 agttttataa aatattttat ttacagtaga gctttacaaa aatagtctta aattaatata 60  
 aatccctttt gcaatataac ttatatgact atcttctcaa aaacgtgaca ttcgattata 120  
 acacataaac tacatttata gttgttaagt caccttgtag tataaatatg tttcatctt 180  
 ttttttgtaa taaggacat accaataaca atgaacaatg gacaacaaat cttattttgt 240  
 tattcttcca atgtaaaatt catctctggc caaaacaaa ttaaccaaag aaaagtaaaa 300  
 caattgtccc tctgttcaac aatacagtc tttttaatta tttgagagtt tatctgacag 360  
 agacacagca ttaactgaa agcaccatgg cataaagtct agtaacatta tcctcaaaag 420  
 ctttttccaa tgtctttcct tcaactgttt attcagtatt tggccagtag aaataaagat 480  
 tggctcaca tctctcttcc attagtctca agtggtccta ttatgcactg agttttcaga 540



ccttcccaac tggcatgtgt ttttaagtgtg agtttctttc tttggcttca agtggagttt 600  
 cacaacattt a 611

<210> 13  
 <211> 394  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(394)  
 <223> n = A,T,C or G

<400> 13  
 caatgttttag attcatttta ttagtggcat atacaaagca ccatataata tatgaaacgt 60  
 anaacaatca tgactatgta attaactgta naaataactg ctaanaaaat atagcaatat 120  
 ttaacacagg atttctaata ccattatatt ttcattactt ttcccaaagc taatgtccca 180  
 tgttttattt tatanacttt gtttatcaag atttatatgc atttggcacc tttttgggct 240  
 gaaaatagtt gatgtactct gtacagtaat gttacagttt tatacaaaaat tcanaaatat 300  
 tgcatttgga atagtcttta tggctcctct ccaagtattc agtttcacac aacagcaaac 360  
 actctgaatg cctttcctcc tgcccaacac aatg 394

<210> 14  
 <211> 361  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(361)  
 <223> n = A,T,C or G

<400> 14  
 agcaggnact ataattttat aattaatttt acaattcatg tagcaaatgg aaaatcatat 60  
 agagaggcca atgtatataa ataagagttt atacagaaac tgccaattca caaaacagca 120  
 ctgcatggtt tctatattgc aagcacaaga catggtcaca tggttccact gtacaggtag 180  
 aaacaagccc acagacaata catagagtac cacctgaaac gaggcccttg gagctgctca 240  
 gcttcttana aaataganaa ctttcaatgg tcataatata ttttgattca aaatgtcttc 300  
 taaaatgttt tcattgtggg agaaaattaa gaaggggcaa aaatccatct atggaacttc 360  
 t 361

<210> 15  
 <211> 537  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(537)  
 <223> n = A,T,C or G

<400> 15  
 acttacaaaa ttaattttat ttgcaaaaac tcaacaaata cacgttcaga tctggtttct 60  
 cttcaaaaaca tgtgtttgtt tttttaacaa acatgcaagt taatttggca tgccaaacat 120  
 ctttctctct agctgcctt ggaaaaattt ttttcataac acaaacaagg gtgcaaatat 180  
 tgtccaaacc tatttacatt ttaccctct agaattacat acattaatat ttattgggag 240  
 gaaagcaaaa ctgcaaaaaca tagtctttgg cattcacatt tgcttcagca gtataattaa 300  
 aaccttatat ttgttttaaa gataaacagt ttgaaggaaa ttaataaat cttgttttgg 360

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ctctgcaaag gagccactat atcaaagcat ttaactggag ctgttgagtt cctgctggta      420
gaatattact tccagcctat ttattagctt gtcttccggg ggcccaatac atgctttttt      480
ccctctacac tgaatgaaag tacaaaaaga aaaccatttc ttttcccaa cacaatg        537

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<210> 16
<211> 547
<212> DNA
<213> Homo sapien

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<220>
<221> misc_feature
<222> (1) ... (547)
<223> n = A,T,C or G

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<400> 16
gggtgtggng atgtatttat tcataatata ttttcagaac acattaataa tggagaataa      60
cacttattca tatactgaat ataacttttc ctggagcact ctagagcttg tttggagttg     120
gagaatactg ccaggctttt cctaactctt ttggtctttg gaagtgggca gggtttctca     180
aaccaagtgt ctccatggg ccattggcaa aggcttccct tcatcagctt ggaggggagcag    240
aaagaccatg gcttcagcac ttccattttg gaaagaagta acaaaaaagt gaattaatga     300
gcaatcggaa agactcaaag cattttgtac tccacagttc atttcttcac acaaacgtcc     360
attactgcag cgggcatgaa aaccggcagg gtgttaggct catggcctga agagaagtca     420
catcaccagc cgatgttttc atgcaaaagg caatcgtgat gattcanaac ctgggtctga     480
atttctccag gtgtgctcgt gagctgaagg tcatgcccat tctgtgcac cgtgcccacaa     540
cacaatg                                           547

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<210> 17
<211> 342
<212> DNA
<213> Homo sapien

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<400> 17
acattaagaa gctcctcttc tagcatgtcc ttaagaagcc tgtcttgag cactttcata      60
tcttctttca tcaaacacat ctcgatgta aaaacagttt ctccactatc agtattacag     120
aagacacttt tagccaatga agttttcaaa agaagaaagc ctctgttgtt cgtttttttg     180
atatgcactg aacttctgaa atatcttttc ccaaaagtcc acaaatcctt tttccaaatc     240
ttttaaagac tgtgaatctt tttcaaaatt ctccagctcc tctatgataa tgaattggaa     300
tttatcaagt tttttaatcc tagagtcctg actttggatg at                          342

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<210> 18
<211> 279
<212> DNA
<213> Homo sapien

```

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<400> 18
catcataagg ttttattcat atatatacag ggtattaaga attaagagga tgctgggctc      60
tgttcttggc ttggaagatt ctatttaatt gaaactctct gttcagaaag caataacttt     120
gtctcgttcc tgttgggctg aaccctaagg tgagtgtgca gtacagtgtg tgtgggtgaa     180
atggagattt ggaattgaac tctctgcctg taaatgttcc ccaataaatt gttgtgtgta     240
tgatacgtgt ataataaaag tattcttggt agaattctga                                279

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<210> 19
<211> 239
<212> DNA
<213> Homo sapien

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<400> 19
ctgccagcgt ttttgtgtgg ctgcagtgtg cctgggcccac gctcacgggc agtgggtgga      60

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cctaactgcc	caggcaggcg	agagctactt	ccagagcctt	ccagtgcattg	ggagggcagg	120
gctagggtga	gcgggtgtctc	ctctttgaaa	ttaagaacta	tctttcttgt	agcaaagctg	180
cacctgatga	tgctgcctct	cctctctgtg	ttgtctgggc	ccttgtttac	aagcacgcg	239

<210> 20  
 <211> 527  
 <212> DNA  
 <213> Homo sapien

<400> 20						
ctgaaccatt	atgggataaa	ctgggtgcaa	ttctttgcct	tctctacttc	tcaactgattg	60
aacataagct	tccagggtc	ccctgatgag	gaggagcctg	tccttttcag	atggatggtc	120
atccagccac	tgagagaagc	gtgtgtggga	ccactctgcc	ctctggaaag	gagatttcag	180
ttcagcgggt	gctctcgtga	acaaaaactg	aataatgatg	ctgaacggaa	tcacatcccc	240
caatgcagga	ctactggcta	catgttcact	tgcttggaag	agcagaggtc	tgaatgatct	300
cagcatccga	taggactttc	ctaaatcaga	tactcgtcta	cagaatgaac	ccacagccaa	360
ctccatctgt	gcaaaatcag	cagcaagtcg	cattttccca	ccttcaccaa	gaggtcttat	420
gagactggca	tggeggataa	aaagttcaac	agctctttgg	gcaataacct	cagtgttgtc	480
aaagacaaaa	tccaagcatt	caaagtgttt	aaaatagtca	ctcataa		527

<210> 21  
 <211> 399  
 <212> DNA  
 <213> Homo sapien

<400> 21						
ctgcaatggt	tgcaagtgtc	atttccacct	agctctgact	ctccacttct	aaccagacaa	60
acagccaacc	aaccaatcaa	catgtattta	ataaccacct	atgggggtgca	aagcacaaaa	120
gggcactcat	cttgaaaagg	aaagaccaag	aatgtgctag	agtaaaagaga	cagagaccag	180
accttactct	caagatcaag	agacttcagt	ctcggagaca	tctgccattt	ctctcttctt	240
aataaaacctc	atttgccctt	aaaaatatac	ttgctttggg	ggcccagaat	caagaaagga	300
aactttacaa	agtaaacaga	agttactccc	cacagggagg	cagaagcaga	ttaaccccaa	360
cagcagacat	ctgcccggaa	gagcaaaactc	cacatctgg			399

<210> 22  
 <211> 532  
 <212> DNA  
 <213> Homo sapien

<400> 22						
ccagaagggtg	aagaaaagtt	atctgataat	gtccaaagtg	cagtagaaat	acttttaacc	60
attgatgata	caaagagagc	tggaatgaaa	gagctaaaaac	gtcatcctct	cttcagtgat	120
gtggactggg	aaaatctgca	gcatacagact	atgcctttca	tccccagcc	agatgatgaa	180
acagatacct	cctattttga	agccaggaat	actgctcagc	acctgaccgt	atctggattt	240
agtctgtagc	acaaaaat	tccttttagt	ctagcctcgt	gttatagaat	gaacttgcac	300
aattatatac	tccttaatac	tagattgac	taagggggaa	agatcattat	ttaacctagt	360
tcaatgtgct	tttaatgtac	gttacagctt	tcacagagtt	aaaaggctga	aagggaatata	420
gtcagtaatt	tatcttaacc	tcaaaactgt	atataaatct	tcaaagcttt	tttcatctat	480
ttattttggt	tattgcactt	tatgaaaact	gaagcatcaa	taaaattaga	gg	532

<210> 23  
 <211> 215  
 <212> DNA  
 <213> Homo sapien

<400> 23						
tgcaaaataag	ggctgctgtt	tcgacgacac	cgttcgtggg	gtcccctggg	gcttctatcc	60
taataccatc	gacgtccctc	cagaagagga	gtgtgaattt	tagacacttc	tcaggggatc	120

tgccctgcac	ctgacacggg	gccgtcccca	gcacgggtgat	tagtcccaga	gctcgggctgc	180
cacctccacc	ggacacctca	gacacgcttc	tgacag			215

<210> 24  
 <211> 215  
 <212> DNA  
 <213> Homo sapien

<400> 24						
cctgaggctc	caggctaaga	agtagccaag	tttcacctgg	agagaagagt	agaggggactt	60
cccaaatttc	tctctgaact	cagctctgat	actcagaagg	tcagtctcac	atcgagagat	120
aaggatgcga	atcaggactt	ggtaattggg	ctcagtttcc	tagtagggga	agaaagagat	180
ggggggtagt	tagtgagagt	ctcactgaga	gtagg			215

<210> 25  
 <211> 530  
 <212> DNA  
 <213> Homo sapien

<400> 25						
ttttttttct	agtaagacta	gatttattca	ataccctagt	aaaagttttg	attataagta	60
tccaacagta	taaaaagtac	aaaacagatc	tgtagatttc	taatataatta	atacaaagtg	120
catgactaca	tacagtacat	cctacaggca	aagagagggtg	gaaggggaaa	aagaagactg	180
tggttgaggt	ctagtaataa	ataaataaat	acagaagtag	agatgatcca	tattatagta	240
tattctacca	ccaatactgc	agccaaaatg	tacaaaaaaa	atcatttcaa	ataactcagg	300
aggatgataa	tggtctggact	tttgtaattc	acctcaaaga	ctgtggggaga	gccaaactcaa	360
ctcactgtat	agtctgtgca	tatgggtggct	tgtagcatgt	aggttttttc	caaaagaagg	420
aaatataaaa	tgtttagatt	aagaactata	aaactacagg	gtgcctataa	aagggtggctt	480
actccttatt	gttattatac	tatccaattt	ttaaaatgca	gtttaaaaaa		530

<210> 26  
 <211> 366  
 <212> DNA  
 <213> Homo sapien

<400> 26						
ccagcagttc	tcggacctcc	tctgggggca	gggagaggcc	attgggtcag	gggctggacc	60
caggaggagt	tggaatgggt	gaaagatggg	gagcaagttt	ttagggtaca	gggtgggcct	120
aagatgggtc	agtagacaga	tgggagcaca	gagcagggca	gggggtgagg	tcaagtgagg	180
gccacaggat	gtgctgaggg	ctcccaggga	gccctaccca	ggctcacgtc	ctcctggtca	240
ccacctgtac	tgtctggggg	ccacagggtg	tgggcgttgc	caggagcac	tgggagggcc	300
tcggtagggg	ccacctgtag	ggagaggatg	tcaggaccac	tagcctctgg	gcaagggcag	360
aggagg						366

<210> 27  
 <211> 331  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(331)  
 <223> n = A,T,C or G

<400> 27						
ccaaactcag	agatgggtacc	agccaggggc	aagcatgacc	agagccaggg	accctgtggc	60
tctgatcccc	catttatcca	ccccatgtgc	ctcaggacta	gagtgagcaa	tcatacctta	120
taaaagactt	ttgtgccttt	ctgctccagt	ctcaaaattt	cctacacctg	ccagttcttt	180

acatTTTTcc	aaggaaagga	aaacggaagc	agggttcttg	cctggtagct	ccaggaccca	240
nctctgcagg	cacccaaga	ccctctgtgt	ccagcctctt	ccttgagttc	tcggaacctc	300
ctccctaatt	ctcccttctt	tccccacaag	g			331

<210> 28  
 <211> 530  
 <212> DNA  
 <213> Homo sapien

<400> 28						
ccatgaatgc	ccaacaagat	aatattctat	accagactgt	tacaggattg	aagaaagatt	60
tgtcaggagt	tcagaaggtc	cctgcactcc	tagaaaatca	agtggaggaa	aggacttggt	120
ctgattcaga	agatattgga	agctctgagt	gctctgacac	agattctgaa	gagcagggag	180
accatgccc	ccccagaaa	cacaccacgg	accctgacat	tgataaaaaa	gaaagaaaaa	240
agatgggtcaa	ggaagcccag	agagagaaaa	gaaaaaacia	aattcctaaa	catgtgaaaa	300
aaagaaagga	gaagacagcc	aagacgaaaa	aaggcaaata	gaatgagaac	catattatgt	360
acagtcattt	tcctcagttc	cttttctcgc	ctgaactctt	aagctgcatc	tggaagatgg	420
cttattgggt	ttaaccagat	tgatcatcgt	gcactgtctg	tgaagacgga	ttcaaagtgt	480
ttcatgtaac	tatgtaaaaa	gctctaagct	ctagagtcta	gatccagtca		530

<210> 29  
 <211> 571  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(571)  
 <223> n = A,T,C or G

<400> 29						
ccataatatt	ctgatgatca	aggagcacac	atatacaaaa	gttattggat	tactgcaatt	60
ctcagaggca	caaaacctga	catgggtgtg	tatagtatat	aatcagtcac	gggggggaaa	120
agaacattaa	gtctttaaaa	aggcttagga	agacataaac	agtaaattct	tgtttttcta	180
ccttcctttg	gacagtgtta	tatttcaatt	tcttctttgc	aaaatgtttc	caaattcatt	240
tgctcaggat	ttatttaaga	taataactta	aaacaactaa	cagttgttta	tgctatatgc	300
atatcatgca	tggttctactg	gttcaaggac	aaaattaaaa	caagatcttc	tctgtaaagc	360
aaatatattt	attatgcact	ttcatatata	cagggatttt	ttgagtacca	angggataaa	420
ataaaaactt	tacaatgtga	aattcaatgt	acatttttgg	ctatttacat	acctcaaacc	480
aagggaaaaa	taaaaagaaa	gcatttgttt	gcaactacat	ttgctgagaa	gtgtaaatgg	540
aggacattaa	gcaaaacaaa	tatttgcata	g			571

<210> 30  
 <211> 917  
 <212> DNA  
 <213> Homo sapien

<400> 30						
actgccagag	agtatgat	gaaggagatg	ggagcagatg	taattcttgg	ctggaatctc	60
tcatttcaaa	atcacttcac	ataatgggtg	catcatttaa	acacttaaca	gtcagtgcaa	120
ctgccactgt	aacatctagt	tgacacaaa	cacaaggagg	gggaggagaa	aatgccatca	180
ctattatggt	aacaaacatt	taattttaa	gggtgtctga	ctagtaaatt	tctgcagaaa	240
acagttttac	ccgccccctt	tcacagttcc	aaattaatca	aggatgcttt	tctataatct	300
gatgcttagc	aaattagctc	atgattcaaa	ttttgccctc	ttgaagcaca	tatacctttt	360
attttaaaag	tccattatag	agaatttgga	atatataagg	tatttgaatt	gcagaacacc	420
cctctaattc	tgtaaatata	gcaaagacaa	aacagtatca	tatacatcaa	gatcatactt	480
ttaaagtaag	tttaaaggtc	tcaattgccc	agatattaaa	tttatatttt	ccttctatta	540
aaaaatatta	catttcaatt	ttgtaattat	gtaacatatt	ttaagatgac	cagcaagacc	600

tagtcaat	ttt	gaaaata	cccc	ttgcatt	ccca	tacacaag	ct	ataccata	aag	taataac	ccca	660
agtatat	gat	gtgtaaa	aagt	tggtga	aggt	cataata	ctg	aatttttt	tg	caaata	gtaaa	720
ctgcttt	cca	agtaat	cagc	accatt	tttt	actagac	tac	atttta	atca	cttcct	tagc	780
tgcttaca	ac	ctctact	tag	gcataa	ataa	aagaat	ctga	aattgg	tata	ttcccc	cttc	840
ctgctgt	gtt	aacaaaa	aat	actatt	tgac	ttaaag	atca	aagagt	cttt	ttcctga	agg	900
ttttt	gtttt	taaat	gt									917

<210> 31  
 <211> 367  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(367)  
 <223> n = A,T,C or G

tctttt	cttt	ctgtatt	ttcc	caaatt	acag	ggagct	atgc	ccttgg	tatt	gcacac	agta	60
cactgca	aaaa	gattcaca	aag	gttagt	tgaa	agtcatt	tttt	gccctg	gtga	ttcaa	agctc	120
aaanaa	tttt	ctagcata	aaa	gtcttatt	aa	aat	ttta	aat	ttta	tttgag	ttta	180
agttta	ataa	aacaata	cca	ctatat	atac	tctcaac	aac	ttcatt	tata	aatcag	tcct	240
atgaggt	gtg	acttgct	ttt	catatc	acac	tgatta	agga	caaaa	ataat	tttgat	gtac	300
atgtacc	ata	cactgat	atg	caatct	acac	actgat	gcat	ttacata	cat	acaac	cccaa	360
cacaat	g											367

<210> 32  
 <211> 847  
 <212> DNA  
 <213> Homo sapien

cattgtg	ttg	ggctgg	cagg	atagaag	cag	cggctc	actt	ggactt	tttt	c	accaggg	aaa	60
tcagag	acaa	tgatgg	ggct	cttcccc	aga	actacag	ggg	ctctgg	ccat	cttcgt	ggta	120	
agtcct	ggat	tttcc	taata	atcacaa	act	tcctg	cttc	ctccct	gtt	aaaga	atatt	180	
atatttg	att	gcaca	atctt	tattata	aat	tctaaa	agga	gtgcag	tgga	aatca	acact	240	
ttgaaat	gaa	atcgtg	aaga	ttacca	attt	ccttct	tttg	ttgttt	ttta	tggtg	tattt	300	
tacata	gaaa	aataaa	accag	aaagaa	atga	gtttta	aaaa	ccattt	tagaa	tttttt	tttag	360	
ttaatg	aatt	aagta	atctt	aatcac	agg	tata	tttt	cca	at	ttt	cttt	420	
aaagtt	atgc	ttttact	agt	ttttcta	acc	cacaa	caa	aacac	agg	ag	ccactt	ctat	480
tttcca	agat	tacatg	tctc	ttagcat	ata	gctaag	aact	ctacac	gcct	gggctt	gata	540	
cctgac	acgc	ttttaaa	agt	aaaaaat	cgc	agaatt	aaaa	tcaaag	cagt	gtttg	actct	600	
agaga	agttg	ggagg	attat	taagta	agta	tttatg	ttta	gctatt	atgt	gccaaa	agaa	660	
aatgtc	agcc	tttggg	gatg	ggggg	aaaga	cataca	acat	tttaa	agcca	tttttt	tcag	720	
aaaag	taata	cttctg	ttga	ttgaga	aatg	cgtaca	atagt	attat	ctaaa	agagaa	acgg	780	
aatgtt	acag	actgtt	taaa	acctgg	atgt	tacaga	actaa	cttact	ccct	aactgt	gttc	840	
ttatag	c											847	

<210> 33  
 <211> 863  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(863)  
 <223> n = A,T,C or G

&lt;400&gt; 33

cattgtgttg	ggcttttatt	tgagtttatg	aacagaaata	gaaagtatgg	tgcttgggtt	60
ttgccctttc	ttactcctga	aagttaaatc	agaagacact	gatttcattt	tgtgaaattt	120
agctcagaga	ctattgatct	tttgtttcat	taatatgaac	aactattagt	aaaaaatagc	180
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aaggagggtg	tgatatattt	aggtgtaaat	atatcacata	tatggtgtga	tatatttggg	720
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&lt;210&gt; 34

&lt;211&gt; 432

&lt;212&gt; DNA

&lt;213&gt; Homo sapien

&lt;400&gt; 34

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&lt;210&gt; 35

&lt;211&gt; 350

&lt;212&gt; DNA

&lt;213&gt; Homo sapien

&lt;400&gt; 35

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&lt;210&gt; 36

&lt;211&gt; 1082

&lt;212&gt; DNA

&lt;213&gt; Homo sapien

&lt;400&gt; 36

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<210> 37  
 <211> 1135  
 <212> DNA  
 <213> Homo sapien

<400> 37						
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<210> 38  
 <211> 1323  
 <212> DNA  
 <213> Homo sapien

<400> 38						
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ggactggaac	acctgcggct	tgctcagtac	accatagagc	ggtatttcac	cttagtcacc	660
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aatgttctgt	atttcatttt	ggaaacctac	gttccttcca	cttctctgggt	ggtgttgtcc	780



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<210> 39  
 <211> 440  
 <212> PRT  
 <213> Homo sapien

<400> 39

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			20					25					30		
Ser	Asp	Lys	Leu	Ser	Leu	Pro	Gly	Phe	Glu	Asn	Leu	Thr	Ala	Gly	Tyr
		35					40					45			
Asn	Lys	Phe	Leu	Arg	Pro	Asn	Phe	Gly	Gly	Glu	Pro	Val	Gln	Ile	Ala
50						55					60				
Leu	Thr	Leu	Asp	Ile	Ala	Ser	Ile	Ser	Ser	Ile	Ser	Glu	Ser	Asn	Met
65					70					75					80
Asp	Tyr	Thr	Ala	Thr	Ile	Tyr	Leu	Arg	Gln	Arg	Trp	Met	Asp	Gln	Arg
			85						90					95	
Leu	Val	Phe	Glu	Gly	Asn	Lys	Ser	Phe	Thr	Leu	Asp	Ala	Arg	Leu	Val
			100					105					110		
Glu	Phe	Leu	Trp	Val	Pro	Asp	Thr	Tyr	Ile	Val	Glu	Ser	Lys	Lys	Ser
			115				120					125			
Phe	Leu	His	Glu	Val	Thr	Val	Gly	Asn	Arg	Leu	Ile	Arg	Leu	Phe	Ser
130						135					140				
Asn	Gly	Thr	Val	Leu	Tyr	Ala	Leu	Arg	Ile	Thr	Thr	Thr	Val	Ala	Cys
145				150					155					160	
Asn	Met	Asp	Leu	Ser	Lys	Tyr	Pro	Met	Asp	Thr	Gln	Thr	Cys	Lys	Leu
			165						170					175	
Gln	Leu	Glu	Ser	Trp	Gly	Tyr	Asp	Gly	Asn	Asp	Val	Glu	Phe	Thr	Trp
			180					185					190		
Leu	Arg	Gly	Asn	Asp	Ser	Val	Arg	Gly	Leu	Glu	His	Leu	Arg	Leu	Ala
		195					200					205			
Gln	Tyr	Thr	Ile	Glu	Arg	Tyr	Phe	Thr	Leu	Val	Thr	Arg	Ser	Gln	Gln
210						215					220				
Glu	Thr	Gly	Asn	Tyr	Thr	Arg	Leu	Val	Leu	Gln	Phe	Glu	Leu	Arg	Arg
225					230					235				240	
Asn	Val	Leu	Tyr	Phe	Ile	Leu	Glu	Thr	Tyr	Val	Pro	Ser	Thr	Phe	Leu
			245						250					255	
Val	Val	Leu	Ser	Trp	Val	Ser	Phe	Trp	Ile	Ser	Leu	Asp	Ser	Val	Pro
		260						265					270		
Ala	Arg	Thr	Cys	Ile	Gly	Val	Thr	Thr	Val	Leu	Ser	Met	Thr	Thr	Leu
		275					280					285			
Met	Ile	Gly	Ser	Arg	Thr	Ser	Leu	Pro	Asn	Thr	Asn	Cys	Phe	Ile	Lys
290						295					300				
Ala	Ile	Asp	Val	Tyr	Leu	Gly	Ile	Cys	Phe	Ser	Phe	Val	Phe	Gly	Ala
305					310					315				320	
Leu	Leu	Glu	Tyr	Ala	Val	Ala	His	Tyr	Ser	Ser	Leu	Gln	Gln	Met	Ala

325 330 335  
 Ala Lys Asp Arg Gly Thr Thr Lys Glu Val Glu Glu Val Ser Ile Thr  
 340 345 350  
 Asn Ile Ile Asn Ser Ser Ile Ser Ser Phe Lys Arg Lys Ile Ser Phe  
 355 360 365  
 Ala Ser Ile Glu Ile Ser Ser Asp Asn Val Asp Tyr Ser Asp Leu Thr  
 370 375 380  
 Met Lys Thr Ser Asp Lys Phe Lys Phe Val Phe Arg Glu Lys Met Gly  
 385 390 395 400  
 Arg Ile Val Asp Tyr Phe Thr Ile Gln Asn Pro Ser Asn Val Asp His  
 405 410 415  
 Tyr Ser Lys Leu Leu Phe Pro Leu Ile Phe Met Leu Ala Asn Val Phe  
 420 425 430  
 Tyr Trp Ala Tyr Tyr Met Tyr Phe  
 435 440

<210> 40  
 <211> 289  
 <212> PRT  
 <213> Homo sapien

<400> 40  
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 Ser Asp Lys Leu Ser Leu Pro Gly Phe Glu Asn Leu Thr Ala Gly Tyr  
 35 40 45  
 Asn Lys Phe Leu Arg Pro Asn Phe Gly Gly Glu Pro Val Gln Ile Ala  
 50 55 60  
 Leu Thr Leu Asp Ile Ala Ser Ile Ser Ser Ile Ser Glu Ser Asn Met  
 65 70 75 80  
 Asp Tyr Thr Ala Thr Ile Tyr Leu Arg Gln Arg Trp Met Asp Gln Arg  
 85 90 95  
 Leu Val Phe Glu Gly Asn Lys Ser Phe Thr Leu Asp Ala Arg Leu Val  
 100 105 110  
 Glu Phe Leu Trp Val Pro Asp Thr Tyr Ile Val Glu Ser Lys Lys Ser  
 115 120 125  
 Phe Leu His Glu Val Thr Val Gly Asn Arg Leu Ile Arg Leu Phe Ser  
 130 135 140  
 Asn Gly Thr Val Leu Tyr Ala Leu Arg Ile Thr Thr Thr Val Ala Cys  
 145 150 155 160  
 Asn Met Asp Leu Ser Lys Tyr Pro Met Asp Thr Gln Thr Cys Lys Leu  
 165 170 175  
 Gln Leu Glu Ser Trp Gly Tyr Asp Gly Asn Asp Val Glu Phe Thr Trp  
 180 185 190  
 Leu Arg Gly Asn Asp Ser Val Arg Gly Leu Glu His Leu Arg Leu Ala  
 195 200 205  
 Gln Tyr Thr Ile Glu Arg Tyr Phe Thr Leu Val Thr Arg Ser Gln Gln  
 210 215 220  
 Glu Thr Gly Asn Tyr Thr Arg Leu Val Leu Gln Phe Glu Leu Arg Arg  
 225 230 235 240  
 Asn Val Leu Tyr Phe Ile Leu Glu Thr Tyr Val Pro Ser Thr Phe Leu  
 245 250 255  
 Val Val Leu Ser Trp Val Ser Phe Trp Ile Ser Leu Asp Ser Val Pro  
 260 265 270  
 Ala Arg Thr Arg Ile Gly Asp Asn Lys Gly Ser Arg Arg Ser Gln Tyr  
 275 280 285

Tyr

<210> 41  
 <211> 265  
 <212> PRT  
 <213> Homo sapien

<400> 41  
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 Ser Asp Lys Leu Ser Leu Pro Gly Phe Glu Asn Leu Thr Ala Gly Tyr  
 35 40 45  
 Asn Lys Phe Leu Arg Pro Asn Phe Gly Gly Glu Pro Val Gln Ile Ala  
 50 55 60  
 Leu Thr Leu Asp Ile Ala Ser Ile Ser Ser Ile Ser Glu Ser Asn Met  
 65 70 75 80  
 Asp Tyr Thr Ala Thr Ile Tyr Leu Arg Gln Arg Trp Met Asp Gln Arg  
 85 90 95  
 Leu Val Phe Glu Gly Asn Lys Ser Phe Thr Leu Asp Ala Arg Leu Val  
 100 105 110  
 Glu Phe Leu Trp Val Pro Asp Thr Tyr Ile Val Glu Ser Lys Lys Ser  
 115 120 125  
 Phe Leu His Glu Val Thr Val Gly Asn Arg Leu Ile Arg Leu Phe Ser  
 130 135 140  
 Asn Gly Thr Val Leu Tyr Ala Leu Arg Ile Thr Thr Thr Val Ala Cys  
 145 150 155 160  
 Asn Met Asp Leu Ser Lys Tyr Pro Met Asp Thr Gln Thr Cys Lys Leu  
 165 170 175  
 Gln Leu Glu Ser Trp Gly Tyr Asp Gly Asn Asp Val Glu Phe Thr Trp  
 180 185 190  
 Leu Arg Gly Asn Asp Ser Val Arg Gly Leu Glu His Leu Arg Leu Ala  
 195 200 205  
 Gln Tyr Thr Ile Glu Arg Tyr Phe Thr Leu Val Thr Arg Ser Gln Gln  
 210 215 220  
 Glu Thr Gly Asn Tyr Thr Arg Leu Val Leu Gln Phe Glu Leu Arg Arg  
 225 230 235 240  
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 260 265

<210> 42  
 <211> 574  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(574)  
 <223> n = A,T,C or G

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 aatttgggaa aacctatgat tacaagtaaa aactcagaaa tgcaaagatg ttggtttttt 180

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ctaaattgct tctatctagc atgttaaaca aagataatat actttcgatg aaagtaaatt      480
ataggaaaaa aattaactgt tttaaaaaga acttgattat gttttatgat ttcaggcaag      540
tattcatttt taacttgcta cctactttta aata                                     574

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<210> 43

<211> 467

<212> DNA

<213> Homo sapien

<220>

<221> misc\_feature

<222> (1)...(467)

<223> n = A,T,C or G

<400> 43

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tccgtagctg gtttctcacc ataccctgca gttctgtgag ccaaaggctt tgcagaaagt      180
taaaataaat cacaaagact gctgtcatat attaattgca taaacacctc aacattgtctc      240
anagtttcat ccgtttgggt aanaaaacat tctttcaatt catctatggc atttgtagtg      300
gcattgtcgt ctatgaactc ttgaagaagt tctttgtatt cagtcttaga cacttgtgga      360
ttgattgtct tggaaatcac attctccaat aaggggcagc cagagcctgc gtagcagtgc      420
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<210> 44

<211> 613

<212> DNA

<213> Homo sapien

<220>

<221> misc\_feature

<222> (1)...(613)

<223> n = A,T,C or G

<400> 44

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gcagccgggc cgattgcagg acgtggcctg tcgggccagg gtcgctgaca tgcacgctgg      180
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aaccctccga gcacaaccac cttaggccaa ctgaatgtaa tctagtttat tcaacaaaaa      360
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aagcatgtag gcctanaaaa aggctctctg aaaccctcaa tggcaactgg tgaacggtaa      600
cactgattgc cca                                     613

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<210> 45

<211> 334

<212> DNA

<213> Homo sapien

<220>

<221> misc\_feature

&lt;222&gt; (1)...(334)

&lt;223&gt; n = A,T,C or G

&lt;400&gt; 45

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cagtataatc aaaatcaatt gtatcatcat tagttttcca ctgcctcaca ctagtgagct	120
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ggaaagccaa atcccagatg agtctcagag agggatcaat atgtccatga ttatcaggta	240
tgctgactat ttccaagggg tttttcagtt gcttcatttg cttgtaaagc aggtaatcct	300
cttgttgtnt tttctttttc tcgatgagcc gtgt	334

&lt;210&gt; 46

&lt;211&gt; 429

&lt;212&gt; DNA

&lt;213&gt; Homo sapien

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; (1)...(429)

&lt;223&gt; n = A,T,C or G

&lt;400&gt; 46

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taatagactt aaacatataa tgatggctaa aaaaaataag tatacgaaaa tgtaaaaaag	180
gaaatgtaag tccactctca atctcataaa aggtgagagt aaggatgcta aagcaaaata	240
aatgtagggt ctttttttct atttccgttt atcatgcagt ctgcttcttt gatatgcctt	300
agggttaccc atttaagtta gaggttgtaa tgcaatgggt ggaaatgaaaa ttgatcaaat	360
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gaagaaaaa	429

&lt;210&gt; 47

&lt;211&gt; 394

&lt;212&gt; DNA

&lt;213&gt; Homo sapien

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; (1)...(394)

&lt;223&gt; n = A,T,C or G

&lt;400&gt; 47

acgcgaantt gtgttatgac tgatagcctt cagctacaaa angataggac tgacctggtt	60
taaagtgttc tattttgtaa atcattccat ttgagtcttt ctgatgaact tggctatact	120
gaaatctgtt atttttagtga ggctccaaaa tgagcaaagc taggcctgat tagagtagag	180
tgactattaa aaaacataac tttctaggag ctataaatca aagttttaaa aagatgtttg	240
gatatatttg agtattccga tcatgaaaac agaaattgcc ctgcctacta caaggacaga	300
ctgatgggaa attatgcacc tggtaactt agcttttaag cagacgatgc tgtaaaaaaca	360
aacggcttct ctgatattta ttgtaagttt tagt	394

&lt;210&gt; 48

&lt;211&gt; 486

&lt;212&gt; DNA

&lt;213&gt; Homo sapien

&lt;400&gt; 48

acaaaggaac cgaggggtga ccacctctga gatgtccttg actttgtcat agcctggggc	60
atattgagca tctctctcac agctgccttt cttatcccca ttcttgatgt agacctcctt	120

ccgagtcagc	tttttctcct	cctcagacac	aaacagagct	ttgatatcct	gtgcagggag	180
cagctcttcc	ttttgttgct	ggcaagtggg	agttggagga	agcctcaaag	ctcgagttgt	240
tccttcgggt	caggggagac	aaatgggcct	gatagtctgg	ccatatattca	gcttattctt	300
gagcttgatc	agggcaacgt	catagtcata	aaattcagga	attcctgctt	cttttttccc	360
attaatgttg	tagttggggg	gaaataggac	tacttctatc	tccagggtccc	gcttctcccc	420
tcctttgatt	gagtgttcct	tgatcatccac	agtgaacaaa	tgtgctgctg	tcagcacaaa	480
gtacct						486

<210> 49  
 <211> 487  
 <212> DNA  
 <213> Homo sapien

<400> 49						
acgggctgac	agagaagatt	cccagagagta	aatcatcttt	ccaatccaga	ggaacaagca	60
tgtctctctg	ccaagatcca	tctaaactgg	agtgatgtta	gcagacccag	cttagagttc	120
ttctttcttt	cttaagccct	ttgctctgga	ggaagttctc	cagcttcagc	tcaactcaca	180
gcttctccaa	gcataccct	gggagtttcc	tgagggtttt	ctcataaatg	agggtgcac	240
attgcctggt	ctgcttcgaa	gtattcaata	ccgctcagta	ttttaaatga	agtgattcta	300
agatttggtt	tgggatcaat	aggaaagcat	atgcagccaa	ccaagatgca	aatgttttga	360
aatgatatga	ccaaaatttt	aagtaggaaa	gtcacccaaa	cacttctgct	ttcacttaag	420
tgtctggccc	gcaatactgt	aggaacaagc	atgatcttgt	tactgtgata	ttttaaatat	480
ccacagt						487

<210> 50  
 <211> 460  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1) ... (460)  
 <223> n = A,T,C or G

<400> 50						
acatatatttg	gttgaagaca	ccagactgaa	gtaaacagct	gtgcatccaa	tttatttatag	60
ttttgtaagt	aacaatatgt	aatcaaactt	ctaggtgact	tgagagtgga	acctcctata	120
tcattatttta	gcaccgttta	tgacagtaac	catttccagt	tattgtttat	tataccactt	180
atatcaactt	atttttcacc	aggttaaaat	tttaatttct	acaaaataac	attctgaatc	240
aagcacactg	tatgttcagt	aggttgaact	atgaacactg	tcataaatgt	tcagttcaaa	300
agcctgaaaag	tttagatcta	gaagctggta	aaaatgacaa	tatcaatcac	attaggggaa	360
ccattgttgt	cttcacttaa	tccatttagc	actattgaaa	ataagcacac	caagntatat	420
gactaatata	acttgaaaat	tttttatact	gagggggtng			460

<210> 51  
 <211> 529  
 <212> DNA  
 <213> Homo sapien

<400> 51						
acacttgaaa	ccaaatttct	aaaacttggt	tttcttaaaa	aatagttggt	gtaacattaa	60
accataacct	aatcagtggt	ttcactatgc	ttccacacta	gccagtcttc	tcacacttct	120
tctggtttca	agtctcaagg	cctgacagac	agaagggtct	ggagattttt	tttctttaca	180
attcagttct	cagcaacttg	agagctttct	tcatgttgct	aagcaacaga	gctgtatctg	240
caggttcgta	agcatagaga	cggtttgaat	atcttccagt	gatatcggct	ctaactgtca	300
gagatgggtc	aacaaacata	atcctgggga	catactggcc	atcaggagaa	agggtgttgt	360
cagttgttct	ataaaccaga	ttgaggagga	caaactgtct	tgccaatttc	tggatttctt	420
tattttcagc	aaacactttc	tttaaagctt	gactgtgtgg	gcactcatcc	aagtgatgaa	480

taaatcatca aggggttgggt gcttgtcttg gatttatata gagcttctt 529

<210> 52  
 <211> 379  
 <212> DNA  
 <213> Homo sapien

<400> 52  
 acttttgcaa gcagtaaagg atccaggaga tagcactgga tgtggtgtca tgcctgcaa 60  
 acatgaacgt ttccacttca gcctggagat ctgcttcaga gaaatctttg gtgttttcgc 120  
 ttttggcact caaaagtatg tccagaaaat cccagcgctt tttctgagta gtatcttggt 180  
 ttagcttata cttaagagac tccttcgggt cctggattac tttctctgtg aactgatgaa 240  
 gttcttggtt aaatttagaa aagatttggc cttgagagct gaatttgaaa accaggtcgt 300  
 tgtgatgtag aaaattgttc atgcgctggg tggagatttt gctaagggtg aacactgctt 360  
 tcaggtatga gtccagggt 379

<210> 53  
 <211> 380  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1) ... (380)  
 <223> n = A,T,C or G

<400> 53  
 acttttatct taaaagggtg gtagttttcc ctaaaatact tattatgtaa gggtcattag 60  
 acaaatgtct tgaagtagac atggaattta tgaatgggtc tttatcattt ctcttcccc 120  
 ttttggcat cctggcttgc ctccagtttt aggtccttta gtttgcttct gtaagcaacg 180  
 ggaacacctg ctgagggggc tctttccctc atgtatactt caagtaagat caagaatctt 240  
 ttgtgaaatt atagaaattn actatgtaaa tgcttgatgg aatnntttcc tgctagtgt 300  
 gcttctgaaa ggcgctttct ccatttattt aaaactaccc atgcaattaa aaggtacctt 360  
 gccgcgacca cncctaanggc 380

<210> 54  
 <211> 245  
 <212> DNA  
 <213> Homo sapien

<400> 54  
 gcgcggcgct tcacttcttc aacttccggg ccggtctgcc cagcgcgctg cgagtgtgtg 60  
 ccgaggtgca ggagggccgc gcgtggatta atccaaaaga gggatgtaaa gttcacgtgg 120  
 tcttcagcac agagcgctac aaccagaggt ctttacttca ggaaggtgag ggacgtttgg 180  
 ggaaatgttc tgctcgagtg tttttcaaga atcagaaaacc cagaccaacc atcaatgtaa 240  
 cttgt 245

<210> 55  
 <211> 556  
 <212> DNA  
 <213> Homo sapien

<400> 55  
 acagaagatg aataataatg aaaaactgtg attttttgac tatcacatac attgtgttaa 60  
 aaaacaggta aatataatga ctattactgt taagaaagac aaggaggaaa actgtttcaa 120  
 tgttcagggt taaatactaa gcacaaaaat ataacaaatt ctgtgtctac aataattttt 180  
 gaagtgtata caagtgcatt gcaaatgagc tctttaaaat ttaaagtcca ttccccctt 240  
 agccaagcat atgtctacat ttatgatttc tttctcttat tttaaagtct cttctgggtt 300

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agtttttttaa aaagtttcat catggetgtc atcttggaat ctagcctcca gctcaaagct 360
gagacttcac gcatacatat tctcctttct gggtgcatct tcacctagtt tctccaagta 420
ttcagagtta aatagcaciaa cttcttttat atgttcactt ttgtccacat gtagtggcag 480
tgctgctgct tcagtaggct ttctcacaca cccttttcct tctttcaaca gcagtcacca 540
aacgttcaca acacaa 556

```

```

<210> 56
<211> 166
<212> DNA
<213> Homo sapien

```

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<220>
<221> misc_feature
<222> (1) ... (166)
<223> n = A,T,C or G

```

```

<400> 56
atgggccctg attacatcat tatgaactac tcaggtnaac atcccaaata ccgacctngg 60
gaaagacttg gtccgagatg tggtcatcca tacaggctac ctcttcaga gncaggncc 120
caagagctgc ntnatcacct acctggccca ggtggacccc anaggg 166

```

```

<210> 57
<211> 475
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1) ... (475)
<223> n = A,T,C or G

```

```

<400> 57
acatccncat gttcctccaa atgacgtttg gggctctgct tgccaacatt ctttattgcc 60
agctgttcag gtgtcatctt atcttcttct tctacagcct tattgtaatt cttggctaatt 120
tccaacatct cttttaccac tgattcattg cgtttacaat gttcactgta gtccgaagt 180
gtcaaacctt ccatccaact cttcttatgc aaatttagca acatcttctg ttccagttca 240
ttttccgat agttaatagt aatggagtaa taatgtctgt ttagtccatg aattaatgcc 300
tggatagatg gcttgtttaa gtgaccaga ttccaagttg tttgtcttgg tcatgtcct 360
aagaccatca tattagcatt gatcaatctg aaggcatcaa taacaacctt tccttttaca 420
ctctgaatgg gatccacaac cactgccaca gntctctccg ataaggcttc aaage 475

```

```

<210> 58
<211> 520
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1) ... (520)
<223> n = A,T,C or G

```

```

<400> 58
actgttnatg tgctacttgc atttgtccct cttcctgtgc actaaagacc ccactcactt 60
ccctagtgtt cagcagtgga tgacctctag tcaagacctt tgcactagga tagttaatgt 120
gaaccatggc aactgatcac aacaatgtct ttcagatcag atccatttta tcctccttgt 180
tttacagcaa gggatattaa ttacctatgt tacctttccc tgggactatg aatgtgcaaa 240
attccaatgt tcatggtctc tccttttaa cctatattct acccctttta cattatagaa 300
aggaatgctg gaaacccaga gtccttctct tgggactctt aatgtgtatt tctaattatc 360

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catgactctt aatgtgcata ttttcaattg cctaattngat ttcaattgtc taagacattt 420  
caaatgtcta attggggaga actgagtcctt ttatatcaag ctaatatcta gcttttatat 480  
caagctaata tcttgacttc tcagcatcat agaagggggt 520

<210> 59  
<211> 214  
<212> DNA  
<213> Homo sapien

<220>  
<221> misc\_feature  
<222> (1)...(214)  
<223> n = A,T,C or G

<400> 59  
ctggcaggaa atgcatcaaa agacttaaag gtanagcgta ttaccctctg tcacttgcaa 60  
cttgctattc gtggagatga agaattggat tctctcatca aggctacaat tgctgggtggn 120  
gggtgcattc cacacatcca caaatctctg atngggaana aaggacaaca naagactgnc 180  
taanggatgc ctgnatncct tggaatctca tgac 214

<210> 60  
<211> 360  
<212> DNA  
<213> Homo sapien

<220>  
<221> misc\_feature  
<222> (1)...(360)  
<223> n = A,T,C or G

<400> 60  
gcatacaaca tggcagcagg gcctcgggaa gangggtagg aggaccgagc agcattctct 60  
gtagaggaag acaggaaagg agaccctctt ggcacacatt tatggagggt tgtccctgaa 120  
gagaagggca ggtgggagag gttccctgtt acttaagaga aggaccagt ggcaaagagc 180  
acaatgaaga ggatgatgat aaaaacaatc acgcagataa ggacaatcat cttcacgttc 240  
ttccaccaga attttcgagc caccttctgc gatgtcgtct tgaagtgtc agatgtggct 300  
tccagatcct ctgtcttggt gcggagatgt tccaagtttt cccccgggc caggatccgc 360

<210> 61  
<211> 391  
<212> DNA  
<213> Homo sapien

<220>  
<221> misc\_feature  
<222> (1)...(391)  
<223> n = A,T,C or G

<400> 61  
tntgggatcg tactcgatta aacagagcca cttttgttcc tgaggcaatg cataantcan 60  
catttttcaa tgactgttc tttttggaag gnttgagat gacttttatc cgcttgctga 120  
ggaacacacc aatgncatca ctgttgccat agaacatctt tacagacaac atgaantgct 180  
ttcgcttgtc tgagtcagat atatacaatg ttttggtgtg gcaatagtgc tttccttcca 240  
agtttagctg ctgcatttct tggncactat ttcctatccc aataaatgca cacggttgag 300  
actcttgntc agaacaacca tcncgttcca tttgttcttt ttttntcttc catccactgc 360  
ccataagata tacacannga ggtgggcaaa a 391

<210> 62

<211> 324  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(324)  
 <223> n = A,T,C or G

<400> 62  
 acaattttat tttaacagat ttcaagagtc catttttttaa aaaatgagca ataaagaacc 60  
 tctatcagtg agacttctca ttttatagca aatacatttt tgcagcttaa attttcttga 120  
 attcatatac gcttctgtca tttaaacaaa ctccagaga aaactggtct ctatataatt 180  
 aagtaacaaa ttgacaaaa tacatattta tacatatata ganctctaata ataaatatta 240  
 aatttgaaaa aatcaaagt gaagcagaaa ctgctataca agtatattgt ntaatatcta 300  
 tntnatacat taaagnnttc cggg 324

<210> 63  
 <211> 360  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(360)  
 <223> n = A,T,C or G

<400> 63  
 acaganncct tgaatatggt gtggttccct cattatggcc ctccattccc ttctgtgtta 60  
 atagtaaagc atgttgccca ataactacaa ccctgaccaa atttgggcct ggatctcatg 120  
 ggtaacgtgg agtttttaaat acgattttta atttacttgg gtaattgagc tgaatcttta 180  
 gttttcagat tactttttta aacagatagg ctcttagaac aaattattaa aaacataata 240  
 ccccatggga ggggaatctg gattaactac ccaactgttc cccccccc aacttttgaa 300  
 aaattttggc catatagaat gcatgaaaaa tcagggtatga tcttatgagg actttatagt 360

<210> 64  
 <211> 491  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(491)  
 <223> n = A,T,C or G

<400> 64  
 nctgactgtg atgtccactt gttccctgat ttttacacat catgtcaaag ataacagctg 60  
 ttcccaccca ccagttcctc taagcacata ctctgctttt ctgtcaacat cccatttttg 120  
 ggaaaggaaa agtcatattt attcccgcac cccagttttt taacttggtc tcccagttgt 180  
 cccctctctc tctgggtgta agaagggaag ttggaaaaaa attatatata tattctcctt 240  
 ttaatggtgg ggggctactg gagaggagag acagcaagtc caccctaact tgttacacag 300  
 cacataccac aggttctgga attctcatct tcgaacctag agaaataggt gctataaaca 360  
 gggaatttaag caaaatgctg gatgctatag atcttttaatt tgncttaatt tttttctat 420  
 tattaacta caggctgtag atntcttagg tctcacagaa cttntatcat tttaaactga 480  
 cttgtatatt t 491

<210> 65  
 <211> 484

<212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(484)  
 <223> n = A,T,C or G

<400> 65  
 accagcacac cggcgccgtc ctggactgcg ccttctacga tccaacgcat gcctggagtg 60  
 gaggactaga tcatcaattg aaaatgcatg atttgaacac tgatcaagaa aatcttgttg 120  
 ggacccatga tgccctatc agatgtgttg aatactgtcc agaagtgaat gtgatgggtca 180  
 ctggaagtgt ggatcagaca gctaaactgt gggatcccag aactccttgt aatgctggga 240  
 ccttctctca gcctgaaaag gtatataccc tctcagtgtc tggagaccgg ctgattgtgg 300  
 gaacagcagg ccgcagagng ttggtgtggg acttacggaa catgggttac gtgcagcagc 360  
 gcagggagtc cagcctgaaa taccagactc gctgcatacg agcgtttcca aacaagcagg 420  
 gttatgtatt aagctctatt gaaggccgag tggcagttga gtatttgac ccaagccctg 480  
 aggt 484

<210> 66  
 <211> 355  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(355)  
 <223> n = A,T,C or G

<400> 66  
 ngaagaaagt atgggtggag gtgaaggtaa tcacagagct gctgattctc aaaacagtgg 60  
 tgaaggaaat acaggtgctg cagaatcttc ttttctcag gaggtttcta gagaacaaca 120  
 gccatcatca gcatctgaaa gacaggcccc tcgagcacct cagtcaccga gacgcccacc 180  
 acatccactt cccccaagac tgaccattca tgccccacct caggagttag gaccaccagt 240  
 tcagagaatt cagatgaccc gaaggcagtc tgtaggacgt ggccttcagt tgactccagg 300  
 aataggtggc acgcaacagc atttttttga tgatgaagac agaacagttc caagt 355

<210> 67  
 <211> 417  
 <212> DNA  
 <213> Homo sapien

<400> 67  
 acgacacccc tcaagaggtg gccgaagctt tctgtcttc cctgacagag accatagaag 60  
 gagtcgatgc tgaggatggg cacagcccag gggaacaaca gaagcggaag atcgtcctgg 120  
 acccttcagg ctccatgaac atctacctgg tgctagatgg atcagacagc attggggcca 180  
 gcaacttcac aggagccaaa aagtgtctag tcaacttaat tgagaagggtg gcaagttagt 240  
 gtgtgaagtc aagatatggt ctagtgaacat atgccacata ccccaaaatt tgggtcaaag 300  
 tgtctgaagc agacagcagt aatgcagact gggtcacgaa gcagctcaat gaaatcaatt 360  
 atgaagacca caagttgaag tcagggacta acaccaagaa ggcctccag gcagtgt 417

<210> 68  
 <211> 223  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature

<222> (1)...(223)

<223> n = A,T,C or G

<400> 68

cacttgcaag	cttgcttaca	gagacctgnt	aaacaaagaa	cagacagatt	ctataaaatc	60
agttatatca	acatatataag	gagtgtgatt	ttcagtttgt	ttttttaagt	aaatatgacc	120
aaactgacta	aataagaagg	caaaacaaaa	aattatgctt	ccttgacaag	gcctttggag	180
taaacaaaat	gctttaaggc	tcctggtgaa	tgggggttgca	agg		223

<210> 69

<211> 396

<212> DNA

<213> Homo sapien

<400> 69

accttttttc	tctccaaagg	aacagtttct	aaagttttct	ggggggaaaa	aaaacttaca	60
tcaaatttaa	accatatggt	aaactgcata	ttagtttgt	tacaccaaaa	aattgcctca	120
gctgatctac	acaagtttca	aagtcattaa	tgcttgatat	aaatttactc	aacattaaat	180
tatcttaaat	tattaattaa	aaaaaaaaact	ttctaaggaa	aaataaacia	atgtagaccg	240
tgattatcaa	aggattatta	agaatcttt	acaaaaaatt	tcaaccctac	aacctaaac	300
cgcaaatttc	tattttttaa	catcagaaaa	taactcttgg	ttcattactt	atgacccaaa	360
gtttttattt	cactattcaa	tatctgaaaa	gtatca			396

<210> 70

<211> 402

<212> DNA

<213> Homo sapien

<220>

<221> misc\_feature

<222> (1)...(402)

<223> n = A,T,C or G

<400> 70

accannccc	accagggcaa	acagctccga	catgtttngt	aagtggagaca	agccagtgc	60
agttttttt	tttttttct	ttttctttt	tttgtcttt	gcttaccttc	ttgcttaatg	120
gaattgttat	ggctaagcac	atagaaggcc	aaaaaaggag	tttttcaaac	ccagcaaata	180
aagtgccttg	attctgaact	gccaaaagaa	aactgcactt	cccctcttaa	gtaaaacgaa	240
atgagtttct	taggtaaatg	tattcatcag	cccagataaa	aaaaaaacca	gttatgtgag	300
cgtagtcac	tgctcatttc	caggaanata	aaacaaaata	ccagcccagc	cagactcaca	360
tgtgggnata	tatatataaa	gcaagagagc	cacacccaca	ag		402

<210> 71

<211> 385

<212> DNA

<213> Homo sapien

<220>

<221> misc\_feature

<222> (1)...(385)

<223> n = A,T,C or G

<400> 71

accagtagag	agtgggccct	gcaggccact	tataaacagg	aagctctctc	ctgagctcac	60
tgatcaacct	gccttgggca	cagacagaac	ctaccagaaa	agaacaagta	caaaacacta	120
tcattatctg	ttttctcaag	acagtcccaa	atgtccttgt	gcgatcgcca	caaactcagt	180
gattggccca	agtcattccc	gggtgccata	aacagtaact	ggtgtgcanc	attagaacaa	240
ggggacacgg	ccttgattct	cttctgagca	acatgaactg	ggatttctgc	cnccccgagt	300

ctcggctgcc acctccgaag aagtcgtgac cagccacctc cacagtaaaa gattcctccc 360  
gtgagtatga ttggaatgc gncct 385

<210> 72  
<211> 538  
<212> DNA  
<213> Homo sapien

<220>  
<221> misc\_feature  
<222> (1)...(538)  
<223> n = A,T,C or G

<400> 72  
caattaatta acagaggtat aattgtctca ctttcagaag tgatcattta tttttattta 60  
gcacaggtca taagaaaaat atatagaaaa ataatacaatt tcatatataa aaggattatt 120  
tctccacctt taattattgg cctatcattt gttagtgtta tttggtcata ttattgaact 180  
aatgtattat tccattcaaa gtctttctag atttaaaaat gtatgcaaaa gcttaggatt 240  
atatcatgtg taactattat agataacatc ctaaaccctc agtttagata tataattgac 300  
tgggtgtaat ctcttttgta atctgntttg acagatttct taaattatgt tagcataatc 360  
aaggaagatt taccttgaag cactttccaa attgatactt tcaaacttat tttaaagcag 420  
tagaaccttt tctatgaact aagtcacatg caaaactcca acctgtaagt atacataaaa 480  
tggacttact tattcctctc accttctcca ggctaggaa tattcttctc tggagccc 538

<210> 73  
<211> 405  
<212> DNA  
<213> Homo sapien

<220>  
<221> misc\_feature  
<222> (1)...(405)  
<223> n = A,T,C or G

<400> 73  
actttatnna tggaattttc ttctacttgt atccatttnc cggggcttat ggaccattc 60  
atactctcca tatttagaat caaaggttcc tttctgaaga gaccttaatt ttaaggtaaa 120  
acgtgggtcca agttcctgaa ttcccacttt cttttcactc ctgaatatgt atctgtgaaa 180  
tctgaagaat atgtaatccc gttgattgtg gaatgtggca acctgccttc cgataaattg 240  
aggattatga ggaagagag atgcaaacat acgtccaatt gaatgaccca gccgtgttgt 300  
aaaattatc agaattattt caggtatgtg ttctgtgggg tccttgccctc ttctcttaat 360  
ttctttacga agacgaacac tgctcatttt aaaaatgagca gttgg 405

<210> 74  
<211> 498  
<212> DNA  
<213> Homo sapien

<220>  
<221> misc\_feature  
<222> (1)...(498)  
<223> n = A,T,C or G

<400> 74  
tgagccctgc acctgtttcc tgcaccccct gcenactggt tctatggcca caaggagttt 60  
taccagtaa aggagtttga ggtgtattat aagctgatgg aaaaataccc atgtgctgtt 120  
cccttgtggg ttggaccctt tacgatgttc ttcagtgtcc atgaccaga ctatgccaag 180  
atttctctga aaagacaaga tcccaaaagt gctgttagcc acaaaatcct tgaatcctgg 240

```

gttggctcgag gacttgtgac cctggatggt tctaaatgga aaaagcaccg ccagattgtg      300
aaacctggct tcaacatcag cattctgaaa atattcatca ccatgatgtc tgagagtgtt      360
cggatgatgc tgaacaaatg ggaggaacac attgcccaaa actcacgtct ggagctcttt      420
caacatgtct ccctgatgac cctggacagc atcatgaagt gtgccttcag ccaccagggc      480
agcatccagt tggacagt

```

```

<210> 75
<211> 458
<212> DNA
<213> Homo sapien

```

```

<400> 75
agccttgac atgatactca gattcctcac ccttgcttag gagtaaaaca atatacttta      60
caggggtgata ataatctcca tagttatttg aagtggcttg aaaaaggcaa gattgacttt      120
tatgacattg gataaaatct acaaatcagc cctcgagtta ttcaatgata actgacaaac      180
taaattatct ccctagaaag gaagatgaaa ggagtggagt gtggtttggc agaacaactg      240
catttcacag cttttccagt taaattggag cactgaacgt tcagatgcat accaaattat      300
gcatgggtcc taatcacaca tataaggctg gctaccagct ttgacacagc actgttcatc      360
tggccaaaca actgtgggta aaaacacatg taaaatgctt tttaacagct gatactgtat      420
aagacaaagc caagatgcaa aattaggctt tgattggc

```

```

<210> 76
<211> 340
<212> DNA
<213> Homo sapien

```

```

<220>
<221> misc_feature
<222> (1) ... (340)
<223> n = A,T,C or G

```

```

<400> 76
accttatacc aaaanaatgc ttattccaaa atattttttg tagctagtag ttctttcctt      60
ggaggtaaag aaaatacacc caaactttta attaccagga ttcagaatat ttaagagaac      120
aatttttagtt aagaatcaaa tatactgaga ttcaaaggagg ggaaaaaaag gaaatattat      180
agaagacaaa ggtcaaaactg gcattccaga tctggagcaa ttttgtaaag caggaaaaca      240
actatgacaa tctgnagctt cttagatcat tatagtgaat gtncccatct actataaggg      300
tttttataat ggtgtttcct aaataaagga acataaatgt

```

```

<210> 77
<211> 405
<212> DNA
<213> Homo sapien

```

```

<400> 77
actccatttg tggaaactcg gtccgagctc ggtaaacagc cgaatgtctt cctccccctac      60
agtttccctc ccttgcatga gagcagtgat gtctcgatta aaggcattaa ttttatctat      120
caggaagaac attttttcat tttcgtcttc cggatgtcg acaccatact tttgtagctc      180
ctctgttatt ctctggtgag tctccttgat ttgattttct aacaggggca gagatttaca      240
gatatgtgtg atgagctcgc tggtaagttt ttctgccagg cagggaaccg tggcctttcc      300
ttctccagc agatccctga aatatgggtg gtctcctcag aagatcttct ctctctgcag      360
ggcttcggac aggtccagct ggtcctggat ctctgctgg ccccg

```

```

<210> 78
<211> 410
<212> DNA
<213> Homo sapien

```

<220>  
 <221> misc\_feature  
 <222> (1)...(410)  
 <223> n = A,T,C or G

<400> 78  
 acagcagntn tagatggctg caacaacctt cctcctaccc cagcccagaa aatatttctg 60  
 cccaccccca ggatccggga ccaaaataaa gagcaagcag gccccttca ctgaggtgct 120  
 gggtagggct cagtgccaca ttactgtgct ttgagaaaga ggaaggggat ttgtttggca 180  
 ctttaaaaat agaggagtaa gcaaggactgg agaggccaga gaagatacca aaattggcag 240  
 ggagagacca tttggcgcca gtcccctagg agatgggagg agggagatag gtatgagggg 300  
 aggcgctaag aagagtagga ggggtccact ccaagtggca ggggtgctgaa atgggctagg 360  
 accaacagga cactgactct aggtttatga cctgtccata cccgttccac 410

<210> 79  
 <211> 512  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(512)  
 <223> n = A,T,C or G

<400> 79  
 acagtgaaaa acaaactaat ataaagcatt ccagnngata aaaacctcct caggcttatg 60  
 gtttgtttcc caaggaaatt atgtttcaat gtaaagtttg aaatactcca gacatacatt 120  
 ccatgtaggt tttgggtgcc aatgttaaaa tttcaattt tgcattgcaag gcttagcaaa 180  
 gaaacactgg cagaattcca gcatttgcaa aattctaagt tttggtgaat attgtaaata 240  
 ttacaattgg tattagaaag ccattgatgaa tccagaatta agagaaaacc catttcataa 300  
 atattttggt tgattaaaaa ataccaggct taccatgttc taaataacac aagaaaatat 360  
 ctttaaaaaa aaaaggactg caatttaaca gtaatctgta tatcttttagc tgccattaaa 420  
 aaaagaaaaa agaacaacca aaaacaatga aaatgttaca actggtataa agtnaccna 480  
 tgatgctccc cttacgagaa aacaaaactg tc 512

<210> 80  
 <211> 174  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(174)  
 <223> n = A,T,C or G

<400> 80  
 tgattcccca gacctcaaat gggctaacac gcttctcttc tncagcagnc ttctgtccg 60  
 tgaagntncc ttccagattg gtacatggaa ctgaaaacaa agggagcctc agctggattg 120  
 aaatctggag catgccacaa agncttgcaac tnggcatttt cnagaagaac ccat 174

<210> 81  
 <211> 274  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(274)

<223> n = A,T,C or G

<400> 81

ttgcaacaag	cacattaaat	taaggcctgc	tngaatttct	tcttcccca	tcaggtaa	60
tttctttgcc	aataaagttt	gaggaggtgg	catttgaaaa	tctcttttaa	aaagaagtct	120
tcatctattc	acnagaaaac	tcaaaaataa	ttttcattat	caacacacaa	actaactcaa	180
tctctgcttt	aagtttctat	tggccaattt	ttctgattna	tacgagaatt	attntcagnt	240
ntagaaaatc	ctgggtctttg	gtcattacaa	gntg			274

<210> 82

<211> 101

<212> DNA

<213> Homo sapien

<220>

<221> misc\_feature

<222> (1)...(101)

<223> n = A,T,C or G

<400> 82

atggagaaga	tgaacctga	gcctnntgag	aattgcctgc	taengcctgg	cagccctgcc	60
cgagtggccc	agcnncttt	cacnagntgg	gcattgattg	n		101

<210> 83

<211> 182

<212> DNA

<213> Homo sapien

<400> 83

tattatgggg	aaagataact	gagaataaag	ctatcatgca	gatatttgca	gagataaaag	60
taatgcagat	actgagtgg	gttttgatca	aactatgctt	gaaagccact	ctaccactag	120
ttacacaaac	caataatttc	ccttcgcagt	ggaagtcagc	ttgagttttt	tcagggtgtt	180
tt						182

<210> 84

<211> 229

<212> DNA

<213> Homo sapien

<220>

<221> misc\_feature

<222> (1)...(229)

<223> n = A,T,C or G

<400> 84

actgtttgta	gctgcactac	aacagattct	taccgtctcc	acaaaggtca	gagattgtaa	60
atgggtcaata	ctgacttttt	ttttattccc	ttgactcaag	acagctaact	tcattttcag	120
aactgtttta	aacctttgtg	tgctggttta	taaaataatg	tgngtaatcc	ttgttgcttt	180
cctgatacca	nactgtttcc	cgnggttggt	tagaatatat	tnngttcng		229

<210> 85

<211> 500

<212> DNA

<213> Homo sapien

<220>

<221> misc\_feature

<222> (1)...(500)



<223> n = A,T,C or G

<400> 85

ggggagtang	tgatttatta	aagcaagacg	ttgaaacctt	tacnttctgc	agtgaagatc	60
aggggtgcat	tgaaagacag	tggaaccag	gatgaaagt	tttacctg	acacactaca	120
tttcttcaat	atcttcacca	ggacttccgc	aatgaggctt	cgtttctgaa	gggacatctg	180
atccgagcat	ctcttcactc	ctaacttggc	tgcaacagct	tccagagggg	catcaaattt	240
ggcaagactt	aacttgaaca	gaggttccact	aatgaagaag	aagtctaaca	gctcagaaac	300
aagagctggg	cagaactcgg	cattggcctg	gtagcagcag	agggccagcg	tgaccagcag	360
gagacacacc	gacagcttca	tggtggcttg	ttttgctgtg	agctcagctt	tcacaacaaa	420
tgagtgattt	ggactccacc	ccaggagcct	gtggagctgc	agagcccagg	gctatttgta	480
cctgcccggg	cggncgctcg					500

<210> 86

<211> 323

<212> DNA

<213> Homo sapien

<220>

<221> misc\_feature

<222> (1)...(323)

<223> n = A,T,C or G

<400> 86

ccgccagtgt	gctggaatc	gcccttgccg	cccgggcagg	tactcagaag	tcatttggtta	60
tttacaattg	ggtttggtg	ggatgggatn	tanggcggat	gagccagtgc	ttttgcaatg	120
aagatgcaat	antcattgtc	ctctcccact	gtctcctctt	tcctcacccc	atggcagctn	180
tcattgacca	ttcccaaagg	gtccaccgag	tcctgaactc	agcttcatca	ccaacattcc	240
tcgccttcag	ttgaattcaa	cactgncaan	ggagnagang	caaagacttg	ggtcagggag	300
aggnggggaa	acacanaaca	aac				323

<210> 87

<211> 230

<212> DNA

<213> Homo sapien

<400> 87

gcagcattga	gccacccctt	tggcaggcga	tacggcagct	ctgtgccctt	ggccagcatg	60
tgagtgagg	gagatgctgc	ccctgtggtt	ggaacatcct	gggtgaccc	ccgaccagc	120
ctcgtgggc	tgccccctgt	ccctatctct	cactctggac	ccagggctga	catcctaata	180
aaataactgt	tggattagac	aaaaaaaaaa	aaaaaaaaaa	aaaaaaaaag		230

<210> 88

<211> 249

<212> DNA

<213> Homo sapien

<220>

<221> misc\_feature

<222> (1)...(249)

<223> n = A,T,C or G

<400> 88

atgtgaccag	gtctaggtct	ggagtttcag	nttgacact	gagccaagca	gacaagcaaa	60
gcaagccagg	acacaccatc	ctgcccagg	cccagcttct	ctcctgcctt	ccaacgccat	120
ggggagcaat	ctcagccccc	aactctgcct	gatgcccttt	atcttggggc	tcttgtctgg	180
aggtgtgacc	accactccnt	ggtctttggc	ccggccccat	ggatcctgct	ctctggaggg	240
ggtntagat						249

<210> 89  
 <211> 203  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(203)  
 <223> n = A,T,C or G

<400> 89  
 tggtttacact gtcaaggatg acaaggaaaag tggttctatc tntgatacca tcatcccagc 60  
 tggttctcct cccactgacc tgcgattcac caacattggt ccagacacca tgcgtgtcac 120  
 ctgggctcca ccccatcta ttgatttaac taacttcctg gtgcggnact cacctgtgaa 180  
 aaatgangaa gatgttgag agt 203

<210> 90  
 <211> 455  
 <212> DNA  
 <213> Homo sapien

<400> 90  
 ctctaagggg gctggcaaca tggctcagca ggcttgcccc agagccatgg caaagaatgg 60  
 acttgtaatt tgcacctctg tgatcacctt actcctggac cagaccacca gccacacatc 120  
 cagattaaaa gccaggaagc acagcaaacy tgcagtgaga gacaaggatg gagatctgaa 180  
 gactcaaatt gaaaagctct ggacagaagt caatgccttg aaggaaattc aagccctgca 240  
 gacagtctgt ctccgaggca cttaaagtca caagaaatgc taccttgctt cagaaggttt 300  
 gaagcatttc catgaggcca atgaagactg catttcctaaa ggaggaatcc tggttatccc 360  
 caggaaactcc gacgaaatca acgacctcca agactatggt aaaaggagcc tgccaggtgt 420  
 caatgacttt tggctgggca tcaatgacat ggtca 455

<210> 91  
 <211> 488  
 <212> DNA  
 <213> Homo sapien

<400> 91  
 actttgcttg ctcatatgca ttagtgcact ttataagtca ttgtatgtta ttatattccg 60  
 taggtagatg tgtaacctct tcaccttatt catggctgaa gtcacctctt gggtacagta 120  
 gcgtagcgtg gccgtgtgca tgccttttgc gcctgtgacc accaccccaa caaaccatcc 180  
 agtgacaaaac catccagtgg aggtttgtcg ggcaccagcc agcgtagcag ggtcgggaaa 240  
 ggccacctgt cccactccta cgatacgcta ctataaagag aagacgaaat agtgacataa 300  
 tatattctat ttttatactc ttctatcttt ttagtgacc tgtttatgag atgctggttt 360  
 tctacccaac ggcctgcag ccagctcacg tccaggttca accacagct acttggtttg 420  
 tgttcttctt catattctaa aaccattcca tttccaagca ctttcagtcc aataggtgta 480  
 ggaaatag 488

<210> 92  
 <211> 420  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(420)  
 <223> n = A,T,C or G

<400> 92  
 tctccggcag gctctgcccc ggctcgtagcn agnnaacct aatacctgac cttttttgta 60  
 gacaaccttg gtgctgaggt taactccatc cattgtagtg gctgtatat caatgggacg 120  
 attgcatatt tttcctgggt gagctttcca gaggtctgaa attttctccc cacttttagt 180  
 ctgagatact ttatcatgat cganccactc cgtccactcc acgtnttgaa cccactcact 240  
 ggacaaagaa acattgaaat attcgccatg ctctgtctgg aacaatttga ataccggggc 300  
 agcagcagag cctcgatgnc caggatattc aatatgggtc tccactgaag atgatggatt 360  
 tcctttcaca gntagaaaac ttncnagggg gtctaaatcc aaggtgcagg aagngngngc 420

<210> 93  
 <211> 241  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(241)  
 <223> n = A,T,C or G

<400> 93  
 accacgaatt ncaacatcca gatccaccac tatectaatt ggattgtaac tgngaactgt 60  
 gcccggctcc tgaaagccga ccaccatgca accaacgggg tgggtgcacct catcgataag 120  
 gtcattctcca ccatacacia caacatccag cagatcattg agatcganga cacctttgag 180  
 acccttcggg ctgctgnggc tgcatacagg ctcaacacga tgcttgaagg naacggncag 240  
 t 241

<210> 94  
 <211> 395  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(395)  
 <223> n = A,T,C or G

<400> 94  
 actctattnt aattctgctt ttttatactt aattctaaat ttttccctc taatttacaa 60  
 caaattttgt gatttttata agaattctatg cctccccaat tctcagattc ttctcttttc 120  
 tcctttattt ctttgcttaa attcagtata agctttcttg gtatttttagg cttcatgcac 180  
 attcttatcc ctaaacacca gcagttcttc agagacctaa aatccagtat aggaataact 240  
 gtgttagttc ttgaaaaagc attaaagaca tttttccctg aaacatacag aacatgtcat 300  
 gccaaatctc ttgtttacat aataaactgg taataccggg gaattgcaca tacagatttt 360  
 atctccaaga tagaataact taaatattaa aacgt 395

<210> 95  
 <211> 304  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(304)  
 <223> n = A,T,C or G

<400> 95  
 cgaggtagag tgatngctcc ccctgggcaa tacaatacaa gaacngnggg ttttgtcaaa 60  
 ttggaacaag gaaacagaac cacagaaata aatacattgg ttaacatcag attagttcag 120

gttacttttt	tgtaaaagt	aaagtacgag	gggacttctg	tattatgcta	actcaagtan	180
actggaatct	cctgttttct	ttttttttct	taaatngggt	ttaatttttt	ttaattggat	240
ctatcttctt	ccttaacatt	tcagttggag	tatgtagcat	ttagcaccac	tggctnaaac	300
ctgt						304

<210> 96  
 <211> 506  
 <212> DNA  
 <213> Homo sapien

<400> 96						
acactgtcag	cagggactgt	aaacacagac	aggggtcaaag	tgttttctct	gaacacattg	60
agttggaatc	actgttttaga	acacacacac	ttactttttc	tggtctctac	cactgctgat	120
atthttctta	ggaaatatac	ttttacaagt	aacaaaaata	aaaactctta	taaattttcta	180
tttttatctg	agttacagaa	atgattactg	aggaagatta	ctcagtaatt	tgtttaaaaa	240
gtaataaaat	tcaacaaaca	tttgctgaat	agctactata	tgtaagtgct	tggtcaaggt	300
attacactct	gtaattgaat	attattcctc	aaaaaattgc	acatagtaga	acgtatctg	360
ggaagctatt	tttttcagtt	ttgatatttc	tagcttatct	acttccaaac	taatttttat	420
ttttgctgag	actaatctta	atcattttct	ctaatatggc	aaccattata	accttaattt	480
attattaacc	ataccctaag	aagtag				506

<210> 97  
 <211> 241  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1) ... (241)  
 <223> n = A,T,C or G

<400> 97						
atthttcttt	taattacttt	agagagctag	ggatgcaa	gttttcagtt	agaaagcctt	60
tatttacttt	tggaattga	acaagaaatg	catctgtctt	agaaactgga	gattatttga	120
tgtaggttaa	aacatgta	atgntctctg	gcaattttgt	atcantnatt	ngaaaatgag	180
atattangaa	aaaccaattc	ttcttaaattc	tagnncatct	ttctttanaa	gaacattana	240
t						241

<210> 98  
 <211> 79  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1) ... (79)  
 <223> n = A,T,C or G

<400> 98						
ggcaaacana	cttatgctgn	ancnggggtt	tancaaggtt	ttcaaagnaa	aaanccatt	60
ngactttatg	gaaaatatt					79

<210> 99  
 <211> 316  
 <212> DNA  
 <213> Homo sapien

<220>

<221> misc\_feature  
 <222> (1)...(316)  
 <223> n = A,T,C or G

<400> 99  
 ccacatatgt aaaaccaga aagaccngnt tngcactttc actgagagtt gagtcatctg 60  
 ggctgtcnac aggtgtctga cgtgtaaact tggaaatcaaa ctgacttaca tcctcttcag 120  
 attgcaacag aggtttaaag gggggctcca ctttcgagc cagaagttct tcccagttaa 180  
 tgtgtctaaa gaatggatga gcttgaactt ctccagcgtc cccaggacca gctcccagac 240  
 gagaagcagc atttcttttc agcagctttt taagcagatc tctggcttct tgngtgaggt 300  
 agggaggcaa attgag 316

<210> 100  
 <211> 425  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(425)  
 <223> n = A,T,C or G

<400> 100  
 accgctttca gaaagtttat atgggttatt cttcagcctc tcttttatgc ctttcgacct 60  
 ctgtttatca accccaaacc aattacgtat ctggaagtta tcaataccgt ggcacaggtc 120  
 acttttgaca tttaattta ttactttttg ggaattaaat ccttagtcta catgttggca 180  
 gcatctttac ttggcctggg ttgacacca atttctggac attttatagc tgagcattac 240  
 atgttcttaa agggncatga aacttactca tattatgggc ctctgaattt acttaccttc 300  
 aatgtgggtt atcataatga acatcatgat ttcccaca ttcctggaaa aagtcttcca 360  
 ctggtgagga aaatagcagc tgaatactat gacaacctgc ctactacaa tttctggata 420  
 aaagg 425

<210> 101  
 <211> 156  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(156)  
 <223> n = A,T,C or G

<400> 101  
 actgacttgg gaatgtcaaa attctttatt atgatcttcc gagtgttgtc ctgagctttg 60  
 ttggccctca actgcaggca gagaaccagg agcagggtgg cagggtctggc cctgaacagg 120  
 agctggagca agcgcattgct ngagaaaaca gaaggc 156

<210> 102  
 <211> 230  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(230)  
 <223> n = A,T,C or G

<400> 102

```

actccaggcc gggntcagg ttatcaaaag tgcaggagct ctgatcagca tggaccactt      60
cttccaaaga atttcctgc tggccgtttg taggggttgt ggtaattcta taaccagtaa      120
tgtctggggt ggtgctcctc tcccaggaga ctgtgagcac tccagtgtca gggtttgcct      180
ccagatgcaa gntngtnggt ggagacaatg gtgncaccac tttgtnnaca                230

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<210> 103

<211> 404

<212> DNA

<213> Homo sapien

<220>

<221> misc\_feature

<222> (1) ... (404)

<223> n = A,T,C or G

<400> 103

```

actgtgaacc ctgnggnttc nangcgacct acctggagct ggccagtgt gtgaaggagc      60
agtatccggg catcgagatc gagtcgcgcc tcggggggcac aggtgccttt gagatagaga      120
taaattggaca gctggtgttc tccaagctgg agaattggggg ctttccttat gagaaagatc      180
tcattgaggc catccgaaga gccagtaatg gagaaaccct agaaaagatc accaacagcc      240
gtcctccctg cgtcatcctg tgactgcaca ggactctggg ttcttgcctt gttctggggg      300
ccaaaccttg gtctcccttt ggtcctgctg ggagctcccc ctgcctcttt cccctactta      360
gtcctcttagc aaagagaccc tggcctccac tttgcctctt gggg                    404

```

<210> 104

<211> 404

<212> DNA

<213> Homo sapien

<220>

<221> misc\_feature

<222> (1) ... (404)

<223> n = A,T,C or G

<400> 104

```

accaggttat ataatagtat aacactgcc aaggagcggat tatctcatct tcctcctgta      60
attccagtgt ttgtcacgtg gttgttgaat aaatgaataa agaattgagaa aaccagaagc      120
tctgatacat aatcataatg ataattatct caatgcacaa ctacgggttg tgctgaacta      180
gaatctatat tttctgaaac tggctcctct aggatctact aatgatttaa atctaaaaga      240
tgaagttagt aaagcatcag aaaaaaaagt gggatttctt acaagtcagg acattctacg      300
tgactataat ataatctcac agaaatttaa cattaatacn ttctaagatt taattcttag      360
antctnngta aacaaagtag ctctgtgga natgattggc atca                    404

```

<210> 105

<211> 325

<212> DNA

<213> Homo sapien

<220>

<221> misc\_feature

<222> (1) ... (325)

<223> n = A,T,C or G

<400> 105

```

acagcagaag ccagtctang atgggtgtgat tcaatttctg cctctagtat ttctttgtct      60
tgtttttcct tcaatttaga agtgagcatt gtgttctcag ctatcagaac tttaagctgc      120
ccactatatt gagatgccct tttagctaat gattcctctt tcagtttttag ggtcatctga      180
agttcagcat tctttttctt taaaatctta atgtcctcaa agtattttatt ttccttttcc      240

```

tggtattggn gtttcagngt ggctatttcc agtttttagca tggcaattnc ctttttcaac 300  
atgcaatttt catgtaagag ataata 325

<210> 106  
<211> 444  
<212> DNA  
<213> Homo sapien

<220>  
<221> misc\_feature  
<222> (1)...(444)  
<223> n = A,T,C or G

<400> 106  
actgtcttca atnctatgcg tgcaggtgtc taccacaggc aaacagtttt ctccccattt 60  
tgtagtaatg tgattttcct attagcaaaa agaggtcacc agcccctgta gacttaaggg 120  
actcaagtca caggatgggg atttcctctt aatatttttt atttngttgt ttgaactctt 180  
gatgcaacat tgtagagcag ggtgttcagg acctgctgtg cccaagggac tgataaagga 240  
aaaagctcta tttattcttt ttgtgatttg atgcacagat gaaaaactta acacacaata 300  
acagaagttg gncgttaata aatcacatcc taggctttca gcgcttncgt aagcagacga 360  
catcttcagt tttctagctc ttgnagnttc aacacngnaa catcaatgat gcatatgtnc 420  
agaatcagtt acaaagacca tccg 444

<210> 107  
<211> 287  
<212> DNA  
<213> Homo sapien

<220>  
<221> misc\_feature  
<222> (1)...(287)  
<223> n = A,T,C or G

<400> 107  
acctgcactc gnacntcagg cantaggcct ccacgtcatg gccaggcact ggcatgggct 60  
ccaccacgtg caggcagttg cagtccttct gggatacatt ctggttgtaa atgtgccac 120  
tgatgtttct ataagggtgg acagatgcat ttgcaccgga tatcttcana actctgttg 180  
gctncagctg ggggcaccaa caaacaccg accacagcca ccaaagataa nagcttcatg 240  
cttatcangc ttgctgggccc agnaaaagccg gacacctaca agcccn 287

<210> 108  
<211> 478  
<212> DNA  
<213> Homo sapien

<400> 108  
acatgtgcaa gaatttgga aagcagggca ttttcctca tctctcctag agggaatatc 60  
acagcatctg tctctactgg tccacactgg actgcagaca atgtcaaaac tctggatttg 120  
gaatgcggct gatttccttt cccctttaag gagttttcca agaatttcat aaccatcagt 180  
tggtatatatt ccagcttctt tgatgtcttt ttctataatt tcatagcagt caatgtaaat 240  
cttaacactt ttgagggtca ctacaatatg aaccttgtga aaacttccat aaaataatgt 300  
ctttacttct tctgtgtcaa atgtaacagt ttgcacctcg cctcttgat ccttggttaa 360  
gaatgataac gtcttgctag aaggatctgc aatcactcca acttggtggt tgtagtctct 420  
gtctgtgatt tgccaaattg caaaagggc actgggagtt tctgggagaa gtctgaat 478

<210> 109  
<211> 361  
<212> DNA

<213> Homo sapien

<220>

<221> misc\_feature

<222> (1)...(361)

<223> n = A,T,C or G

<400> 109

gaatttttct	tctanaataa	gtattctgtt	gacacagact	attggttaaga	ttttcaacat	60
aaggtaatgc	taggactggc	ctcctagcat	gagttgtgag	taaagatctg	gtctgttggt	120
tctccaaaag	aagnttctta	ctgcttgtct	ctcatgagtt	ttctgtttct	gctttctctt	180
tttcatattg	atatatacgg	ntttttaaat	ggtnattgta	attaaatata	tcctcatttt	240
tctcttttag	gagatgatgt	tgcattttcc	tctcaagaaa	atgaatatca	attgttatct	300
tgcttttgnt	gncagctttc	ttatgtgcat	gaactaattg	ctgttgaagc	cacatatttt	360
t						361

<210> 110

<211> 305

<212> DNA

<213> Homo sapien

<220>

<221> misc\_feature

<222> (1)...(305)

<223> n = A,T,C or G

<400> 110

acataatgac	tnncanagtg	aagctgattg	gctgcggttc	tggagtaaata	ataagctctc	60
cgttcctggg	aatccgcact	acttgagtc	cggtgcctggc	ctaccaaata	cttgccaaaa	120
ctatgtgcct	tatcccacct	tnnaatctgn	ctcctcattt	ntcagctggt	ggatcagaca	180
atgacattcc	tntagatntg	gcgatcaagc	attccanacc	tgngccaaact	gcaaacgggtg	240
cctncaagga	gaaaacgaag	gcncaccacaa	atgnaaaaaa	tgaangnccc	ttgaatgtac	300
taaaa						305

<210> 111

<211> 371

<212> DNA

<213> Homo sapien

<220>

<221> misc\_feature

<222> (1)...(371)

<223> n = A,T,C or G

<400> 111

cggggggccag	cgggggggtat	tcagccatcg	atcaaactca	aaacctggaa	tgatatccac	60
tctctttttc	ttaagctcag	ggaaatatcc	caagtagaag	tccagaaaag	catcggctaa	120
gatgcttcgg	aatttgaatt	catgcacata	ggccttgaga	aaactgtcaa	actgatcctg	180
atcacccacc	aagtggggcca	ggtatgagac	aaagcagaaa	cctttctcgt	aggggggtctc	240
attataggtg	tcgtccgggt	caacgcctgg	ttcaatcttc	acgcggagct	tggtgagtgg	300
gttttcctct	ccagtgatgt	ccatgtgctg	acgcagcaga	ncccgccccg	ttgcagcctc	360
caagcaggng	t					371

<210> 112

<211> 460

<212> DNA

<213> Homo sapien



<220>  
 <221> misc\_feature  
 <222> (1)...(460)  
 <223> n = A,T,C or G

<400> 112  
 acatcttagg ttttnttcc tttantgtga agaggcggtt ccaccaaccc acagctctgc 60  
 gtcgagtttt tactagattg ctgcaaattt catggaatct ttgctgttgt tcagtgggtcc 120  
 atttattgga gccaaaaatt ctagggcgct agaattggga caaggtagtc agccaagcac 180  
 aaaaacataa caaaacagga aacgccggac agaacagatg gatctagata gtagataatc 240  
 agaaacacca aagaaaccac acccatgatg gcaggtggaa accaggctct ttctcatcgg 300  
 aggactttat cagccatcag catcacttct ccccatcctt gcagctgttc ttccagactt 360  
 gcagctcttg cagccagcag gttgggtgct gcgattacct ccctccgcca tcgtctcggg 420  
 gatgcagtct ctacaagcgc aggccacctc cccaacgagt 460

<210> 113  
 <211> 204  
 <212> DNA  
 <213> Homo sapien

<400> 113  
 gagaagacag cagagctgct ttccgcctct ttgagaccaa gatcacccaa gtctctgact 60  
 tcaccaagga tgtcaaggcc gctgctaatac agatgcgcaa cttcctgggt cgagcctcct 120  
 gccgccttag cttggaacct gggaaagaat atttgatcat gggcttagat gggggcacct 180  
 atgacctcga gggacacccc cagt 204

<210> 114  
 <211> 137  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(137)  
 <223> n = A,T,C or G

<400> 114  
 accgcaagaa atgggacagc aacgtcattg agacttttga catcgncgcg tngacagtca 60  
 acgtgacgt gggctattac tcctggagggt gtcccaagcc cctgaagaac cgtgatgtca 120  
 tcaccctccg ntccctg 137

<210> 115  
 <211> 278  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(278)  
 <223> n = A,T,C or G

<400> 115  
 gcggggcggt ttntggactc gtcattttac agagcatgcg tggctcttcac ccttggcatg 60  
 ttctccgccg gcctctcgga cctcaggcac atgcgaatga cccggagtgt ggacaacgctc 120  
 cagntcctgc cttttctcac cacggangtc aacaacctgg gctggctgan ttatggggct 180  
 ttgaagggag acgggatcct catcgtcanc aacacagtgg gtgctgcgct tcanacctg 240  
 tatatctttg gcatactctg attactgccc tcggaagc 278

<210> 116  
 <211> 178  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(178)  
 <223> n = A,T,C or G

<400> 116  
 acaccgtcat angtcaaaag tncagtgtctg gccatcttgc atcaaagtgt ctttaaggcag 60  
 tgactggcta tcaaccacag nttctgtctc cccagntgca aacacaggat ccatgcaaca 120  
 gttctgagac catacactta gaaaccacng ggagatgcgg atcanatgca naactnnc 178

<210> 117  
 <211> 360  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(360)  
 <223> n = A,T,C or G

<400> 117  
 actccccaat ggnggattta ttactattaa agaaaccagg gaaaatatta attttaatat 60  
 tataacaacc tgaaaataat ggaaaagagg tttttgaatt ttttttttaa ataaacacct 120  
 tcttaagtgc atgagatggt ttgatgggtt gctgcattaa aggtatttgg gcaaacaaaa 180  
 ttggagggca agtgactgca gttttgagaa tcagttttga ccttgatgat tttttgtttc 240  
 cactgtggaa ataaatggtt gtaaataagt gtaataaaaa tccctttgca ttctttctgg 300  
 accttaaatg gtagaggaaa aggctcgtga gccatttgtt tcttttgctg gttatagtgt 360

<210> 118  
 <211> 125  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(125)  
 <223> n = A,T,C or G

<400> 118  
 gcgtcgtgct atgaccggac ttngtcttga aaggggatga cagcatggga ggcaatggnt 60  
 ncacatgtaa accccacact gaaagacaag gcactctctc cacagcagcc ccaacaacta 120  
 gccct 125

<210> 119  
 <211> 490  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(490)  
 <223> n = A,T,C or G

```

<400> 119
nacaaagaaa agcaaaaaga atttacgaag attgtgatct cttattaaat caattgttac      60
tgatcatgaa tgttagttag aaaatgttag gttttaactt aaanaaaatn gtattgngat      120
tttcaatntt atgttgaaat cngngtaata tcctgangtt nttttccccc cagaagataa      180
agaggataga caacctctta aaatattttt acaatttaat ganaaaaaagn ttaaaattct      240
caatacnaat caaacaattt aaatatttta agaaaaaagg aaaagtagat agtgatactg      300
agggtaaaaa aaaattgatt caattttatg gtaaaggaaa cccatgcaat tttacctaga      360
cagccttaaa tatgtctggt tttccatctg ctagcatttc agacatttta tgttcctctt      420
actcaattga taccaacaga aatatcaact tctggagtct attanatgtg ttgtcacctt      480
tctnaagctt

```

<210> 120

<211> 361

<212> DNA

<213> Homo sapien

<220>

<221> misc\_feature

<222> (1)...(361)

<223> n = A,T,C or G

```

<400> 120
caggtagagt aaaattaaca cttccgttac aggaaatgta tgacgcaat aatataaaat      60
taaaaaggta aaaaaagggtg acactgggtt cctaagatac aatttactct ttacaaccag      120
gggccacagg tccaggctgc anagcgggca tcaggaagca gagcctncca cctgcttctg      180
ggggacctgg taataaaaaat cagcccatga tggcgctatg gcctctcaga caccacacgc      240
tgctaataca cctagagctc tggaaatagt caacaggaga gtgatttcca tgggggaaat      300
tttaanaaag atgcacatgg gacaggcaat agaaagttag ccaaggntaa atttggtacc      360
t

```

<210> 121

<211> 405

<212> DNA

<213> Homo sapien

<220>

<221> misc\_feature

<222> (1)...(405)

<223> n = A,T,C or G

```

<400> 121
acacaaaacc ttttnacata ttgggggctt accgctccaa attgctactg atcctttaag      60
ttcacaatat agaatttctt caccaattaa gtaataaccc tcattacaaa taaagtgcac      120
ctgataacca aactcgtaag tcccatttgc agggactgct tggccattta aaggatcccg      180
tatatatgga catgtttctc tataacaggc gtcactctgag acaggtagcc atgtatgatt      240
ccgatcacia atagtatggg tggcaagagg aggtatatag aagtatcctt ttttacactt      300
ataatctact cgttcaccaa tctcatagta ggggttttgg ttaccaatga gcctccatan      360
cttcaaatgt tgggtggctn ctcacaggca tcnggcanaa ngagt

```

<210> 122

<211> 152

<212> DNA

<213> Homo sapien

<220>

<221> misc\_feature

<222> (1)...(152)

<223> n = A,T,C or G

<400> 122  
 accccgctcc gttgncacag atcgctgtct gccactcca tcggccattc acttggcagg 60  
 tgcgattggc agagccccgg agagtgtaac cgtcatagca gtggaaagag atctcatcac 120  
 tcacattgta gtagggagac cggggccaan ta 152

<210> 123  
 <211> 336  
 <212> DNA  
 <213> Homo sapien

<400> 123  
 acatctgaca tatTTatata gcacataaat tagggagtgc tctgaccctt gcccgTggag 60  
 cccaagcact gagcagggag gtgaacgccg gtccagaaag aaggTgctgg agcccttGct 120  
 ctgtctcttc catcacgggg ctcccctagg gcctccccag gcctccttgg ctCagtccag 180  
 gtgtctgcag gaggaaggTg ttgtctgcat ttagTgtctg agactgggtt tgaggaggca 240  
 ccagataaaa ggagatacac ttgcagctat aaagtCagct tcaaaccCCA gggcttGtaa 300  
 ttccaaggag agggTgggga ggcgaggcca tagtct 336

<210> 124  
 <211> 253  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(253)  
 <223> n = A,T,C or G

<400> 124  
 ctgcaagagc ccagatcacc cattccgggt tcaTccccg cctccccaag tcagcagTcc 60  
 tagccccaaa ccagcccaga gcagggtctc tctaaagggg acttgagggc ctgagcagga 120  
 aagactggcc ctctagcttc taccctttgt ccctgtagcc tatacagTtt agaattttta 180  
 tttgttaatt ttattaaaat gctttaaaaa aacaaaaaaa aaaaaaaaaa aaaaaaaaaa 240  
 aaaaaagntt gtn 253

<210> 125  
 <211> 522  
 <212> DNA  
 <213> Homo sapien

<400> 125  
 acaactgcaa gtctaagata atgttcattc attcccatca taaatgtaac attctaaata 60  
 ggtgtcttct gatgtcatct gtcagaattt cttttaaact ttttcttcat cttcaacatt 120  
 atcaaagttc atccttattc ctcttgccctt gatttcggag agtttccaat ttttcaactta 180  
 ttaaggcagc gattgtcttt gcattctctg tatttatctg ctcttcttga aaatttctct 240  
 ttgtctcttc gtagaaataa aacttaacag ttggataggc cctgatccca gctttctggc 300  
 atgtctgagc ataagcctga cagtctactt ttccagcttt cacttttcct ttaatcatcc 360  
 tagccaagag ctcaaattct ggagcaaaat tctggcaagg tccacaccaa ggagcataga 420  
 aatcaatcac ccaatgattt ttcccttgta gaacttttct actgaaagtc tgaggTgtta 480  
 gatctgtgga tacttgaggt aaaaatccta gacccagat tc 522

<210> 126  
 <211> 374  
 <212> DNA  
 <213> Homo sapien

<220>

<221> misc\_feature  
 <222> (1)...(374)  
 <223> n = A,T,C or G

<400> 126  
 tttttaagat attaacttta cttttataaa tctttgtgtg aaatgaaaa aaaaatcaag 60  
 gcatacaaat ttcattgtgt tctacatttt taaataccat cttttgtctc cgttaaaaga 120  
 ttttcatcca tttattcaaa aaccttttaa gttcaactgt ccaatttaag acagagtga 180  
 gacatttttg agtatctgaa ctaagcattg tcttgactga aacgaagtaa gaactcaatg 240  
 agagtccttg tgggcctccc aggcattgct ttccgtagat agggaaacttc atctttgttg 300  
 gncatcacgc ctgctatgtc taaatgtgcc cacttaggat gagttacgaa ttctttcagg 360  
 aatgctgcag ctgt 374

<210> 127  
 <211> 130  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(130)  
 <223> n = A,T,C or G

<400> 127  
 aaagccaaga cngccattgg cactgctatg gtaaggncac agggcancca gggccttctg 60  
 gcaaaaggng atacnaccag cactatnaac agacaggaca tgggtgagag gnagnctaca 120  
 caantcctaa 130

<210> 128  
 <211> 350  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(350)  
 <223> n = A,T,C or G

<400> 128  
 aactgattt ccgntnaaaa gaancatcat ctttaccttg acttttcagg gaattactga 60  
 actttcttct cagaagatag ggcacagcca ttgccttggc ctcaactgaa gggctctgat 120  
 ttgggtcttc tggctctctg ccaagnttcc cagccactcg agggagaaat atcgggaggt 180  
 ttgacttctt ccggggcttt cccgagggtc tcaccgtgag ccctgcggcc ctgagggtg 240  
 caatcctgga ttcaatgtct gaaacctcgc tctctgcctg ctggacttct gaggccgtca 300  
 ctgccactct gtctccagc tctgacagct cctcatctgt ggctgttga 350

<210> 129  
 <211> 505  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(505)  
 <223> n = A,T,C or G

<400> 129  
 acaataccaa agcttcataa tgctaaagaa aaccaaaca aaagacaatg gtttacacag 60

```

ggaaataacc ctaaggcaat atgaaaacag tcataattta ttactgataa agagtaaagg 120
catccttccc atagaggggg ggaattcaca gggaacacta attatatcag atgaaccacg 180
gggatagaaa ataggcccat ttttaaaatt cattgagaaa ttattacttt ttctccacaa 240
ctgtgattct atacaaaata taaaccctgc aaaccttatg tgctacctga cagataaaaag 300
tagcaggagc cagactcttg aagcacttga gactgatttc tacaaagtcc aggaagagca 360
atgattccag tgtgcagtgc tgatgcatgt gtgagcctaa catgtttatc agctctgggt 420
gcagcccat ctacatggg cccagttagt ttttagggag tcacagatta ngcaggcaac 480
cgaggggcat gatttaaaaa gcaca 505

```

&lt;210&gt; 130

&lt;211&gt; 526

&lt;212&gt; DNA

&lt;213&gt; Homo sapien

&lt;400&gt; 130

```

acaaaagagc ctgattcttt ttaattccac aaatacctag catctcaaag taacatgtaa 60
acaaacttct atgctgctca atgaatcctt ccaatttcga taataaacta aatagtattg 120
gatctagtat atgactttca tgtgtaagtt atgggttctat ccattacttt aacaatatta 180
ctgatgtaac agagaaaaat tttcaactat tgtacttatt taaaacaaac tgacaagttc 240
aagcacctgt ctccagaaaa gccagcagca tttttttttt tttaacatac tcaaagtaag 300
atgtggccta agcccttaat acctttctga acagccatgc aactaaacac cctcaggaga 360
tgttacataa gggagagaag aacatggagc aatttgact ttttccccta gataatatta 420
acaaggtaaa gcaaatccag atctttatga atgaatggct gtcatgttta atacacttgg 480
agctctataa aactagagcc actatcatat atgtttatat agatat 526

```

&lt;210&gt; 131

&lt;211&gt; 477

&lt;212&gt; DNA

&lt;213&gt; Homo sapien

&lt;400&gt; 131

```

ctcagttttc ccagcaacag atgctcctga gcaatttatt agtcaagtga cgggtgctgaa 60
atacttttct cattacatgg aggagaacct catggatggg ggagatctgc ctagtgttac 120
tgatattcga agacctcggc tctacctcct tcagtggcta aaatctgata aggccctaata 180
gatgctcttt aatgatggca cttttcaggt gaatttctac catgatcata caaaaatcat 240
catctgtagc caaaatgaag aataccttct cacctacatc aatgaggata ggatatctac 300
aactttcagg ctgacaactc tgctgatgtc tggctgttca tcagaattaa aaaattgaat 360
ggaatatgcc ctgaacatgc tcttacaaaag atgtaactga aagacttttc gaatggacct 420
tatgggactc ctcttttcca ctgtgagatc tacagggaa ccaaaagaat gatctag 477

```

&lt;210&gt; 132

&lt;211&gt; 404

&lt;212&gt; DNA

&lt;213&gt; Homo sapien

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; (1)...(404)

&lt;223&gt; n = A,T,C or G

&lt;400&gt; 132

```

accacacgan cgggnatcnt ttgnacatag tgagaccggg ctgattccca tacatgaatc 60
cattcatgga gtgcatttta ttagatncct gaaagtcttc atcttcttta tccacctgat 120
caggngcagt tgtaaactn cctaataatta tcttcagga gtaaactctc attctcatca 180
aatactgtag gaaacaaata gaattccttg tctacatctt tctgtctccc atttgcatat 240
aaacttcctt tcttgcatat tttcattggc ccaataagcc cagtgaatat atcttttagtg 300
ggatccacag cagaataata catcttagct agacacacag ggatctgcat tacgnggggc 360
ctacttcttt ggggacagcc cttcatacgn gaatgtttnt gtgg 404

```

<210> 133  
 <211> 552  
 <212> DNA  
 <213> Homo sapien

<220>  
 <221> misc\_feature  
 <222> (1)...(552)  
 <223> n = A,T,C or G

<400> 133  
 accccaaatt atctctctcc tgaagtcctc aacaaacaag gacatggctg tgaatcagac 60  
 atttggggccc tgggctgtgt aatgtataca atgttactag ggaggccccc atttgaaact 120  
 acaaactctca aagaaactta taggtgcata aggggaagcaa ggtatacaat gccgtcctca 180  
 ttgctggctc ctgccaagca cttaattgct agtatgttgt ccaaaaaccc agaggatcgt 240  
 cccagtttgg atgacatcat tcgacatgac ttttttttgc agggcttcac tccggacaga 300  
 ctgtcttcta gctgtttgtca tacagttcca gatttccact tatcaagccc agctaagaat 360  
 ttctttaaga aagcagctgc tgctcttttt ggtggcaaaa aagacaaaagc aagatatatt 420  
 gacacacata atagagtgtc taaagaagat gaagacatct acaagcttag gcatgatttg 480  
 aaaaagactt caataactca gcaaccagc aaacacaggg acagatgang agtccacca 540  
 cctaccacca ca 552

<210> 134  
 <211> 496  
 <212> DNA  
 <213> Homo sapien

<400> 134  
 acattgatgg gctggagagc aggggtggcag cctgttctgc acagaaccaa gaattacaga 60  
 aaaaagtcca ggagctggag aggcacaaca tctccttggg agctcagctc cgccagctgc 120  
 agacgctaatt tgctcaaaact tccaacaaag ctgcccagac cagcacttgt gttttgatcc 180  
 ttcttttttc cctggctctc atcatcctgc ccagcttcag tccattccag agtcgaccag 240  
 aagctgggtc tgaggattac cagcctcacg gagtgcacttc cagaaatata ctgaccaca 300  
 aggacgtaac agaaaatctg gagaccaag tggtagagtc cagactgacg gagccacctg 360  
 gagccaagga tgcaaatggc tcaacaagga cactgcttga gaagatggga gggaagccaa 420  
 gaccagtggt gcgcacccgg tccgtgctgc atgcagatga gatgtgagct ggaacagacc 480  
 ttttctgggc cacttt 496

<210> 135  
 <211> 560  
 <212> DNA  
 <213> Homo sapien

<400> 135  
 actgggagtg atcactaaca ccatagtaat gtctaattatt cacaggcaga tctgcttggg 60  
 gaagctagtt atgtgaaagg caaatagagt catacagtag ctcaaaaggc aaccataatt 120  
 ctctttgggt cagggtcttgg gagcgtgac tagattacac tgcaccattc ccaagttaat 180  
 cccctgaaaa ctactctca actggagcaa atgaactttg gtcccaaata tccatctttt 240  
 cagtagcggt aattatgctc tgtttccaac tgcatttcct ttccaattga attaaagtgt 300  
 ggctcggtt ttagtcattt aaaattgttt tctaagtaat tgctgcctct attatggcac 360  
 ttcaattttg cactgtcttt tgagattcaa gaaaaatttc tattcttttt ttgcatcca 420  
 attgtgcctg aacttttaaa atatgtaaat gctgccatgt tccaaacca tcgtcaagtg 480  
 tgtgtgttta gagctgtgca ccctagaaac aacatattgc ccatgagcag gtgcctgaac 540  
 acagaccctt ttgcattcac 560

<210> 136  
 <211> 424

<212> DNA  
<213> Homo sapien

<220>  
<221> misc\_feature  
<222> (1)...(424)  
<223> n = A,T,C or G

<400> 136  
accagcaaat ctccattagc atttctcagg ttctcatgac cttttcagat atgttggttg 60  
attttatgta tatattgctt agaaacaaaa atccacctga tattaaaaca aaccaaaaaa 120  
aatcataaaa gcaagcaaat gaacaaaaaa ccctagtttt gttgtgcttt tctttcacat 180  
ttcctacagg gagatttgta tatctcagat actttcaaaa tctaataagg aagtaaaatt 240  
agtgccttaa ccaaacagta agataccaaa gaatcctcca tcacaagtta ctgaatcaaa 300  
cttctcatga catttgcggt atattcagat ttgaagattt tttaaattta gaatttaaaa 360  
caaacttttag actgctgatt ttccatattt caaagactgt agctgtntgc agcatataaa 420  
tgga 424

<210> 137  
<211> 392  
<212> DNA  
<213> Homo sapien

<220>  
<221> misc\_feature  
<222> (1)...(392)  
<223> n = A,T,C or G

<400> 137  
tgccgggntg aaggctagca aaccgagcga tcatgtcgca caaacaaatt tactattcgg 60  
acaaatacga cgacgaggag tttagagtac gacatgtcat gctgccaag gacatagcca 120  
agctgggccc taaaaccat ctgatgtctg aatctgaatg gaggaatctt ggcgatcagc 180  
anagtcaggg atgggtccat tatatgatcc atgaaccaga acctcacatc ttgctgttcc 240  
ggcgccact acccaagaaa ccaaagaaat gaagctggca agctactttt canctcaag 300  
ctttacacag ctgnccttac ttcctaacat ctttctgata acattattat gctgccttcc 360  
tgttctcact ctganatnta aaagatgttc aa 392

<210> 138  
<211> 284  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1)...(284)  
<223> n = A,T,C or G

<400> 138  
tgccctgtgca cctcttttgc tgaaatatgg caagacttgg aaaaatgttt gcccttagaa 60  
tctatctcac tactttagtt agttgtctcc ttggggcctg ggcacagttc tggccctgat 120  
ctggaacaga ctcccttttc taaaactgaa cttgaccaca tcaaaagntt gnaaaacaat 180  
ctccatggta attaaacttg cattcaacac catatggnaa cagaagatgg caggaggata 240  
anatncagat cttatgatct ttccangnan ggcagtgtac atga 284

<210> 139  
<211> 249  
<212> DNA



<213> Homo sapiens

<220>

<221> misc\_feature

<222> (1)...(249)

<223> n = A,T,C or G

<400> 139

```
gaggaagggg ggactgaatc tancacntg acngaactag agacagccat gggcatgac 60
atagacnnct ttacccgata ntccgggcagc gagggcagca cgcagaccct gaccaagggg 120
gagctcaagg ggctgatgga gaaggagcta ccaggcttcc ngcagagngg aaaaanacaag 180
gangccgtgg ataaattgct caaggaccta gagcccnatg gaggatgcc cagggtggactc 240
cagcgagnt 249
```

<210> 140

<211> 390

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> (1)...(390)

<223> n = A,T,C or G

<400> 140

```
tcataatggt tggggcagct ataatnnact acaanaatca natgtttcac atctagacct 60
cgggcagcaa cagaggtagc cacaagaagt ttgcangtcc cattcttaaa gtcatttatg 120
atgctatctc tgtcatattg atcaatgcct ccatgaagag acatgcaagg ataagatgct 180
ctcattaaat ccttaagaag accatcagca tgttctctgct tatccacaaa tataatgaca 240
gatcctgact cttgataatg gcctagaagc tcaagtaact tcaagaattt cttttcttct 300
tcaatcacaa tcacttgtn gctccacatc gagcaaacca cactcctgcc tccaacttgt 360
acctgccccg ggccggcgct caagggcgaa 390
```

<210> 141

<211> 420

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> (1)...(420)

<223> n = A,T,C or G

<400> 141

```
gacactcagg gaaaagcatn ngncaaanag agcttaaaat gcatcgccaa cggggtcacc 60
tccaagggtc tcctcgccat tcggagggtgc tccactttcc aaaggatgat tgctgagggtg 120
caggaagagt gctacagcaa gctgaatgtg cgcancatcg ccaagcggaa ccngaagcc 180
atcactgagg tcgtgcagct gcccaatcac ttctccaaca natactataa cagacttgnn 240
cgaagcctgc tggaatgnga tgaanacaca gggcagcaca atcaggagac agcctgatgg 300
anaaaantgg gcctancatg gccaggcctc ttccacatcc tngcangaca gaccactgtg 360
cccaaacaca ccnctgagc tgacttnnac aggagacgca cnaaggagcc cggcagangc 420
```

<210> 142

<211> 371

<212> DNA

<213> Homo sapiens

<400> 142

```

gggttcgaca atgctgatcc gcaattagaa gacactggta agctgtgtta cactgggctt 60
cattgaaatc ttcaaggata tagccagctc ctgctcgaag ctgggattct gtatactgct 120
tgttgaaagg aggaatttcc aaaaattcct cctcttcttc actgcttctt gtaggacct 180
ctggcagttt ggagcggctg gccaaactgt cactggttgt ggccatggta aggagaaatg 240
cgtagcccag aaacaaggtc ttgttgagag gcaaaggccc tctctgctct tccagggcag 300
agggttcacc ggtgttgtct ccactctcac agggggtcac aaactctcct gccctactt 360
gcaccagggt t

```

371

&lt;210&gt; 143

&lt;211&gt; 270

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; (1)...(270)

&lt;223&gt; n = A,T,C or G

&lt;400&gt; 143

```

ggtggctgtg atnacctttn ttagtttaca aataaaaaag ntaaaaagaa atactgtgtt 60
tagggtaagg taacannttc atctaatacag aggagagtga agangaggcn ctgccttcta 120
ggngctgtga ccttctcctt ttcgngattc ttcnccacct tgggnaacat cttccccgct 180
atgctggaan tacttcggng ttctgcggtg gccatgntga acatctgatg aactgaaant 240
ncatccnaat gcacacgaag anatagncna

```

270

&lt;210&gt; 144

&lt;211&gt; 259

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; (1)...(259)

&lt;223&gt; n = A,T,C or G

&lt;400&gt; 144

```

ttctctttgc tttttataat tttaaagnaa ataacacatt taactgtatt taagtctgtg 60
caaataatcc ttcagaagaa atatccaaga ttctgtttgc agaggtcatt ttgtctctca 120
aagatgatta aatgagtttg tcttcagata aagtgtcctt gtccagnaga actcaaaagg 180
ccttcaagct gttcagtaag tgtaggttca gataagactc cgncatacga attccagctt 240
cccgtgcccc ctgtacctc

```

259

&lt;210&gt; 145

&lt;211&gt; 433

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; (1)...(433)

&lt;223&gt; n = A,T,C or G

&lt;400&gt; 145

```

accacatnta ccatagtgtg attagtttta attttcacat gaatcaaagg tttcctttca 60
tgtctattta cagtccaatt gtgccaaact cttacttgtg tgctgactaa caaggcattt 120
aggtgtgcag catcctagag tgctccaggg cagtgtcagc gttctcggga gtaaaagggtg 180
ccacttggtg gcaatgatat tccagaatta aatgggtttt tgttgccatg gagactgcat 240
ttatataaat gtagcctgta gcttaagtta actaaacctt atgctgctgt taaaaacagt 300

```

ttattttaat attaaaatac agttgattag caacagcggg gctgtatttt aagagacact 360  
ttattggaag tgcaatcata gttatttggg ttcacaattt tacagngcat tctaattact 420  
gatgggtgca att 433

<210> 146  
<211> 576  
<212> DNA  
<213> Homo sapiens

<400> 146  
acctcaggcc tgtgcacctc tttgcttgaa atatggcaag acttggaata atgtttgccc 60  
ttagaatcta tctcactact ttagttagtt gtctcctttg ggcctgggca cagttctggc 120  
cctgatctgg aacagactcc cttttctaaa actggacctt gaccacatca aaagtttgta 180  
aaacaatctc catggtaatt aaacttgcac tcaacaccat atggtaacag aagatggcaa 240  
aggataagat tcagatctta gatctttcca agtagggcat gttagatgat agaaggatta 300  
gttgcaagct ggatctgagc tcaggcttgg gcatgaagga aactgtctcc catgtgggtt 360  
ggaagagtta ggggctccct gagctctatt gtgaactata cgggtttcat ccaaggaatg 420  
gtatgatgtg ggcataaaac cattcttcag acaactgaag atggctccct tctgtagcca 480  
gaaacactag ctgtcctgca ttgccatttc ctttacccca ggcggcctgc agaaggaaaag 540  
gccataatta attaaaaggc ttaatgaagt tttgga 576

<210> 147  
<211> 300  
<212> DNA  
<213> Homo sapiens

<400> 147  
ccagccccc ggaggaaggt gggctctgaat ctagcaccat gacggaacta gagacagcca 60  
tgggcatgat catagacgtc tttaccgatg attcgggcag cgagggcagc acgcagaccc 120  
tgaccaaggg ggagctcaag gtgcttatgg agaaaggagc taccaggctt ctgcagagtg 180  
gaaaagacaa ggatgccgtg gataaattgc tcaaggacct agacgccaat ggagatgccc 240  
aggtggactt cagtgaagtc atcgtgttcg tggctgcaat cacgtctgcc tgtcacaagt 300

<210> 148  
<211> 371  
<212> DNA  
<213> Homo sapiens

<400> 148  
acataatcct cataatgggt ggggcagcta taatttacta caagaatcag atgtttcaca 60  
tctagacctc gggcagcaac agaggtagcc acaagaagtt tgcaggtccc attcttaaag 120  
tcattttatga tgctatctct gtcattatga tcaaatggcc tccatgaaga gacatgcaag 180  
gataagatgc tctcattaaa tccttaagaa gaccatcagc atgttctctg ttatccacaa 240  
atataatgac agatcctgac tcttgataat ggcctagaag ctcaagtaac ttcaagaatt 300  
tcttttcttc ttcaatcaca atcacttggt gctccacatc tgagcaaacc acactcctgc 360  
ctccaacttg t 371

<210> 149  
<211> 585  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1)...(585)  
<223> n=A,T,C or G

&lt;400&gt; 149

```

cgaggtacan cactgctaaa tttgacactn anggaaaagc attcgtcaaa gagagcttaa 60
aatgcatcgc caacgggggtc acctccaagg tcttcctcgc cattcgaggg tgctccactt 120
tccaaaggat gattgctgag gtgcaggaag agtgctacag caagctgaat gtgtgcagca 180
tcgccaagcg gaaccctgaa gccatcactg aggtcgtcca gctgcccaat cacttctcca 240
acagatacta taacagactt gtccgaagcc tgctggaatg tgatgaagac acagtcagca 300
caatcagaga cagcctgatg gagaaaattg ggcctaacat ggccagcctc tccacatcc 360
tgcagacaga ccactgtgcc caaacacacc cagagctga cttcaacagg agacgcacca 420
atgagccgca gaagctgaaa gtctcctca ggaacctccg aggtgaggag gactctcctt 480
cccacatcaa acgcacatcc catgagagtg cataaccagg gagaggntat tcacaacctc 540
ccaaactagt atcatttttag gggngttga cacaccagt ttgag 585

```

&lt;210&gt; 150

&lt;211&gt; 642

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; (1)...(642)

&lt;223&gt; n=A,T,C or G

&lt;400&gt; 150

```

acttncgggt tcgacaatgc tgatccgcaa ttagaagaca ctggtaagct gtgttacact 60
gggcttcatt gaaatcttca aggatatagc cagctcctgc tcgaagctgg gattctgtat 120
actgcttggt gaaaggagga atttccaaaa attcctcctc ttcttactg ctctctgtat 180
gaccatctgg cagtttgag cggtggcca acttgctact ggttggtggc atggttaagga 240
gaaatgcgta gccagaaac aaggtcttgt tgagaggcaa aggccctctc tgctcttcca 300
gggcagaggg ttcaccggtg ttgtctccac tctcacagg gctcaciaac tctcctgcc 360
ctactgcacc aggttttact gtggcagact tgcgacctcg cttggcaggg gaccgttctt 420
cttcagaagt gataagttt cttttgctg agagaactcc catggaggca cgaggacttt 480
ctgtgatctt tcgggtaggg gttgtgctgc tactggaggc agtanggggtg gctggggagc 540
tgacgttact gcgcggttgc cgcttcttcc caccaaattg ctaagctgat atctgctgcc 600
tttgtaagaa gnggtactgc ttcatanggg ccaagcccat ac 642

```

&lt;210&gt; 151

&lt;211&gt; 322

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; (1)...(322)

&lt;223&gt; n=A,T,C or G

&lt;400&gt; 151

```

nttgacaac atcttccccg ctatgctgga attacttcgg tgttctgcgg tggccatgg 60
gaacatctga tgaactgaaa tccatcggga atgcacagga agatatagtt gatcttcaaa 120
aatgtccttt ccaggaccac catactgggg aagttcttcc ggggcctgc naatgggctg 180
caccctgggg ctgggcccga gctctagctc tgtcatgcca tcgccactga aatcggtttn 240
cagatgatta gtctcttcat gccccgtcca tttttcggtt tttctccagt gttcagaaat 300
tcaaatagatt aacttctggg aa 322

```

&lt;210&gt; 152

<211> 262  
<212> DNA  
<213> Homo sapiens

<400> 152  
acaaagtctt ctctttgctt ttataaattt taaagcaa ataacattta actgtattta 60  
agtctgtgca aataatcctt cagaagaa atccaagatt ctgtttgcag aggtcatttt 120  
gtctctcaaa gatgattaaa tgagtttgct tttagaataa agtgctcctg tccagcagaa 180  
ctcaaaaggc cttcaagctg ttcagtaagt gtagttcaga taagactccg tcatacgaat 240  
tccagcttcc cgtgccact gt 262

<210> 153  
<211> 284  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1)  
<223> n=A,T,C or G

<400> 153  
ctcgggagta aaagtgcca cttggtagca atgatattcc agaattaaat gggtttttgt 60  
tgccatggag actgcattta tataaatgta gcctgtagct taagttaact aaaccta atg 120  
ctgctgttaa aaacagttta ttttaattt aaaatacagt tgattagcaa cagcggtgct 180  
gtattttaag agacacttta ttggaagtgc aatcatagtt atttgttttc acaattttac 240  
ngtgcattct aattactgat gggngcaatt acttttaatc gngg 284

<210> 154  
<211> 531  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1)...(531)  
<223> n=A,T,C or G

<400> 154  
accacccta aatttgaact cttatcaaga ggctgatgaa tctgaccatc aaataggata 60  
ggatggacct ttttttgagt tcattgtata aacaaatttt ctgatttgga ctttaattccc 120  
aaaggattag gtctactcct gctcattcac tctttcaaag ctctgtccac tctaactttt 180  
ctccagtgtc atagataggg aattgctcac tgcgtgccta gtctttcttc acttacctgg 240  
cctctgatag aaacagttgc ccctctcatt tcataaggctc gaggacttgt gaccctggat 300  
ggttctaaat ggaaaaagca ccgccagatt gtgaaacctg gcttcaacat cagcattctg 360  
aaaatattca tcaccatgat gtctgagagt gttcggatga tgctgaacaa atgggaggaa 420  
cacattgccc aaaactcacg tctggagctc tttcaacatg tctcctgat gaccctggac 480  
agcatcatga agtgtgcctt cagccaccag ggcagcatcc agttngacag t 531

<210> 155  
<211> 353  
<212> DNA  
<213> Homo sapiens

<220>

<221> misc\_feature

<222> (1)...(353)

<223> n=A,T,C or G

<400> 155

```
tcttgacaag actgagagag ttacatgttg ggaaaaaaaa agaagcatta acttagtaga 60
actgaaccag gagcattaag ttctgaaatt ttgaatcatc tctgaaatga agcagggtga 120
gcctgccctc tcatcaatcc gtctgggtgc cagaactcaa gggtcagtgg acacatcccc 180
ctgttagaga ccctcatggg ctaggacttt tcatctagga tagattcaag acctttacct 240
canaattatg taaactgtga ttgtgtttta gaaaaattat tatttgctaa aaccatttaa 300
gtctttgtat atgtgtaaat gatcacaaaa atgtatttta taaatgttc tgt 353
```

<210> 156

<211> 169

<212> DNA

<213> Homo sapiens

<400> 156

```
agtttgttct actacatttg tgggccacta gttcactttg ctgtgttgat aagcggttacc 60
accaattgca ctttctatag cctcttttac aatgttgctc acttcatcaa caacaaaagc 120
agtctctccc gcagcctggt agtcttccat ctttctccc gcgcgtccc 169
```

<210> 157

<211> 402

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> (1)...(402)

<223> n=A,T,C or G

<400> 157

```
gttaactacc cgctccgaga cgggattgat gacgagtcct atgaggccat tttcaagccg 60
gtcatgtcca aagtaatgga gatgttccag cctagtgcgg tggctttaca gtgtggctca 120
gactccctat ctggggatcg gttaggntgc tttaatctac tatcaaagga cagcccaagt 180
gtgtggaatt tgtcaagagc tttaacctgc ctatgctgat gctgggaggc ggtgggttaca 240
ccattcgtaa cgttgcccg tgctggacat atgagacagc tgtggccctg gatacggaga 300
tccctaata gcttccatac aatgactact ttgaataact tggaccagat ttcaagctcc 360
acatcagtc ttccaacatg actaaccaga acacgaatga gt 402
```

<210> 158

<211> 546

<212> DNA

<213> Homo sapiens

<400> 158

```
actttgggct ccagacttca ctgtccttag gcattgaaac catcacctgg tttgcattct 60
tcatgactga ggttaactta aaacaaaaat ggtaggaaag ctttctatg cttcgggtaa 120
gagacaaatt tgcttttgta gaattggtgg ctgagaaagg cagacagggc ctgattaaag 180
aagacatttg tcaccactag ccaccaagtt aagttgtgga acccaaaggt gacggccatg 240
gaaacgtaga tcatcagctc tgctaagtag ttaggggaag aaacatatc aaaccagtct 300
ccaaatggat cctgtgggta cagtgaatga ccactcctgc tttatttttc ctgagattgc 360
cgagaataac atggcactta tactgatggg cagatgacca gatgaacatc atcatcccaa 420
gaatatggaa ccaccgtgct tgcatcaata gatttttccc tgttatgtag gcattcctgc 480
catccattgg cacttggtc agcacagtta ggccaacaag gacataatag acaagtccaa 540
```

aacagt

546

&lt;210&gt; 159

&lt;211&gt; 145

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; (1)...(145)

&lt;223&gt; n=A,T,C or G

&lt;400&gt; 159

```
acttttgcta taagtttcct aaaaatattt aatacttttt tttttcaatt taaattaaat 60
ctnttgatga acaggggggg gntggcaaaa tttccaagcn ctggactgga attttganan 120
aggcatttac ngacctnat aactt 145
```

&lt;210&gt; 160

&lt;211&gt; 405

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;400&gt; 160

```
tgtaaatcgc tgtttggatt tcctgatattt ataacagggc ggctgggttaa tatctcacac 60
agtttaaaaa atcagcccct aatttctcca tgtttacact tcaatctgca ggcttcttaa 120
agtgacagta tcccttaacc tgccaccagt gtccccctc cgccccccgt cttgtaaaaa 180
ggggaggaga attagccaaa cactgtaagc ttttaagaaa aacaaagttt taaacgaaat 240
actgctctgt ccagaggctt taaaactggg gcaattacag caaaaaggga ttctgtagct 300
ttaacttgta aaccacatct tttttgcact ttttttataa gcaaaaacgt gccgttttaa 360
ccactggatc tatctaaatg ccgatttgag ttcgcgacac tatgt 405
```

&lt;210&gt; 161

&lt;211&gt; 443

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; (1)...(443)

&lt;223&gt; n=A,T,C or G

&lt;400&gt; 161

```
tttgctttta atgaaggaca agggattaag acncatagag actggccana caaatgggaa 60
accgaccaga ccagcccatg accaaaatat cacaggcaga ccaccacaa atgcagaggc 120
ctcagagtcc acagtgggag gttggaaccc agggcccag ggaatctttc agctgcattc 180
cggctgtgat cggcgggcaa caggtagagg tgctggaggg ggctgagtcg tgattttcgg 240
tgtctgtcat attcgatcaa gtgtgtcata gagcttctctg tttcatctec cagttattca 300
aggagaggct ggtggctcca cttcccagg aactgtgctg tgaagatctg aagacaggca 360
cgggctcagg caccgcttgt ctggaatgtc aatttgaaac ttaaaaagca gcgaccatcc 420
agtcatttat ttccctccat tcc 443
```

&lt;210&gt; 162

&lt;211&gt; 228

&lt;212&gt; DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> (1)...(228)

<223> n=A,T,C or G

<400> 162

```
tcgttatcaa aatggaagac accaaacccat tactggcttc taagctgaca gaaaaggagg 60
aagaaatcgt ggactagtgg agtaaatttt atgcttnctc aggggaacat gaaaaatgcg 120
gacagtatat tcagaaaggc tattccnagc tcaagatata tnattgtgaa ctanaaaata 180
tagcanaatt tgagggcctg acagacttct canatacnnt caagttgt 228
```

<210> 163

<211> 580

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> (1)...(580)

<223> n=A,T,C or G

<400> 163

```
acccaaggct acacatcctt ctgtgaaaca gtctcacgga gactctcaga atcccaagaa 60
ttttcttcaa ccttcttttg ttttgattct gaagggaaca tctgatctgc tctcaatgtt 120
tggtcattct tcaattccaa ggctttattt ggaacagact ttgcatttca atggcaggct 180
cgaaggcaga tggcttctcg ggaggctctg ctttgaaagt ttgcntgtcc atcaattcta 240
aggctttagn tggaaatagaa actttcattc tgcagggagc cttcagaaaa ccatcattat 300
caggagactc ttctaatttt ccatttattt tatctatttc tttttgatgc gcagccttgg 360
gtanacacac atccttctgt gaaacagtct cacagagact ctccagaatcc caagaacttt 420
cttcatagtc cttttgtttg gattctgatg ggagtatctc atctgctctc aatgtttgtt 480
cattcttcaa ttccaaggct ttatttggaa cagacttttg catttcaatg gcaggctcga 540
aggcagatgg cttctcggga ggctctgctt tgaaaagtgt 580
```

<210> 164

<211> 140

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> (1)...(140)

<223> n=A,T,C or G

<400> 164

```
acttatatct tttggnccttg ggcttctcaa agttcacgac agacataggc actctcacag 60
tatcaagccc atttaccgnc acctcacacc aatactcgcc ccaccgngng ataggntctg 120
ctggnaactt taatgnatgn 140
```

<210> 165

<211> 370

<212> DNA

<213> Homo sapiens



<220>  
 <221> misc\_feature  
 <222> (1)...(370)  
 <223> n=A,T,C or G

<400> 165  
 acatggagcc actgccacca gtggtgatgg aaagcactgc cttcttactc cggaaggggc 60  
 ctttgtcata catggcagcg taagtgttaag caaactctcc tatgaacact cgctcaaacc 120  
 agcctttcag aatggcaggg actccaaacc actgcnnngg ggaactggaa tatcacaagg 180  
 tctgcggctt ccagcttctt ttgttcagcc acaatatctg ggctcanatg gncttcttta 240  
 taagccagaa cagactcggg aggatactga aagttcgcag ggnccctcan ttacctgng 300  
 atgncccttn tggaaatgat gggattgaag ntcattggnat aaaggncgga ctncaccacc 360  
 tccattcttt 370

<210> 166  
 <211> 258  
 <212> DNA  
 <213> Homo sapiens

<400> 166  
 gtcaaaagtc atgatttttta tcttagttct tcattactgc attgaaaagg aaaacctgtc 60  
 tgagaaaaatg cctgacagtt taatttataaa ctatgggtgta agtctttgac aagaaaaaaa 120  
 aacaaacaaa cacttctttc catcagtaac actggcaatc ttctgttaa ccactctcct 180  
 tagggatggg atctgaaaca acaatgggtca ccctcttgag attcgtttta agtgtaattc 240  
 cataatgagc agaggtgt 258

<210> 167  
 <211> 345  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> (1)...(345)  
 <223> n=A,T,C or G

<400> 167  
 ggtcagccaa acaccagga tctctgtaaa actgaagaac aggncaatgc caccaacaaa 60  
 tctcaaaacc tctccagcat attctcctat gattggagca catggngagc acnantgggc 120  
 acttttaaca canctagcca gacaggngnc atttgggtta acacttcgga acccacagca 180  
 ntttanantt ctctggatgt catttcgagc acttgtattt attggtcann tttctgtatc 240  
 tngcgcttgg ttagccctga accaggagca acaggngcag cttctggagg ntggttgga 300  
 caatacggca agtgnrngaa atgacatcca acctncngaa atgac 345

<210> 168  
 <211> 61  
 <212> DNA  
 <213> Homo sapiens

<400> 168  
 gatagtgtgg tttatggact gaggtcaaaa tctaagaagt ttcgcagacc tgacatccag 60  
 t 61

<210> 169

<211> 344  
 <212> DNA  
 <213> Homo sapiens

<400> 169  
 acattggtgc tataaatata aatgctactt atgaagcatg aaattaagct tcttttttct 60  
 tcaagttttt tctcttgtct agcaatctgt taggcttctg aaccaagacc aaatgtttac 120  
 gttcctctgc tgcataccaa cgttactcca aacaataaaa aatctatcat ttctgctctg 180  
 tgctgaggaa tggaaaatga aacccccacc ccctgacccc taggactata cagtggaaac 240  
 tgttcattgc tgatgaatgc agcagtcacc aaaaaatata cccaatcttc cagataacct 300  
 cagtgcactt taggaaatca aaaattacct ggaagcaatt tagt 344

<210> 170  
 <211> 114  
 <212> DNA  
 <213> Homo sapiens

<400> 170  
 agcagtgtgt cctccatgaa taaacaggag ttctggaggg ccatcttctg catctttctg 60  
 tgattgttct tccccaattt tacttaaate ccacacattc aggcggcggt cagt 114

<210> 171  
 <211> 150  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> (1)...(150)  
 <223> n=A,T,C or G

<400> 171  
 actgagagca tttataatct gaccaaaattc ataggcatta ttaggcttgg ctatcggaag 60  
 tttctcaggg tcttctggng acctgctgct tttgctccc ttctcanaag caaggcatcc 120  
 catggagacc tcccctgcag ggcttccagg 150

<210> 172  
 <211> 435  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> (1)...(435)  
 <223> n=A,T,C or G

<400> 172  
 atttgttttc cactgcctca cactagtgag ctgtgccaa tagtagtgtg acacctgtgt 60  
 tgtcatttcc cacatcacgt aagagcttcc aaggaaagcc aaatcccaga tgagtctcag 120  
 agagggatca atatgtccat gattatcttc tggtttaggt ctacagtcaa tgtgatgggt 180  
 gtctttgctt ccagctctgc cagaatatct ttgtgcttct ctaatcattg gctttaaagc 240  
 taatcaatgt gttggcagca tctctgtcac tcttgtttaa cacgtgaaga aatcaggtag 300  
 atttttttct gtggcattgt tttcggacct aaaatcaggt atgctgacta tttccaaggg 360  
 gtttttcagt tgettcattt gcttgtaaaag cagggaatcc tcttgntgct tttctttttc 420  
 tcgatgagcc cgtgt 435

<210> 173

<211> 622  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1)...(622)  
<223> n=A,T,C or G

<400> 173  
actgntttcc cccaagtcca tgacatgtat acataattaa tggtttgctt ccttgattgt 60  
tttctccaac atccagacat agaggctgac caacgctttt aatgtatcca gatataacag 120  
gattaagggtc tggcacatac acctctggat aaatgttggt cagataccat gtaaaatttt 180  
tacctgaag gcggtgtttt atttcaaate tttttgaaag atcaccaaat gctttttgtt 240  
taacaatttt tgcctgcact gtattttctc tataaaatat ttctttgtat tcatccatcc 300  
agactttctgc aaggcgaact tggtttctag caatcacctg agtgcctttt ggaaagctat 360  
gagggtctttt gctgcgaaaa acatgtccaa caacagagca aggcataatc tccaactgcc 420  
caccacattg ccatactctg aaagacattt ctatattttc acctccccag atttccattt 480  
cttcatcata gcttccaata tactcaaaat attcttttga tatggaaaaa agtcctcctg 540  
caaaagtggg tgttttaatt gggtagggtt catctttcct tctttgcttc tcatgatcag 600  
gaagcgactt ccaccaatg aa 622

<210> 174  
<211> 362  
<212> DNA  
<213> Homo sapiens

<400> 174  
acggtgcagt tgacccactg ttggctctcc ttgcagttcc tgatatgtca tcttttagcat 60  
gtggctactt acgtaatctt acctggacac tttctaactt ttgccgaac aagaatcctg 120  
caccctccgat agatgctgtt gagcagattc ttctacctt agttcagctc ctgcatcatg 180  
atgatccaga agtgttagca gatacctgct gggctatttc ctaccttact gatggtccaa 240  
atgaacgaat tggcatgggt gtgaaaacag gagttgtgcc ccaacttgtg aagcttctag 300  
gagcttctga attgccaatt gtgactcctg ccctaagagc catagggaat attgtcactg 360  
gt 362

<210> 175  
<211> 486  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1)...(486)  
<223> n=A,T,C or G

<400> 175  
acagntnctc tactacactc agcctcttat gtgccaagtt tttctttaag caatgagaaa 60  
ttgctcatgt tcttcatctt ctcaaatcat cagaggccga agaaaaacac tttggctgtg 120  
tctaaaactt gacacagtca atagaatgaa gaaaattaga gtagttatgt gattattttca 180  
gctcttgacc tgtccctctt ggctgcctct gactctgaat ctcccaaaga gagaaaccaa 240  
tttctaagag gactggattg cagaagactc ggggacaaca tttgatecaa gatcttaaat 300  
gttatattga taaccatgct cagcaatgag ctattagatt cattttgga aatctccata 360  
atttcaattt gtaaaacttt ttaagacctg tctacattgt tatatgtgtg tgacttgagt 420  
aatgttatca acgtttttgt aaatatttac tatgttttcc tattagctaa attccaacaa 480  
ttttgt 486

<210> 176  
 <211> 461  
 <212> DNA  
 <213> Homo sapiens

<400> 176  
 accctggcca ctcttttcc tttggctggc caatgtctcc tctgtaggct ccagaagget 60  
 ctcagggatg caggcggcct cctgcagggt tgagttgcaa tgggaacaaa gacagctgtg 120  
 gtcccatagc accctcatct ggtgacatcc tgctactgac agtcaaaaga agccttccca 180  
 gatgaaatth tagtctctctg cgcagccatg ctcttcttcc agcaaaagag ccatgtgcag 240  
 tggggtctgc tccccatggg ggctttgatg tgggcccagc agtggatcag ccttccagac 300  
 acgtcgaact ctgcacactc ttctgcccgc ctcaggcttt ccaggaccct cccgagcctt 360  
 atcagagtcc ttaccctcag ggctactgat accttgctgg gtgaccttgg acagattcac 420  
 ttacctggag tcagtttcat aatatgaaaa tgatagggtt g 461

<210> 177  
 <211> 234  
 <212> DNA  
 <213> Homo sapiens

<400> 177  
 acacattttg taattacctt ttttgttgtt ttgtagcaac catttgtaaa acattccaaa 60  
 taattccaca gtcctgaagc agcaatcgaa tccctttctc acttttggaa ggtgactttt 120  
 caccttaatg catattcccc tctccataga ggagaggaaa aggtgtaggc ctgccttacc 180  
 gagagccaaa cagagcccag ggagactccg ctgtgggaaa cctcattgtt ctgt 234

<210> 178  
 <211> 657  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> (1)...(657)  
 <223> n=A,T,C or G

<400> 178  
 gagctcggan ccctagtaac ggccgccagg gtgctggnat gngcccttgc gagcgnngncg 60  
 cccgggcagg nactttnatc cccctcatc ttctgtagc tcatttgtnt ctctcatttt 120  
 ttggcatatt tttcaagtca cacttaaaaa ctcttccatg tattcacttc tcatcacttg 180  
 gtctacatgc cgaacctaa gtcaggatcc caaaaagatg agtatcctct caaacgcctc 240  
 ctaagcctct ggtatacatg actttggctg tgcacttcat ttagacttca cctttttgtt 300  
 tgctgttgtt ttttactacta gattcctttg tcttcattaa agataatgaa agattcacat 360  
 cacagtgcag ctcttcgctt tgcctttctg taagtccgta gcaactgccg agagtctctg 420  
 tctgctaggc atgtgtgaaa tccgctttgt ggctctctgt gatttgttcc gcttaacgtt 480  
 tttatttgtc ttatttacac atgccaaggt ggcaacgtga aaaatgtctc tgacgctatt 540  
 ttccgactgt aaagctgagc attcgatata agtagctgct ccaatctgtt tggccatact 600  
 tgccccctgg tcataggaca ctggcgctctg cctgtgattg gagagctcta ctaatgt 657

<210> 179  
 <211> 182  
 <212> DNA  
 <213> Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; (1)...(182)

&lt;223&gt; n=A,T,C or G

&lt;400&gt; 179

```

acaaaanctt ttaaatttta tattattttg aaactttgct ttgggtttgt ggcaccctgg 60
ccaccccatc tggctgtgac agcctctgca gtccgtgggc tggcagtttg ttgatctttt 120
aagtttcctt ccctaccag tccccatttt ctggttaagg ttctaggagg tctgttaggt 180
gt 182

```

&lt;210&gt; 180

&lt;211&gt; 525

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;400&gt; 180

```

acacgctttt ggccccgacc aatgaggcct tcgagaagat ccctagttag actttgaacc 60
gtatcctggg cgaccagaa gccctgagag acctgtgaa caaccacatc ttgaagtcag 120
ctatgtgtgc tgaagccatc gttgcggggc tgtctgtaga gacctggag ggcattgacac 180
tggaggtggg ctgcagcggg gacatgctca ctatcaacgg gaaggcgatc atctccaata 240
aagacatcct agccaccaac ggggtgatcc actacattga tgagctactc atcccagact 300
cagccaagac actatttgaa ttggctgcag agtctgatgt gtccacagcc attgaccttt 360
tcagacaagc cggcctcggc aatcatctct ctggaagtga gcggttgacc ctctggctc 420
ccctgaattc tgtattcaaa gatggaaccc ctccaattga tgccataca aggaatttgc 480
ttcggaacca cataattaaa gaccagctgg cctctaagta tctgt 525

```

&lt;210&gt; 181

&lt;211&gt; 444

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;400&gt; 181

```

acaccacaat gtgcatcaag gagacgtgcc gattgattcc tgcagtcccg tccatttcca 60
gagatctcag caagccactt accttcccag atggatgcac attgcctgca gggatcaccg 120
tggttcttag tatttggggg cttcaccaca atcctgctgt ctggaaaaac ccaaaggtct 180
ctgaccttct gaggttctct caggagaatt ctgatcagag acacctctat gcctacttac 240
cattctcagc tggatcaagg aactgcattg ggcaggagtt tgccatgatt gagttaagg 300
taaccattgc cttgattctg ctccacttca gactgactcc agacccacc aggcctctta 360
ctttcccaa ccattttatc ctcaagccca agaatgggat gtatttgcac ctgaagaaac 420
tctctgaatg ttagatctca gggg 444

```

&lt;210&gt; 182

&lt;211&gt; 441

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;400&gt; 182

```

acaaccttta ttgcttctcc agcattttcc agaagaatgg tgtcattaga gggccacagg 60
ggatggggga gtaaaaaata acataaacga actgaacaga aatgcaggag ggtggcaaga 120
ggggccgaga ttgggtgttc agggcagaga ggtggaagac caggggcagt cagtgttct 180
tagctttcag ccaccagagt ggagaattcg tcaaccccaa ttttgccgtc cccatctttg 240
tctccagcag ccattcagcat cttggtttct ttagcagaca ggtctctggc atctggggag 300
aagcctttta ggatgaatcc cagctcatcc tcctcgatga agccactttg tcctgtcca 360
gcatgtgaaa caccttcttc acatcatccg cactctttt cttcaggccg accatttga 420
agaacttttt gtggtcgaag g 441

```

<210> 183  
 <211> 339  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> (1)...(339)  
 <223> n=A,T,C or G

<400> 183  
 tgtntcatcn taaggggatt gggctctaga tctgtcgacg gcgcattgag gatttgcnat 60  
 cgggttangtg gtccgcgagt catgaatttt tgctctggag cgttattggt tgtgaagttt 120  
 atccaggaga gaactatgat tgtgtcgatg cgtttactgc aggaagantc acggtctcag 180  
 tcacggagggt gtaaggggtgg actgactgan tgagacaagg gatatntngt tnttatannc 240  
 ttgtgatgaa cctgcctacc gtttatgtct ctttgctaag gggctctcng tncgtgnatt 300  
 cncncaagct gcgggggctt ccncggttct gggtctga 339

<210> 184  
 <211> 490  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> (1)...(490)  
 <223> n=A,T,C or G

<400> 184  
 atatagcaag cttgtacgac cgacacatac ggcgcatgtg gctggattgc ttatcttgtc 60  
 gcgcgacgtc tatataancg anactacata gtctcggaaa tccactcant ttcaagttcc 120  
 caaaanacng ganaaaaacc catgccttat ttaactaanc atcagctcgc ttctccttct 180  
 gtaaccgcgc ttntngctcc cagcctatag aagggtaaaa cccacactcg tgcgncagtc 240  
 atcnnataac tgattcgccc gggactgccc gggcggcgct cganaccaat tngcanaatt 300  
 cacacattgc ggcgctcnan aagctctaga aggccaatcg ccatattgat ctatacatta 360  
 tggcgcgtcg tnacacgtcg tgacgggana ncctggngta ccattaatcg ctgcacantc 420  
 ccttcgcagc tggggtnnac aaaagccgcc catcncacca cgttgcncc gatggcaagg 480  
 acncctnat 490

<210> 185  
 <211> 368  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> (1)...(368)  
 <223> n=A,T,C or G

<400> 185  
 ctnnanatag cangcttgta cgaccgacac aatacggcca ntgtgctgga ttcgcttcag 60  
 cgccgccccg gcagtaccgg cgctcatcta tcngatgatg gcgcaccaat gtgggggttt 120  
 aaccttttta tatggctggg gacanaaagc gcggttacnn aaccnataac gagctgatgg 180  
 tcatttaaaa atgcttgggg ttttcccggt cttttgggga attgaaactg agtgggactt 240

```

canaaactgt gctactttcg cttatctaag tactcgggcg caacacctag ccgaatccgc 300
anatatcatc acnctgggcg gcgtcancat gcntctaaag ggccaattcn cctanatgag 360
tcttatac                                     368

```

```

<210> 186
<211> 214
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> (1)...(214)
<223> n=A,T,C or G

```

```

<400> 186
ngggagatcg cagcttgtac gactcgatcat ataacgnnca atgtgctgga tcgcttcanc 60
gccgccggcg gtctaactcg gtccgatttn tgtgtgnttt gtctntntta canggtgcta 120
tccccctctt cctcctcttc tgccatcttc atcctttatc tcctttttgg acaagtgtca 180
nancagacag angcagggtg gtggcaccgt tgaa                               214

```

```

<210> 187
<211> 630
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> (1)...(630)
<223> n=A,T,C or G

```

```

<400> 187
cagctgggac gagtcgatca tatacggcgc atgtgttgna tcgctatcgt gtccggcgag 60
tanttattn attactgtta tttctgctcc tactggatat gatctcttga nggcangtct 120
gtgtcgctcg gtcacacccat gttctcaggc tgggcaaata ccttcctata atagtttatg 180
gataatgaat gacgactang tctanaaana cgctagctaa ataacacact cagggaaga 240
gtcttaaata ttgtgaagggt gtttttanta tacaacnttt gtttacataa taggaaataa 300
tttttagact tttaaacaga cacttgagcc agatttggtta atgttaccat ctatagtgtc 360
ttgaaaatat tcctcttagt ttccaatatg aatgaatcta aaatccatct tttcaattat 420
gccaggccc gtggtcaatg cncctcnac acttcattaa cggattatac cttgggaaac 480
cataatctgg cntaggacga atcgctggc ncangctaan aactgcctg tattgagggg 540
ttatnctga ttgcnagagt gcctctccag gtccccaag ggtcgtagt ttgaanctgg 600
ctctaanttt ntcttgccn acaggtctcc                               630

```

```

<210> 188
<211> 441
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> (1)...(441)
<223> n=A,T,C or G

```

```

<400> 188
cnngcaanac anggtcggat tccgntgagg naanaattcc ctnatagggc tcgcccccta 60

```

```

ttcaccaaac caancngaaa ctcttgcggt caaatctaag ctatnncaca accccactct 120
gnagggtatg cgccccgccc ctgcaatgaa atcaatanca tatttgagaga cagagagata 180
gagagagaga gggtcctggc cttnnctatt ctgctcttac ttggnagatn tcaganatag 240
aaaaacctat cctaggtcn nccaatgatn gcggttncg aatcccgnng tggccantcc 300
ccggatcgga ctaaataaaa gaagatcctc cgtctcctg ttcctccaca ctggagtcct 360
attgtatgca tgggtntttc actggctnat cataccnnag gatctgtcca ccttnaactc 420
ttctctngga antcctncc c 441

```

&lt;210&gt; 189

&lt;211&gt; 637

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; (1)... (637)

&lt;223&gt; n=A,T,C or G

&lt;400&gt; 189

```

agggngtata taccacttg tacnactcga tcatanacgc gcatntctga atcgcttntc 60
ggccgcgatg tactgtggc acttaagcac tgagtactgt ttgcgtcatg ccnggtcana 120
agatgctgct gcaaaggac tccaacnaaa tacactgtct tcaacaggag ttaacacctc 180
acacttggtg ganaanagaa ctactgggtg gtgatgcaca cgactgnatc catcaagtgc 240
gtttgcctgt tgactgctaa ccaaggctct ggagtagcct gccggggcgg cgctcgaaac 300
caaatctgca aatatcatca cactggcggn cgctcagcat catctanaag gccatcgctc 360
atagttagtc tatacatcat ggccgcnttt acactcctac tggaaaacct gcgtaccact 420
taatcgcttc acacatcccc ttctgcngtn gcttatancn aaaagccac gatgcctcca 480
cattgcncnc tgatggcatg anccccctac gcgcatancc gcggtntgtg taccncangt 540
accgtntgc acgctacnnc tcttcttctt cctcttcccc ttcctgttcc tcaccattcg 600
gggccttagg tcnatatctc gnccacccaa atntagg 637

```

&lt;210&gt; 190

&lt;211&gt; 653

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; (1)... (653)

&lt;223&gt; n=A,T,C or G

&lt;400&gt; 190

```

aggggggtata taccacttg tacgactgna tcatatacgc gcatgtctgg aatcgcttnc 60
gtggctgcca tgtattgaca ctacttctaa gaactacaaa agtgatactg angatacatt 120
acacagaang gctnacattc tencagatcc tcatttntca tgatatgtgg acatcangan 180
cacgtggata agtgatatcta aanaatggct ttcaaaatat ttccacttta ttaaggtttg 240
acatganatt cataaaatgt ctttaatacta ttcttnaaaa taacatctaa tcggaaacta 300
tgctnaact gcacnttttn tgtgtanata atcntanttg tacgcccggc ggcgccaaag 360
ccnaatctgc gattcctcac ctggcgccgc tcaacatcat cttaaaggcca atcgcttata 420
ntantctata cctctggcc gcgtttacac gtctaattgg aaaccggcgt accattatc 480
gcttgacgca ctccccctcc cactgggtta tacnaaagcc gcncgatgcc tcccacattc 540
canctgatgc aatgaccctt gtctgcctta ncccgcggtt tgtgtaccca ntnaccant 600
cagcgctgcn cntcttctt ctctcttctt gccttncgt tccctcactc nng 653

```

&lt;210&gt; 191



<211> 663  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1)...(663)  
<223> n=A,T,C or G

<400> 191  
angnggtata tacccactgt negactcgat catatacgcg catgtcggat cggtccanc 60  
gcgcgcgcat gtactatatc tacatcaact gtattatcat ttanatattg atnaaagaca 120  
aaatcatact tccatctgct cactgatgat aattactatg atacatgatc atgtaaactg 180  
atcaatataa caatggaaga tccctctgac tatgcaagcc taattttcca atcncatgca 240  
ctctcatagc tcaaanatnt caengacatc ctgatgaaac tatnatacan tttccacaca 300  
aatcacttcg ctttagatct ctccattatt ctgtcttttc cccctaaca actacaaatc 360  
ctcntgggat gggaagaata tatatcatct actaaaaata atatataatc ccctgcanat 420  
ttgtggnaaa tcnggtgtct caanagccac aggagnacaa gggggnacca actaggactt 480  
ttgtatgctt atctctgtac tcgcgcacac ctaagcgatt ctgcnattct ccctggcggc 540  
gtcacanctc tanaggccat cncnatatga tctatacatc ntggcgctct tacactctga 600  
cggaaccggg gtnccantta ccctggacca tcccttcgcn ctgntatata aagccccga 660  
ncc 663

<210> 192  
<211> 361  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1)...(361)  
<223> n=A,T,C or G

<400> 192  
antttttata tacccactgg tacaactcga ncctatacgg cgcanttncg gaatcanctt 60  
cancggcgcc ggcatgtacc ggtnatcatc atcngatgat ggcgctcnaa tgtgggtttt 120  
acctnttata cggctgagat canatcgctg acataacaaa nncaactgat ggtnaatnta 180  
aatnecggttg ggttctcccn ntctgttggg gaacttgana ctgagtngga cntccatana 240  
cgtgctattn tcggctancn antcctcagc gnacacctat ngnagtgcgc naattcatcc 300  
atgntggcct cgactnttcc aaaangccnt ncgcccacnt gntcgcnana cantctcggc 360  
c 361

<210> 193  
<211> 314  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1)...(314)  
<223> n=A,T,C or G

<400> 193  
aggngnata taccaactgg tncgaactcga tectatacgc gcatttcgga ttcgcttcaa 60  
cggcgccggc atgtacaaaa cctcaatccc aaccgtctca nttngacggg ctcaagtctg 120  
tcacagccac ccacatttcc tttgttttgg tctgcccact caaaagaatt ccaaataaga 180

```

attctgctgc agctccgtac aaggatatgg gcagcacagc acacacagag tngtgctcct 240
cacacttctc tggnaatgtc tcgtgaatat ctcaacagtc angaagtggg gcgttatcaa 300
aaacaatcag ggcc                                     314

```

```

<210> 194
<211> 550
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> (1)...(550)
<223> n=A,T,C or G

```

```

<400> 194
aggngngata taccactggt tncgactcga tcctatacgc gcatgtcggg ncgctatgtg 60
gtcncgcaag tacctcttct gcagtgatgg tctgtntcct ctatgatnag tgategaata 120
atcatcgaat tancgaaag ttattcgagt gatantgtg gcttgtagaa tctatgctcc 180
atgggtgtgg cactgtcaag attaacacag aatggaagan ncngcactgc ataaaagatg 240
ttgtcaaatt ggggtgcgtt atcngatagc tcntcccaag aggtcantgg tgttcaggat 300
tncnacataa gatnttggat caccngacga ccagangata ccngtgcaaa ctgtgaancn 360
ngtaatctgc ctatncttgc cctctcggan gatccctcgg ggacgacgag atcattcttg 420
aaacagcnan tgatagtcca gttnnangatt gatgancgac ganacgcntg atanatgtct 480
gacgtgagat tnggatgtga atcttcccnt gtgtgacctg cncntaccn aanggtgcgn 540
ctccactcnn                                     550

```

```

<210> 195
<211> 452
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> (1)...(452)
<223> n=A,T,C or G

```

```

<400> 195
nngcgggnat gataccaact ggtacgaact cganctctat nacggcgctn tttcnngatc 60
tgctatgtgg tctcggcaat gtacattata acngggcana catataatct acntctgtct 120
ttntctcccc cngagagcgc aancatctcc aaatcgggtt ctgggtcatc caatggtctc 180
cantaatcac acaactcata tatatttatg gaangtgtct gtcacgtgcc ccacgangga 240
agtnncgtcg ctgtntgtct gtcactaggt gngtactctc cagtacttga aancgtgtna 300
nggctgtctg tngtactggc cggcgccctc gaaancgaat ctgtnnatat catcacatng 360
cgncgcccga ncatcactna gggncanttc gcctatactg atcgtntgcg annctgcgn 420
cncttacacg tcgnacggga naccggcctt cc                                     452

```

```

<210> 196
<211> 429
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> (1)...(429)
<223> n=A,T,C or G

```

```

<400> 196
gcggggnnat gataccagct ngtagcactc gatcctataa cggcgcatgt gngtatcggc 60
tacgtgtctc ggcatgttac atataacggg gcaacatata atnatacant ctgtcttttt 120
ctcccccgga aacggcaacc atctccaata tcggctctggg tctccaatgg tctccaacta 180
aatcacacaa gtcaaatata nttanggaaa gtgtctgtct cntccccaga aggagtancg 240
ttagctgttg tctgtcatta ggttggtacc tccagtnaca tgaaaactgg tgaggggtgtc 300
cttgtagaag ctctgcctca ccagatccta tactattagg gggcccacgg ttatctatct 360
taagggtctn aaaacctgga cttcatctgc tccggcggan gaatgtcccg cttactttacg 420
ntgttccac                                     429

```

```

<210> 197
<211> 471
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> (1)...(471)
<223> n=A,T,C or G

```

```

<400> 197
atgatacga gctngtacga gccgtcacta tnacggcnca ttgtgtggat tcngetntga 60
tcggcgcccc ggcatgtcca tcnagagcgc atcatgggan tgnactcccc atatinntgac 120
caangttcgc gcaaggagcc naganccgat actacctgag ctgtcgtctn gttatacacg 180
tttctggcca angancaact ccacatncaa caagttagtg ttgaaatgtt gtttatnagt 240
ccaccaaccg gccgtctctg ccttccccga tgatccgaag ataagcttcc tgtccggaan 300
acgaacggcg tgggtgngg acatantgat atgtgcgggt caggaagtac tcgncgcaac 360
ncgcaagcna atctgcnata tcatcacctg gcggcgctcg agctgccana ngcccnttcg 420
cctatatgag tctatacatt cctggccgctc tnttactc ngacgggaaa c 471

```

```

<210> 198
<211> 643
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> (1)...(643)
<223> n=A,T,C or G

```

```

<400> 198
tngtncgacc gtcactatac gcccatgtgt ggatccgntc caccggcgccg ggcangtacg 60
anactatatt gatcctctga tattgaaagt tggctctanca ataaccttta angcaaatca 120
ctcantgagt tttgaccaga agtcaccaca tcatgaatca cagtctatgg caaatgatac 180
cagtgtctct aagtcctatg ctcaaggtaa gagcatgcta ttccgtttta catttactgg 240
aatttactgt tcattcatna ttaaaatctc tagttttcat cctcaactgt ctaanaccag 300
tgtgcacaga cttaagactc tgttctctc attttctcca acagaaacat tctcagtgtc 360
tactgttcta aaagggaatt tccgaggtgg cacttctcgg aatatcgacc ctonggetct 420
atcaggcggt acttcnngca ctctgcattt gggcttggtc anttgtctta tctgtccagt 480
cacttcattt taagaaaaca attgatcgct ggtcacatgt nattcattgg cagccggtgt 540
gactgctgag tctcgcgcac acnctagcaa tcgnnattct ccatggngcg tcaactctcta 600
naggccatcc cctatatgat ctataatctg gcgtctttac act 643

```

```

<210> 199

```

<211> 292  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> (1)...(292)  
 <223> n=A,T,C or G

<400> 199  
 ncggcnggag ttcgcagttg nacgaccgat cctatacgn cgcatttctga tccgctacnt 60  
 gtccggcgag tctatgctat ttatttntga ttaaatacaat attttctttc tgaatattaa 120  
 tcttatctnt acttttatac tattgacctt gctatatgta ttganctttt tgaactccta 180  
 tcagnttttt tcatgctatc gtatatatttc cacttggtac ctntngctga ntccatagata 240  
 tcgtaaaaca tctctnnatc ntcacacnga gnccagggnt ctgtatngaa tt 292

<210> 200  
 <211> 275  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> (1)...(275)  
 <223> n=A,T,C or G

<400> 200  
 atacgcaagc ttggtaccga gctnggatcc ctattaaccg gcgcgaatat tctggaattc 60  
 tgcttancgt ggtcncggcc gaagtactat gctatnttac ttttttgga tataaatca 120  
 atatatctct tctnaagta tataaatctt atcncgctat cnttcnatac ctntctgaca 180  
 ntaagcttat angtatntga tctntgttga actcctatca agtgntttcn catgctatcg 240  
 tganntcttc cacnttggtg ccttttacgc tgaat 275

<210> 201  
 <211> 284  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> (1)...(284)  
 <223> n=A,T,C or G

<400> 201  
 cgnnnatcca gtgtanaccg tcnttaacgc cattctgatc gttcacgccc gcgtctttat 60  
 atctatctcg actgattcac ctgtcattgt aaanaattcg tgtcagctgt ctaccnctta 120  
 nacatcatct aatcnaacta ncctgataaa tttcttcaat agggatanac ntntagtaca 180  
 tacgnttcca ttgagntacn tccgcggacc cncatcgcaa acnnatgcg gtcagtcnna 240  
 gcatecteta tcttaatccg tccttacent ntgaacgctc cact 284

<210> 202  
 <211> 448  
 <212> DNA  
 <213> Homo sapiens

<220>

<221> misc\_feature  
 <222> (1)...(448)  
 <223> n=A,T,C or G

<400> 202  
 atgatacgca agcttgtacg actcggatca tataacggcc gcaatgtgct ggaattccgc 60  
 ttcgacggac gccgggcatg tacttttata atnctactcc tcagaccttg catctcnacc 120  
 gctnggtcca gtttgtaaaa acnnaactcc gtngtgcagc cctggttctg ancantctct 180  
 atcacnctct atcctcncat ccncaanact anategcgtg aattcatatt tattcatttt 240  
 ccataatgat gggggaanga ctatcnctna tnatgcttan cacnctngct gcanttcgnc 300  
 natctcgcnat ngcntgaaac gattactctg tcgcgaaccc tctangntga attctgcnaa 360  
 atatctntna cncgtggcngg cgctcnangn atgcctctcg anggccaatc cgccnngcat 420  
 gattctaatt anaccntng gtcccntt 448

<210> 203  
 <211> 321  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> (1)...(321)  
 <223> n=A,T,C or G

<400> 203  
 ggggtcnaga tcgcagtngt acgaatcgnt catatacggc gcatgtgntg antcgctacg 60  
 tgtccggcga ngtaccatat aatcgaanta ncatagttct ggangeccnc tcattttcaa 120  
 tttcccaaaa nacgggaaaa ccnaagcctt atttaactaa ctatctgctc gttctctcgt 180  
 tctgtaccgc gctatntgct nccagcctat aanaagggtg aaaccacac tcggtgcgtc 240  
 agtctccnat atantgagtc nccgggtact ggccgggcgg tcgttcnaaa ncaattcncg 300  
 aanttacta ctggcggcgc c 321

<210> 204  
 <211> 369  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> (1)...(369)  
 <223> n=A,T,C or G

<400> 204  
 ntgtngtatg taccagtggt tacgactcga tcctagtacg gcgcagtgtg ctgaatcggt 60  
 acttgctcgc gccaaagtac tataaagcaa actatcacag ttctgaaagt ccatctcant 120  
 ttcagttccc aaaagancgg gaaaacccaa gccttattaa actaacaatc agtcgctctc 180  
 gcttctgtac cgcgcttttg gccccagcc tataaaaggg taaaaccac actcgggtgcg 240  
 ccagtcacgc ataactgaat cgcccgttac tgcccgggcg gcgctcnann ccaaactcgc 300  
 agatacaca cactggcggc gctcancatg ctctagaagg ccaattcncc tatantgatt 360  
 ctattacaa 369

<210> 205  
 <211> 2996  
 <212> DNA  
 <213> Homo sapien

&lt;400&gt; 205

cagccaccgg	agtggatgcc	atctgcaccc	accgcccctga	ccccacaggg	cctgggctgg	60
acagagagca	gctgtatgtg	gagctgagcc	agctgaccca	cagcatcact	gagctggggcc	120
cctacaccct	ggacagggac	agtctctatg	tcaatggttt	cacacagcgg	agctctgtgc	180
ccaccactag	cattcctggg	acccccacag	tggacctggg	aacatctggg	actccagttt	240
ctaaacctgg	tccctcggct	gccagccctc	tccctgggtgct	attcactctc	aacttcacca	300
tcaccaacct	gcgggtatgag	gagaacatgc	agcacccctgg	ctccaggaag	ttcaacacca	360
cggagaggggt	ccttcagggc	ctggctccctg	ttcaagagca	ccagtgttgg	ccctctgtac	420
tctggctgca	gactgacttt	gctcaggcct	gaaaaggatg	ggacagccac	tggagtggat	480
gccatctgca	cccaccaccc	tgaccccaaa	agccctaggg	tggacagaga	gcagctgtat	540
tgggagctga	gccagctgac	ccacaatatc	actgagctgg	gcccctatgc	cctggacaac	600
gacagcctct	ttgtcaatgg	tttcaactcat	cggagctctg	tgtccaccac	cagcactcct	660
gggaccccca	cagtgtatct	gggagcatct	aagactccag	cctcgatatt	tggcccttca	720
gctgccagcc	atctcctgat	actattcacc	ctcaacttca	ccatcactaa	cctgcggtat	780
gaggagaaca	tgtggcctgg	ctccaggaag	ttcaacacta	cagagaggggt	ccttcagggc	840
ctgctaaggc	ccttgttcaa	gaacaccagt	gttggccctc	tgtactctgg	ctgcaggctg	900
accttgtctca	ggccagagaa	agatggggaa	gccaccggag	tggatgccat	ctgcacccac	960
cgccctgacc	ccacaggccc	tgggctggac	agagagcagc	tgtatttgga	gctgagccag	1020
ctgaccacca	gcatactga	gctgggcccc	tacacactgg	acagggacag	tctctatgtc	1080
aatggtttca	cccatcggag	ctctgtacc	accaccagca	ccggggtggg	cagcgaggag	1140
ccattcacac	tgaacttcac	catcaacaac	ctgcgtaca	tggcggacat	gggccaaccc	1200
ggctccctca	agttcaacat	cacagacaac	gtcatgaagc	acctgctcag	tcctttgttc	1260
cagaggagca	gcctgggtgc	acggtacaca	ggctgcaggg	tcacgcact	aaggctctgtg	1320
aagaacgggtg	ctgagacacg	ggtggacctc	ctctgcacct	acctgcagcc	cctcagcggc	1380
ccaggctctgc	ctatcaagca	ggtgttccat	gagctgagcc	agcagaccca	tggcatcacc	1440
cggtctgggccc	cctactctct	ggacaaagac	agcctctacc	ttaacggtta	caatgaacct	1500
ggctccagatg	agcctcctac	aactcccaag	ccagccacca	cattcctgcc	tcctctgtca	1560
gaagccacaa	cagccatggg	gtaccacctg	aagaccctca	cactcaactt	caccatctcc	1620
aatctccagt	attcaccaga	tatgggcaag	ggctcagcta	cattcaactc	caccgagggg	1680
gtccttcagc	acctgctcag	acccttgttc	cagaagagca	gcattgggccc	cttctacttg	1740
ggttgccaac	tgatctccct	caggcctgag	aaggatgggg	cagccactgg	tgtggacacc	1800
acctgcacct	accaccctga	ccctgtgggc	cccgggctgg	acatacagca	gctttactgg	1860
gagctgagtc	agctgaccca	tgggtgtcacc	caactgggct	tctatgtcct	ggacagggat	1920
agcctcttca	tcaatggcta	tgcaccccag	aatttatcaa	tccggggcga	gtaccagata	1980
aatttccaca	ttgtcaactg	gaacctcagt	aatccagacc	ccacatcctc	agagtacatc	2040
accctgctga	gggacatcca	ggacaaggtc	accacactct	acaaaggcag	tcaactacat	2100
gacacattcc	gcttctgcct	ggtcaccaac	ttgacgatgg	actccgtgtt	ggtcactgtc	2160
aaggcattgt	tctcctccaa	tttggacccc	agcctgggtg	agcaagtctt	tctagataag	2220
accctgaatg	cctcattcca	ttggctgggc	tccacctacc	agttgggtgga	catccatgtg	2280
acagaaatgg	agtcacagt	ttatcaacca	acaagcagct	ccagcaccca	gcacttctac	2340
ctgaatttca	ccatcaccaa	cctaccatat	tcccaggaca	aagcccagcc	aggcaccacc	2400
aattaccaga	ggaacaaaag	gaatattgag	gatgcgctca	accaactctt	ccgaaacagc	2460
agcatcaaga	gttatTTTTT	tgaactgtcaa	gtttcaacat	tcaggctctgt	ccccaacagg	2520
caccacaccg	gggtggactc	cctgtgtaac	ttctcgccac	tggctcggag	agtagacaga	2580
gttgccatct	atgaggaatt	tctgcggatg	acccggaatg	gtacccagct	gcagaacttc	2640
accctggaca	ggagcagtg	ccttggtgat	gggtattttc	ccaacagaaa	tgagccctta	2700
actgggaatt	ctgaccttcc	cttctgggct	gtcactctca	tgggcttggc	aggactcctg	2760
ggactcatca	catgcctgat	ctgcgggtgc	ctgggtgacca	cccgcggcgg	gaagaaggaa	2820
ggagaataca	acgtccagca	acagtgccca	ggctactacc	agtcacacct	agacctggag	2880
gatctgcaat	gactggaaact	tgcgggtgcc	tgggggtgcct	ttcccccagc	cagggtccaa	2940
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&lt;210&gt; 206

&lt;211&gt; 914

&lt;212&gt; PRT

&lt;213&gt; Homo sapien

<400> 206

Met	Ser	Met	Val	Ser	His	Ser	Gly	Ala	Leu	Cys	Pro	Pro	Leu	Ala	Phe
1				5					10				15		
Leu	Gly	Pro	Pro	Gln	Trp	Thr	Trp	Glu	His	Leu	Gly	Leu	Gln	Phe	Leu
			20					25					30		
Asn	Leu	Val	Pro	Arg	Leu	Pro	Ala	Leu	Ser	Trp	Cys	Tyr	Ser	Leu	Ser
		35					40					45			
Thr	Ser	Pro	Ser	Pro	Thr	Cys	Gly	Met	Arg	Arg	Thr	Cys	Ser	Thr	Leu
	50					55					60				
Ala	Pro	Gly	Ser	Ser	Thr	Pro	Arg	Arg	Gly	Ser	Phe	Arg	Ala	Trp	Ser
65					70					75					80
Leu	Phe	Lys	Ser	Thr	Ser	Val	Gly	Pro	Leu	Tyr	Ser	Gly	Cys	Arg	Leu
				85					90					95	
Thr	Leu	Leu	Arg	Pro	Glu	Lys	Asp	Gly	Thr	Ala	Thr	Gly	Val	Asp	Ala
			100					105					110		
Ile	Cys	Thr	His	His	Pro	Asp	Pro	Lys	Ser	Pro	Arg	Leu	Asp	Arg	Glu
		115					120					125			
Gln	Leu	Tyr	Trp	Glu	Leu	Ser	Gln	Leu	Thr	His	Asn	Ile	Thr	Glu	Leu
	130					135					140				
Gly	Pro	Tyr	Ala	Leu	Asp	Asn	Asp	Ser	Leu	Phe	Val	Asn	Gly	Phe	Thr
145					150					155					160
His	Arg	Ser	Ser	Val	Ser	Thr	Thr	Ser	Thr	Pro	Gly	Thr	Pro	Thr	Val
				165					170					175	
Tyr	Leu	Gly	Ala	Ser	Lys	Thr	Pro	Ala	Ser	Ile	Phe	Gly	Pro	Ser	Ala
			180					185					190		
Ala	Ser	His	Leu	Leu	Ile	Leu	Phe	Thr	Leu	Asn	Phe	Thr	Ile	Thr	Asn
		195					200					205			
Leu	Arg	Tyr	Glu	Glu	Asn	Met	Trp	Pro	Gly	Ser	Arg	Lys	Phe	Asn	Thr
	210					215					220				
Thr	Glu	Arg	Val	Leu	Gln	Gly	Leu	Leu	Arg	Pro	Leu	Phe	Lys	Asn	Thr
225					230					235					240
Ser	Val	Gly	Pro	Leu	Tyr	Ser	Gly	Cys	Arg	Leu	Thr	Leu	Leu	Arg	Pro
				245					250					255	
Glu	Lys	Asp	Gly	Glu	Ala	Thr	Gly	Val	Asp	Ala	Ile	Cys	Thr	His	Arg
		260						265					270		
Pro	Asp	Pro	Thr	Gly	Pro	Gly	Leu	Asp	Arg	Glu	Gln	Leu	Tyr	Leu	Glu
		275					280					285			
Leu	Ser	Gln	Leu	Thr	His	Ser	Ile	Thr	Glu	Leu	Gly	Pro	Tyr	Thr	Leu
	290					295					300				
Asp	Arg	Asp	Ser	Leu	Tyr	Val	Asn	Gly	Phe	Thr	His	Arg	Ser	Ser	Val
305					310					315					320
Pro	Thr	Thr	Ser	Thr	Gly	Val	Val	Ser	Glu	Glu	Pro	Phe	Thr	Leu	Asn
				325					330					335	
Phe	Thr	Ile	Asn	Asn	Leu	Arg	Tyr	Met	Ala	Asp	Met	Gly	Gln	Pro	Gly
			340					345					350		
Ser	Leu	Lys	Phe	Asn	Ile	Thr	Asp	Asn	Val	Met	Lys	His	Leu	Leu	Ser
		355					360					365			
Pro	Leu	Phe	Gln	Arg	Ser	Ser	Leu	Gly	Ala	Arg	Tyr	Thr	Gly	Cys	Arg
	370					375					380				
Val	Ile	Ala	Leu	Arg	Ser	Val	Lys	Asn	Gly	Ala	Glu	Thr	Arg	Val	Asp
385					390					395					400
Leu	Leu	Cys	Thr	Tyr	Leu	Gln	Pro	Leu	Ser	Gly	Pro	Gly	Leu	Pro	Ile
			405					410					415		
Lys	Gln	Val	Phe	His	Glu	Leu	Ser	Gln	Gln	Thr	His	Gly	Ile	Thr	Arg
		420						425					430		
Leu	Gly	Pro	Tyr	Ser	Leu	Asp	Lys	Asp	Ser	Leu	Tyr	Leu	Asn	Gly	Tyr
		435					440					445			
Asn	Glu	Pro	Gly	Pro	Asp	Glu	Pro	Pro	Thr	Thr	Pro	Lys	Pro	Ala	Thr

450		455		460
Thr Phe Leu Pro Pro	Leu Ser Glu Ala Thr	Thr Ala Met Gly Tyr His		
465	470	475	480	
Leu Lys Thr Leu Thr	Leu Asn Phe Thr Ile	Ser Asn Leu Gln Tyr Ser		
	485	490	495	
Pro Asp Met Gly Lys	Gly Ser Ala Thr Phe	Asn Ser Thr Glu Gly Val		
	500	505	510	
Leu Gln His Leu Leu	Arg Pro Leu Phe Gln	Lys Ser Ser Met Gly Pro		
	515	520	525	
Phe Tyr Leu Gly Cys	Gln Leu Ile Ser Leu	Arg Pro Glu Lys Asp Gly		
	530	535	540	
Ala Ala Thr Gly Val	Asp Thr Thr Cys Thr	Tyr His Pro Asp Pro Val		
545	550	555	560	
Gly Pro Gly Leu Asp	Ile Gln Gln Leu Tyr	Trp Glu Leu Ser Gln Leu		
	565	570	575	
Thr His Gly Val Thr	Gln Leu Gly Phe Tyr	Val Leu Asp Arg Asp Ser		
	580	585	590	
Leu Phe Ile Asn Gly	Tyr Ala Pro Gln	Asn Leu Ser Ile Arg Gly Glu		
	595	600	605	
Tyr Gln Ile Asn Phe	His Ile Val Asn Trp	Asn Leu Ser Asn Pro Asp		
	610	615	620	
Pro Thr Ser Ser Glu	Tyr Ile Thr Leu Leu	Arg Asp Ile Gln Asp Lys		
625	630	635	640	
Val Thr Thr Leu Tyr	Lys Gly Ser Gln Leu	His Asp Thr Phe Arg Phe		
	645	650	655	
Cys Leu Val Thr Asn	Leu Thr Met Asp Ser	Val Leu Val Thr Val Lys		
	660	665	670	
Ala Leu Phe Ser Ser	Asn Leu Asp Pro Ser	Leu Val Glu Gln Val Phe		
	675	680	685	
Leu Asp Lys Thr Leu	Asn Ala Ser Phe His	Trp Leu Gly Ser Thr Tyr		
	690	695	700	
Gln Leu Val Asp Ile	His Val Thr Glu Met	Glu Ser Ser Val Tyr Gln		
705	710	715	720	
Pro Thr Ser Ser Ser	Thr Gln His Phe Tyr	Leu Asn Phe Thr Ile		
	725	730	735	
Thr Asn Leu Pro Tyr	Ser Gln Asp Lys Ala	Gln Pro Gly Thr Thr Asn		
	740	745	750	
Tyr Gln Arg Asn Lys	Arg Asn Ile Glu Asp	Ala Leu Asn Gln Leu Phe		
	755	760	765	
Arg Asn Ser Ser Ile	Lys Ser Tyr Phe Ser	Asp Cys Gln Val Ser Thr		
	770	775	780	
Phe Arg Ser Val Pro	Asn Arg His His Thr	Gly Val Asp Ser Leu Cys		
785	790	795	800	
Asn Phe Ser Pro Leu	Ala Arg Arg Val Asp	Arg Val Ala Ile Tyr Glu		
	805	810	815	
Glu Phe Leu Arg Met	Thr Arg Asn Gly Thr	Gln Leu Gln Asn Phe Thr		
	820	825	830	
Leu Asp Arg Ser Ser	Val Leu Val Asp Gly	Tyr Phe Pro Asn Arg Asn		
	835	840	845	
Glu Pro Leu Thr Gly	Asn Ser Asp Leu Pro	Phe Trp Ala Val Ile Leu		
	850	855	860	
Ile Gly Leu Ala Gly	Leu Leu Gly Leu Ile	Thr Cys Leu Ile Cys Gly		
865	870	875	880	
Val Leu Val Thr Thr	Arg Arg Arg Lys Lys	Glu Gly Glu Tyr Asn Val		
	885	890	895	
Gln Gln Gln Cys Pro	Gly Tyr Tyr Gln Ser	His Leu Asp Leu Glu Asp		
	900	905	910	
Leu Gln				



<210> 207  
 <211> 2627  
 <212> DNA  
 <213> Homo sapiens

<400> 207  
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 tagcatcatc attattctgg ctggagcaat tgcactcatc attggctttg gtatttcagg 180  
 gagacactcc atcacagtca ctactgtcgc ctcagctggg aacattgggg aggatggaat 240  
 cctgagctgc acttttgaac ctgacatcaa actttctgat atcgtgatac aatggctgaa 300  
 ggaaggtgtt ttaggcttgg tccatgagtt caaagaaggc aaagatgagc tgtcggagca 360  
 ggatgaaatg ttcagaggcc ggacagcagt gtttgcgtat caagtgatag ttggcaatgc 420  
 ctctttgcgg ctgaaaaacg tgcaactcac agatgctggc acctacaaat gttatatcat 480  
 cacttctaaa ggcaagggga atgctaacct tgagtataaa actggagcct tcagcatgcc 540  
 ggaagtgaat gtggactata atgccagctc agagaccttg cgggtgtgagg ctccccgatg 600  
 gttccccag cccacagtgg tctgggcac ccaagttgac cagggagcca acttctcgga 660  
 agtctccaat accagctttg agctgaactc tgagaatgtg accatgaagg ttgtgtctgt 720  
 gctctacaat gttacgatca acaacacata ctctgtatg attgaaaatg acattgccaa 780  
 agcaacaggg gatatcaaag tgacagaatc ggagatcaaa aggcggagtc acctacagct 840  
 gctaaactca aaggcttctc tgtgtgtctc ttctttcttt gccatcagct gggcacttct 900  
 gcctctcagc ccttacctga tgctaaaata atgtgccttg gccacaaaaa agcatgcaaa 960  
 gtcattgtta caacagggat ctacagaact atttcaccac cagatatgac ctagttttat 1020  
 atttctggga ggaaatgaat tcatatctag aagtctggag tgagcaaaac agagcaagaa 1080  
 acaaaaagaa gccaaaagca gaaggctcca atatgaacaa gataaatcta tcttcaaaga 1140  
 catattagaa gttgggaaaa taattcatgt gaactagaca agtgtgttaa gagtgataag 1200  
 taaaatgcac gtggagacaa gtgcatcccc agatctcagg gacctcccc tgctgtcac 1260  
 ctggggagtg agaggacagg atagtgcatt ttctttgtct ctgaattttt agttatatgt 1320  
 gctgtaattg tgctctgagg aagccccctg aaagtctatc ccaacatata cacatcttat 1380  
 attccacaaa ttaagctgta gtatgtacct taagacgctg ctaattgact gccacttcgc 1440  
 aactcagggg cggctgcatt ttagtaatgg gtcaaatgat tcacttttta tgatgcttcc 1500  
 aaaggtgcct tggcttctct tcccaactga caaatgccaa agttgagaaa aatgatcata 1560  
 atttttagcat aaacagagca gtcggcgaca ccgattttat aaataaactg agcaccttct 1620  
 ttttaaacaa acaaatgcgg gtttatttct cagatgatgt tcatccgtga atggtccagg 1680  
 gaaggacctt tcacctgac tatatggcat tatgtcatca caagctctga ggcttctcct 1740  
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 cagctggggg gatttcgccc cccatctccg ggggaatgtc tgaagacaat tttggttacc 1860  
 tcaatgaggg agtggaggag gatacagtgc tactaccaac tagtggataa aggccaggga 1920  
 tgctgctcaa cctctacca tgtacaggac gtctcccat tacaactacc caatccgaag 1980  
 tgtcaactgt gtcaggacta agaaaccttg gttttgagta gaaaagggcc tggaaagagg 2040  
 ggagccaaca aatctgtctg ctctctcaca ttagtcattg gcaataaagc attctgtctc 2100  
 tttggctgct gcctcagcac agagagccag aactctatcg ggcaccagga taacatctct 2160  
 cagtgaacag agttgacaag gcctatggga aatgctgat gggattatct tcagcttgtt 2220  
 gagcttctaa gtttctttcc ctctattcta ccctgcaagc caagttctgt aagagaaatg 2280  
 cctgagttct agctcagggt ttcttactct gaatttagat ctccagacct ttcctggcca 2340  
 caattcaaat taaggcaaca aacatatacc ttccatgaag cacacacaga cttttgaaag 2400  
 caaggacaat gactgcttga attgaggcct tgaggaaatga agctttgaag gaaaagaata 2460  
 ctttgtttcc agcccccttc ccacactctt catgtgttaa cactgcctt cctggacctt 2520  
 ggagccacgg tgactgtatt acatgtgtt atagaaaact gattttagag ttctgatcgt 2580  
 tcaagagaat gattaaatat acatttccta caccacaaaa aaaaaaa 2627

<210> 208  
 <211> 282  
 <212> PRT  
 <213> Homo sapiens



&lt;211&gt; 309

&lt;212&gt; PRT

&lt;213&gt; Homo sapiens

&lt;400&gt; 209

```

His Ala Ser Ala His Ala Ser Gly Arg Gln Arg Gln Leu His Ser Ala
      5                      10                      15

Ser Thr Gln Ile Arg Trp Glu Pro Ser Pro Ala Met Ala Ser Leu Gly
      20                      25                      30

Gln Ile Leu Phe Trp Ser Ile Ile Ser Ile Ile Ile Ile Leu Ala Gly
      35                      40                      45

Ala Ile Ala Leu Ile Ile Gly Phe Gly Ile Ser Gly Arg His Ser Ile
      50                      55                      60

Thr Val Thr Thr Val Ala Ser Ala Gly Asn Ile Gly Glu Asp Gly Ile
      65                      70                      75                      80

Leu Ser Cys Thr Phe Glu Pro Asp Ile Lys Leu Ser Asp Ile Val Ile
      85                      90                      95

Gln Trp Leu Lys Glu Gly Val Leu Gly Leu Val His Glu Phe Lys Glu
      100                     105                     110

Gly Lys Asp Glu Leu Ser Glu Gln Asp Glu Met Phe Arg Gly Arg Thr
      115                     120                     125

Ala Val Phe Ala Asp Gln Val Ile Val Gly Asn Ala Ser Leu Arg Leu
      130                     135                     140

Lys Asn Val Gln Leu Thr Asp Ala Gly Thr Tyr Lys Cys Tyr Ile Ile
      145                     150                     155                     160

Thr Ser Lys Gly Lys Gly Asn Ala Asn Leu Glu Tyr Lys Thr Gly Ala
      165                     170                     175

Phe Ser Met Pro Glu Val Asn Val Asp Tyr Asn Ala Ser Ser Glu Thr
      180                     185                     190

Leu Arg Cys Glu Ala Pro Arg Trp Phe Pro Gln Pro Thr Val Val Trp
      195                     200                     205

Ala Ser Gln Val Asp Gln Gly Ala Asn Phe Ser Glu Val Ser Asn Thr
      210                     215                     220

Ser Phe Glu Leu Asn Ser Glu Asn Val Thr Met Lys Val Val Ser Val
      225                     230                     235                     240

Leu Tyr Asn Val Thr Ile Asn Asn Thr Tyr Ser Cys Met Ile Glu Asn
      245                     250                     255

Asp Ile Ala Lys Ala Thr Gly Asp Ile Lys Val Thr Glu Ser Glu Ile
      260                     265                     270

Lys Arg Arg Ser His Leu Gln Leu Leu Asn Ser Lys Ala Ser Leu Cys
      275                     280                     285

```

Val Ser Ser Phe Phe Ala Ile Ser Trp Ala Leu Leu Pro Leu Ser Pro  
 290 295 300

Tyr Leu Met Leu Lys  
 305

<210> 210  
 <211> 742  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> (1)...(742)  
 <223> n=A,T,C or G

<400> 210  
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 aggcccgacc gtcacctgag agccagcaac gggcagtgat gtttagcccc gaggaataat 120  
 tacatgcgga atggaaagca ggcgctcagg gtggctcctg ctggaatgag agctggagtg 180  
 caggctccgt ggttctctggg catgcgggtg tggctcagtt ctacacctgc agatggagtg 240  
 ggactgttga cccaggccag cctggggact gcctctcac ctccctgcgc aggttgacct 300  
 tgtcaccttg cctcttgagc ttgcctctct cctgccaga ngctcttga gcaaaatgga 360  
 ggtcgagagg catttggcac tcacgcctca ccacggacac tgggtgcattc ttgggtacct 420  
 cttggcctca atctattgct gggggangga ngactganc ccattgctgg ggccctgaat 480  
 gcagggactg taaccacca tcccctctc agggcacctc tccctctcca gcacncttg 540  
 tttgctatta atgctaccta atttctact gangtggctc agaagctcct ccgccattgc 600  
 ccttgccgcc agcaaatttt tatccctagg gttaagataa cagaaggcan ccttgggcct 660  
 tgcctgccac attctcaggt ntncactgaa gcacagtatc tatttctcca aaaatagggg 720  
 ctgtnaactt gttactacc cc 742

<210> 211  
 <211> 946  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> (1)...(946)  
 <223> n=A,T,C or G

<400> 211  
 ggcacgaggc acatcgctgg atttctcatt gccaaagctct attaattcat tctttttcat 60  
 aacctcttat tcttatttca tggatgcaac attttctttg tctctcaggg aataataatt 120  
 attctacttt ttaaaggtct aatttcttta ttactttatt tctctgggag tgagtttttc 180  
 ctaaagggat aatgagatgg aaaatgaaaa aacaaaagttg agacatggag ataccttctg 240  
 aaactcaagc attcctctac gtggatgtgc cagagggaaa gaacagaaca aaggagggtg 300  
 gacactattt aaataaaaaat atataagaat attacataac aaacaaaaaa gcccaaatcc 360  
 tcaggttgaa aaggaggaga aaatgtcaag caagacaaaa acagatgaag caacaaaaaa 420  
 agtgacatag ctggtcacct atattgaaat ttcagaacat gagtataaa ggactcccag 480  
 aaaaaaaca aacccaaact aaaaaacaga aaaaaaggac tttaccacn aaaacttgan 540  
 gaatcaggaa gactcagtct ctcatlaaga aaantgctat aggggatggg ggcaaggcct 600  
 tcaaagtngc aggggatacc aataacctct ctgaagtgtt ggaacttcat actccaaaat 660  
 ngaatttttg tttgaatagc cccggttagg ggccaatttt aggacttaga aaggaccng 720  
 gnaaatcatt ccncttgcc ccccccgaa agaaattaat agaaggggtt tattcccgcc 780  
 attannaaaa aaggaatcca ggaatnccg nttttttcca gtgttangnt ggggntgtan 840

aaactgaggg cttagcaagg gcggnattaa ccaccnnggg tcccacccca aaantggng 900  
gggtgggccc caaatcggg nttntncct ttaangcgtt aaaccc 946

<210> 212  
<211> 610  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1)...(610)  
<223> n=A,T,C or G

<400> 212  
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gtggtangag ggcaaccagt aacgggagct tctcctgcca ggcaggaaga cgagtagaag 120  
ggagcggcat gctggaggct ggagcctgag cccctggggc tgccttgct gtgtttggtg 180  
gtgacgtggg aactgcagc tcggccagag tggtaaaaaa tgcctggtg tacgcttttc 240  
tggctttgcc cgtctatctg ctccaagcca ggctgganga ngagganaag gaatcacctg 300  
tggtacgctg gagcctgcat gtggcgtgac tctgcaactc gcctcgtgtg actgatggca 360  
gccacggaga ctgcagctcg acagggagtg aggtcttctca ntggcttgaa agctcagctg 420  
actcccacga aatttgccgg aaactcaagg ctgtcagtga cnttcgtggc gccaaagactt 480  
aancangcgc gttgcatgca tccggccagt gtctgtgcca cgtgccctga cnccaccttg 540  
anataancac ccggaacgcg cncgcgcgag gccgcgcgca cacgnccggg cancaacttg 600  
gtggtcttc 610

<210> 213  
<211> 438  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1)...(438)  
<223> n=A,T,C or G

<400> 213  
ccganagcgg tttaaacggg ccctctagac tcgagcggcc gccctttttt tttttttttg 60  
aaataaattt ctagattatt tattacataa gcagaccact gaaacattta ttcaaaaagta 120  
ttccattgag agtcaaaaac atattgatat gattattatt ggtctgttaa agaaaacaaa 180  
ataaaaagaa caaactggga attatcaata aacaaatcaa aacttagatg taattataac 240  
ctaaagggct cacagggcaa atgtgaagca agcttctgtc tcagagcctg catatggaag 300  
acatgtagta cttagctttg gcattcttct ttcctcctct tggttgagtt taagtattaa 360  
taaaaggtgg actgagaaaa ccttttttta caatcttatg gggattttt agtggaacg 420  
ttttagaagt aggaatat 438

<210> 214  
<211> 906  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1)...(906)  
<223> n=A,T,C or G

```

<400> 214
gccctctaga tcgngcggcc gccctttttt tttttttttt gaaataaatt tctagattat 60
ttattacata agcagaccac tgaaacattt attcaaaagt attccattga gagtcaaaaa 120
catattgata tgattattat tgggtctgtta aagaaaacaa aataaaaaga acaaactggg 180
aattatcaat aaacaaatca aaacttagat gtaattataa cctaaagggc tcacagggca 240
aatgtgaagc aagcttctgt ctcagagcct gcatatggaa gacatgtagt acttagcttt 300
gncatctttc tttcctcctc ttgnttgagt ttagtattaa taaaagttgg actgagaaaa 360
ccttttttta caatcttatg gggtattttt agtggaacg tttagaagta gaataacat 420
attaaaactg cncagaacaa atgnggtgca tctcaaatgg nggtccattt tcaaaatatg 480
aacacatatg ggcagcantt ttttttttaa aaagtcagaa ggggcctnct catgccctt 540
tccacttctt cactcattgg nccttcaacc caagcttaac tactntcctg acctccaaca 600
tcataaacta gtttccnagc tttgaaactt ttttccaatg agtcntaccg gaatagatgn 660
tcacagaanc ctcttaaaaa ttttggaccc tgcccgggnt ntaaaaaggg tgcaataaac 720
ccaccaacat cttggctggg ggggcagggg ccaaaagaan ttcccaaac cgtttttgat 780
naaaaaaggg gacttttgaa aaaaaaatta aaatttttgc cagnaaagca tgggnccccc 840
cccttgaana aacccctgc atnaaaccaa cntnttgga nttttttngg tanggtttt 900
ctggct                                     906

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<210> 215
<211> 312
<212> DNA
<213> Homo sapiens

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<220>
<221> misc_feature
<222> (1)...(312)
<223> n=A,T,C or G

```

```

<400> 215
ggcacgagga aaccaggttg gctgggtttt ggggtgtaaac ttaaaaatga caatcagcat 60
gagctggccg tgggctgtgg ggggtgtagg ggcatcttgg taagggaacc ctgcgtcagt 120
ccctctctgt tctgggtggg aggacaagga gggccaatag gggccaatag ggaggctgct 180
gctaggangg tttcctaaaa gaacaggtgt agggctaggg ctgggtctta gttcaggttg 240
ctctgggcag tgatttatat ccacacacct ttctgcaaag tgctctaagg aganggcagg 300
gataggagtg tc                                     312

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<210> 216
<211> 341
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> (1)...(341)
<223> n=A,T,C or G

```

```

<400> 216
taagcctntc gaanataatg aatgagtcn ggagaggctn atgangaaat nccaaacacc 60
tgactaatng gtgccacatg attncaatgg nctanacatg ggtagatct cntcngnga 120
atgagcaata acacnttaa antcntcaat tgacctagac acttcacact tgaaanatca 180
tcacttttna ngaccacgaa tgatgcttaa gaatcacatt ttgtgnngaa ntggantctg 240
gctacttaca cgaacagatt cttattcctg ttcattgagc agtagaccg gaanaagact 300
taagagcttc tganctttct cttagctcca nngcttgaan g                                     341

```

```

<210> 217

```

<211> 273  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1)...(273)  
<223> n=A,T,C or G

<400> 217  
nnccttcncc ccttnacnga catgaacaaa acagcngtct ngaaatttta ttaacattnn 60  
aagggttaacn ctccctnctt ntgttttccg ntaaannta nacctgcgcn ggggcggccg 120  
atncagccct atagtgagaa gcctaattnc agcacactgg cggccgttac tanngnatec 180  
cgactcggta ncaanttttg gngtaaagat ggacatanct ctatccnnga gnactcgtca 240  
nccnttctct atnttacatg cnctaacgna gac 273

<210> 218  
<211> 687  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1)...(687)  
<223> n=A,T,C or G

<400> 218  
ttttcagtgc tgttttggtc tcaattttga tgtcaaaatc tctgggttct tctaancnng 60  
ttatgttctt ccancaaatc ctccagttt ttgtaatttt tttctatatc agaagcgcct 120  
gancccaatg cccaattnat acaccggtct tctccggaac gcttggtcna aagggtntag 180  
tcnatnnggc tcttgaagc atctnaaatg ctccaggtta ctcccangnc cctggannac 240  
ttcanttgtc tanacgaatc ctgggtttcg agcggtcctt gatatcgcaa ggaaatacgg 300  
taaaaattat ccaagctctc tcccactna gganttcgga tctcatcagc cgggtaaagg 360  
aaaactctc angaagtttg ggcttcccct cgggtctacc ggctaattgt aggaattact 420  
tctggctctc tccgataca tctctcttc aaagtnaaga aggttaaaag aatnttaacn 480  
tctcccagtg gctaattggtc aaacaccatc ctcatnagtc agactggggt ttcgaaagga 540  
ggatataacc tcttgcnag tttnaattaa aagggattaa ccanatggac tanccctcnc 600  
cccgggattt nctctctcac aggagaaggg gtctcncnc ttggctcatc cgaagcatag 660  
gcaaaccnccn ggaattttc agaaacc 687

<210> 219  
<211> 247  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1)...(247)  
<223> n=A,T,C or G

<400> 219  
gggcccttcn cctttnaatc gagagatcca aggttcaagg catgaaatac cagnctataa 60  
aatgtctcaa gacntaaata atacggatng ngatagagag gttgaataat aaatgaanaa 120  
anatgaaagn nattatgngg gaatacnaaa aaancngact aanggcggca ctgctgggca 180  
tggnaaatc ggattaattc ctcataggac agccnaaccc cttaaaatct cantttccgt 240  
naccoga 247

<210> 220  
<211> 937  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1)...(937)  
<223> n=A,T,C or G

<400> 220  
cgggctcgag tgcggccgca agcttttttt actatagacc aatattaaag tcagttaagt 60  
tccaaataca ganttggaag actaaagtaa aatatttaat gggagaatat ctgcatctga 120  
atatgtcaac tgtttgctat ttttcagcta tttaatcctt ctacctgtat ctcagaaaca 180  
aattttaaaa ttaatagatt tgacagcaaa atcattcagc actttactta ctccatcagc 240  
aaggatttta tgtagtcatt tccatccatg tggccaaact gaaaatccct aaccaccacc 300  
aaccaaaaat aaataaataa aaggagaggg ggtgggggga gagagagaga gaaagctcat 360  
taaataagtaa aaaagtaaata aaaacaatga agttaaattc aggcctcagt aggccagaa 420  
actgtaaacat tttcacatgt aaatcatata caataaacac tgctaaaagt gtaaattcta 480  
ctggcttctg agatacaaat acacgagtag aggaaattct aagacatttc tacttggttt 540  
atgcatattt aaaattcagg gaaatatcag ctattctacc tgaaatatgt ttaagaaaaa 600  
ttcctatttt ctctaaaaaa aggaataatc agaagacgct acatactatg taagaaaaact 660  
atacaatgac ccatcattag aagattcaga ataggaaaga aataataatt cactaataaa 720  
atatatttat attgactgtc tttttttatg atagcaacaa tgattcagca taaagtaaaa 780  
atatatgtat ttccgatgcc attttttatt cagttattct tttgagtttc tgttagaata 840  
attatctgcc tatctctgac ttctgancag tcatttatgt ccaattataa gtacatgtgc 900  
atattttatt accttaaacy cctctcaaat cctttca 937

<210> 221  
<211> 353  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1)...(353)  
<223> n=A,T,C or G

<400> 221  
ggctatnnna tnnntntaan atcntgncnn ccttgacgct gttantaaan aaaaacaaac 60  
gaatatcctt tttttgctcc cccctgtncg gatactaata tcacactaat acttacagta 120  
taactnttcc tttcaactac caatattaag ttccaagcca cctgggctta agtatcccaa 180  
caacttaggt aatttggtgc taaccacat actatatgct aattataaca ctctaagccc 240  
caaggaattt ttgttcagat ttcttatant ttccacttat aaatatnatt ccncctctat 300  
gggtatatnn nncctctagn cccatatnnc ccacnggat ttgttgaggg ggc 353

<210> 222  
<211> 813  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1)...(813)



<223> n=A,T,C or G

<400> 222

```

ggcacgagggc tttactaagg ccagactcac tatccccgct tctgttctgt ggtacactgt 60
tcactcctca gtccatccta acctgacttc ctggccactg cagctcttcc gataaggggc 120
agcagtgggt tagttattgc taaataataa gcgcacatgc actccctctt tccctgaaaca 180
ttgtccctcc ttggtttctg ttccttccta ggtctcctat cactcctcct tagtcttctg 240
tgcggacttc tgttccttct gccctttaa agttgggtatt ttccaggatt ctgtcctagg 300
cccacttact tctcattctg cacgttcttg ttggatgatt ctatcacatc cctaacttct 360
gctgcccagt atgcacttaa aattcccaaa tctgtatata tggatctggc ctgtgtctct 420
agcctagaag tgtgctttat ccagaagca cctcaaaccac tgcactttgg aaattaagct 480
tactgagtct cgagtctcaa gtcccaaact gacttctttt tctctatttt ggtagtgac 540
aacactatth attcagtcac gcaaaccaga gccctgagaa ccatcttaca ttctctttct 600
ccctttactc agttcttctg tctgttcttt ctctccncc tctcctgcct gtgggcctag 660
ngnccattaa ctggttgga ctgctttact ttcnattttt ttggctganc taaccnaag 720
ancctnttgt aggggccttt ctntcaggcn tnacttctnn caagancccc cgaaaccaga 780
tcnnggggan tgctatggnn tggaaatatt ttg 813

```

<210> 223

<211> 882

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> (1)...(882)

<223> n=A,T,C or G

<400> 223

```

tcacactact gagaagcagg gaaacccact gaaagggcac gtttcttaac ctgagaatgg 60
ggctactagc ctctaaagca ggaattgcgt tttgtttagt atttccatgg tctgctgcaa 120
ggcgtggcct ttaccaatg gataaatgcg tacaaggctc ttgtgagcag tcaagtttct 180
cgaggtttac agttgaaggg aagtgggatt gtttctctgc gcatttaaat gaaggtaggt 240
gggtgatcac ctttccttaa atgtgtgaag gtagagata aagagatagg catcttaatt 300
gccactgatg gccttcaggt gaggacaggc atgagccaac tgaagctttg acaattgtgc 360
tgaacccaaa acttcaaaaa caagaaaaaa catagactgg ctgaaatgat ctaagtcaac 420
agagcatggc cagcgcttca tacaaggcag gaccacaggg gaacactgac agcccaggag 480
gactgagac agaggcagt ggaagaagt acagacccca gggactcccc accaacagca 540
gctgctgttg attaggaacc ccagtagac tgtcaggcac ctggtagtgg agaggctacc 600
aaggcccgga ctggagagga gccaaaggaa gaaacagtgc agtgcttaga cccctctggg 660
tctgcccgtg tccatacccc tagggagatt ccattccaga agtggacata ttcccacaga 720
gtgcctgggg ctactcacc acagctgccc ctncatgaag gcattctcac tgcagcctta 780
ncagggaaca gggtcatttg cattaggcan cttgctgtcc tagaaggcnt cggngtccc 840
tacactgccc atgttcccaa ngnggttcaa nctcnaaaan tn 882

```

<210> 224

<211> 660

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> (1)...(660)

<223> n=A,T,C or G

<400> 224

```

gattaaactc aatcattcac ccgggctcga gtgcggccgc aagctttttt tttttttttt 60
ttttttttt ttttggncct ctgggcttgt gcccggaagg ggantgctgg gccacntggg 120
tgtecggtgt tgattttctg ggacctgccc ccccgntcc cgcgccgnt gccgcgtctc 180
actccccgcc gcggtgcnag gggccccgtg tgccgcgcac ccttccaccc gtgttttgct 240
gtttttttga cnttgggctg ccaggggtg cancgccgt ggggccctgg tttgctttca 300
cctcttcac tgctcactgg ccgnantgn gtctnttca aacaaacgtn tgaaggncaa 360
nccctgggct cctgtgaacc cggcgtctt tgccgcaaan tctgaggctc cttcggtatt 420
ctggatccgg cctntggctg gangcgtgct ctgcaggcac tgctccatt gctggcancc 480
ttttctcccc gtggccgcc gcccgcccat naaaggcgtt gcaaacgcc gccctcgcca 540
gcgcaaagtc aaacnccggt ggcccgcgga ccccccggcg gncgggaaca cccancagg 600
cgggcaccac aanaagcgc gncctccggc gtctaaaact nccatgtggc nccccccgn 660

```

&lt;210&gt; 225

&lt;211&gt; 438

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; (1) ... (438)

&lt;223&gt; n=A,T,C or G

&lt;400&gt; 225

```

aaaaaaaaag gaaaagtacc cagtgtcttc agcttctgag cctcctctac agccctgttg 60
gnttttaaac ctgtgccctg tgtctgtgtc cccacttaat atatatagta cacagctgga 120
gagatggctc agccaggaga gggaccata ggtctgtgaa ttccagagga naggcaggna 180
tttatagggtg gntctgtcag gtgaaatcng aggagccaaa gctattgtat gtgcatatgt 240
cagccgggct ctgtgggagg tgggtgaaga cctatggnat gggacangtg tncacgctgg 300
gatctctggc cggttccgaa aagtgaggat caggtagtgg gtggctgatt gcacaagttt 360
anaaccagg attagggaca cacaggtcag cacctgtctc tcagcatcct gactgggtgt 420
gatgggcata ctcaaggc

```

438

&lt;210&gt; 226

&lt;211&gt; 480

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; (1) ... (480)

&lt;223&gt; n=A,T,C or G

&lt;400&gt; 226

```

aaaattaaaa ccaaaggat cttagaggtc ctttacttca gtggttctca atgtcagagg 60
atgttatgat acctaataa aatctccagg ggaactgttt tgaactcaac agactctctc 120
ctgttctgag agactctggc aaagttggga gagctgccag gtactgtcca catgaccctg 180
actgcccatg attcaattac cttgaatggc ttatccagtc caataccttc atttcttaca 240
tgaggaaact gaagcacgta tcacatagtg atacaatgaa aacttgccct taatcgattt 300
tcagtgtcgc cagtacaatg tcttgagcat atcaatttct tccaaccctt gacaacataa 360
ggtacgacca tcaaatTTTT tatttctgct aatttattag accaaaaaaa aagggnatct 420
cnccatttgt tttacaggga tgattttatt ncagaggatt tcatcntggg gctgattent 480

```

&lt;210&gt; 227

&lt;211&gt; 423

&lt;212&gt; DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> (1)...(423)

<223> n=A,T,C or G

<400> 227

```
cattgtgttg ggctctgctt agcacatcac atcggagcac agaggtgacc tgttctgccca 60
cagggatggt caccttagtc acctgattga ttcctcttca ctttggtcac gtgattcctc 120
caggaggatg ttcaccttgg tcgcctgatt cctccaggag gatgttcacc ttggtcgcct 180
gaccacacag gcatctatca ggctttctca ctgcagccac tatgtcccca taatggatga 240
gtgtcttgtg gagagatagt ccaaagtaca ctgatacctt ttgcctcata cggcctcacc 300
ccccacaat cnaccactaa tgactgcctc atagcagttt ttccatttcc acagttcctt 360
ctatatgtat taattgtcat tctactataa agaanacttt ttctttttaa aaaaaaaaaa 420
aag 423
```

<210> 228

<211> 249

<212> DNA

<213> Homo sapiens

<400> 228

```
cattgtgttg ggctgtagta aaatatgtgt ctggtaagat atgtgaagaa ataaaataag 60
atcaattaaa tctggcccat tgaatgacac attaattgta tattaatatg taatgttaaa 120
gatattagga gatggtggga cattatggca aactaaattt gggaggagggt tgaattgtat 180
aatttatgaa atcctaaagt ctagtacatt aacactctct actgtcaact tttcaaagca 240
gtgagaaac 249
```

<210> 229

<211> 436

<212> DNA

<213> Homo sapiens

<400> 229

```
cattgtgttg ggatgttatt tgaccatcac aatatgattt ataatatgga ggcattgaagt 60
catttctcat tggggcagga gtgtggcaag ggggaagaag agctttacca attaactcaa 120
gattatttgg tgacatttct cttacctttt aggtgaggag aaagagacag aggatggaga 180
attggtgctt ttagtatgct gatacattaa gctgcctgga agcagatgct aaatcctatt 240
gaaaataatt ttatttgcgt tttgcttagg gcattgttta gcaaaatact acacaaaaag 300
tcttgacctg tgtgtttgaa atggcagatg ttcacagtga ggactgagcc ttggggcaac 360
atcaatcttc acaattctgc acctatttgc tcaataactg gcttggttgg aaaaaaagg 420
aaaaaaaaa aaaaag 436
```

<210> 230

<211> 760

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> (1)...(760)

<223> n=A,T,C or G

&lt;400&gt; 230

```

cattgtgttg ggnngtggaa ggaaaanttt gaggcaatga agctaaacat aaaagaggaa 60
aagcanatgt tacctcaatg accacaatct acaaagtcca aatanaaaac ctgggagtat 120
gataggatga aactataacc tccagcaaag agcttaacag caattaaaat aaagacaaat 180
ttctgggatg gatnagacaa agtagcatat attacaaagg aaaatanact agtatcatnt 240
acgtttgatt aagtaactgc tttcaaataa ttgaatcata aacaatgatt tctgcggttt 300
taagctcatt attttggttc cctgggtttct cctaggatgc agtatagaat ctccatgcct 360
gatgtttatg taccaacaga agctgctgct tctttctttc attatttctt ttttaagtga 420
aagttaatac cttttatatg ttacagagaa gaggcagaaa aagccacact cccactatgc 480
tattaaatgc cctgaggatc aactgaggga tgattatacn catggctgaa tacagtntat 540
tcatttggtt ctttggttg tanataacaa aagggtggtat tctgtaacat cttgtgncaa 600
ttanccaaat gttaaggcga aaatggaatc tttcaaacaa gtgttntaaa caggttttga 660
ttttccaaaa tttantatta gaacntttc aattctggaa gtncccaat ttccangttg 720
tgttttctct tccaattctt ctttcctttg naaatccccc 760

```

&lt;210&gt; 231

&lt;211&gt; 692

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; (1)...(692)

&lt;223&gt; n=A,T,C or G

&lt;400&gt; 231

```

cattgtgttg gggggtgctn tggggagaa acgcttatgt tganatnggg ctccccgaga 60
aagcctcatt gacacnttcg aataaggacc cntngggaaa ttcangtgag ttgtggacat 120
nontagataa natcaaaggc cttgangaag tccgcctggc accttcnctg ctgagaggag 180
gttgatacca aatgctaagg ggtccagntg cantgtanta tctgagatc agagtgatgg 240
gcaggtgttg gcatgcgggc cctcaanang aagtgccag gatgactcag acttatgcct 300
atatccattc antcctgttc attattttta ncnttccctc naaggacccc caatttnaac 360
catttggttat tcanggctat acttataaaa gtcatttggt ttnagtctgg gtgatattaa 420
aaccatttgg acgccangca tgggtggtcn nggcctataa tcctntccac cttgggggag 480
cgaagctgg ttnaatccct naaggtcngg aatttgaaaa ccctcctggg ncaacattgg 540
gngaaaccct gtctctactn caaaaaacan aaaattttct ggggcctnng ttngcaggtn 600
gcctgaaaat ttccancnt tactccggga aggccgaatg ccntaaaaaa nnnaccttta 660
acccccccga angggcggaa agtttccatt tn 692

```

&lt;210&gt; 232

&lt;211&gt; 518

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; (1)...(518)

&lt;223&gt; n=A,T,C or G

&lt;400&gt; 232

```

actcaaatgn ccncttgaag gtcaccaga ctcanaangt gtcaagcttt ggggtggggtg 60
gtaatnaata nctcggntc ctgattagtn ctctagctc gatcncctgg tgagatnngt 120
tcgagcacc ttcttttgat cccgtcaaac nccnggnaaa agcngcctgc gtagtcnct 180
nagccgaatc tgnnttcccg acaccctccg ctccgtcggc tgccctggtn aagcngcntc 240
ctnaaanaaa aaagngaagt ctccccnctg tcncccnant cctngggaaa acngcctgaa 300
ccaatatgnt cccccaaggn cnccccaggg cacntaaccg gttaggaggg cccccnctg 360

```

```
gcgttttggc cnaagccn gcccngnaa taaccnct anaaccacn aaaaatgcaa 420
agtcccaaag ggtaaagaat ctccnacc cccggtccc tcgcaanctt cccctnngna 480
cttgtgtccc gggaaaaccc ttancccgan cctttcca 518
```

```
<210> 233
<211> 698
<212> DNA
<213> Homo sapiens
```

```
<220>
<221> misc_feature
<222> (1)...(698)
<223> n=A,T,C or G
```

```
<400> 233
gcacgagttt ctgtctgtct gtctctctct ctctctctct ctctctctgt ctctctctca 60
cagttagaat ttggctgttt tctttattca atacccaat atatgttcat tagggttata 120
ctgtatacac tacacataac agttttgttt tttgttttgg atattatttg ataataagaa 180
ttttaccaca tcattaaaaa aagtttcccc aagctataat ttttgataat tgcactcttc 240
cactattcaa atgtttattt aactctttct ctctggagt aggtttacat tccatttttag 300
ctatgatact gctttaagag aaattgtttt aagataaatt tccatagaca ggtcaaagga 360
ggtagaatata tgtaagcttt tcgatgcctg ttactgaatc tcattctgga aaacataact 420
gtcaatgccc tctttttctc atggtaaaaa aatacataac aaaatttacc atcttaatcg 480
tttttaaatg ttacagtacg atagtgttna ctgtatgtac cttgtgcaac agattctctg 540
aaaacttttt catttttcaa aatgaaaact ctgtactcat tgaacaggca gcttcccaac 600
ttcccatctc ctccanncc ctacccctgg ttaanagtct nacaaaaccc gggaatttta 660
tgaaatttga aacactttta naataccnct tattaggg 698
```

```
<210> 234
<211> 773
<212> DNA
<213> Homo sapiens
```

```
<220>
<221> misc_feature
<222> (1)...(773)
<223> n=A,T,C or G
```

```
<400> 234
ggcacgagcg cagcttttct aaagctgtaa tttgttttgt atcaaaagtc ctgcagtata 60
ttagtctcat tgcattttta agagtttcca agtgatcagt gatggttgtc tgtttttttag 120
tattacgggc ttatgtaatg ttcgaaaact agtcagtttg gtgctgtcgt acggggcgga 180
aagatcaggg caggcaaagt actctggccg ccaaagtaaa tgcttaaggc cgccaacgga 240
ttatgtcctg gggttcgatg agggccgtaa ttaggttgag ctggtgtang ctaacctcgc 300
agccatgtcg gagagagatg agagacataa nattttaaag taggggcgta ttttacgaag 360
ttctgancca tttcctttgt tatcggtccc ggcaaaagca actgagataa atgtgttaaa 420
agactcgatg attttttcga cttcagcaac gtactcagcc ttgggttctc gtagtttttc 480
aaaggcagct atttgctgag attcatgaaa agtttgactt ganctgcttg tcaattttctg 540
cagcncgggc ttcaactgtt attgaatttg tttgattaag cncaatacgt tgcnggtcac 600
caagggtttc catgttttga ctncacctgg tcgaaccaat ttgaattatg tntttttgcc 660
tgnctgttc cccnccctt aaatccatct cttttttnga aacctttgng nggttgaatt 720
cngccgccc gttcccaacn tttggttcna ccttggaana aaanatgggt agt 773
```

```
<210> 235
<211> 849
```

<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1)...(849)  
<223> n=A,T,C or G

<400> 235  
attgggtacg ggccccctc gagcagcctc cactgcaatg ccgctgaatc aagagacttt 60  
tcaatacgtt ttatcagtga aaatgatgtg atctgaagag tcctatcttg agcactttgc 120  
atgacatcca acgttaatgt ccacaacgtt cttagctgcc caacccttt atcggcaagc 180  
tccaaagggtg tgtgcaaacg ttctacggcg tcatgaaaag ctgaaaaatg ctgtgtcaac 240  
actgcaccgc tgcgcatctt caaaagcagc gcccttatag tctccgcatt cgaagacgat 300  
aaccgcgcta gaatagcctc ataatcactt ttgtagaaat caatcagagc tgtgctagga 360  
acctttccat ccaaaacata cgactgtgctg accacgtctg caaaagcaga cgtcacatta 420  
tgcataatgcc ctcttaccgt cagccgatca tctcactca tagcgacgcg agaaagctct 480  
tggtccagct cgtgcacggt atccaattca gtaatcctac gcaacgccgt ctgaatcgtg 540  
ttcataagtt cagtttttaa gctcaaaact tegtctctta ntttaccctt tgtgactttc 600  
aaactgggcg antcttcacc attttattaa tegtcttttt gangganggc ccagcgttag 660  
atctgcatcg ccagcgggaat cgttactccc tccattcctt cctccgggta acgcanntag 720  
tttctccgaa gccttaaaat tagccgggga aagggaantt atttgcccca acaanggnat 780  
cgcggnctg gtggttaaaa ggaactgaaa taaaattaaa nccncttgg gggaaangcc 840  
cgcatactg 849

<210> 236  
<211> 310  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1)...(310)  
<223> n=A,T,C or G

<400> 236  
ggggtgggtt gcttccgaaa nccggggccc ggccaacttg ttggcttggg aatattctgg 60  
caagaaaatt tccagggcgg cgccaatttn atcaagcccg ggcggcctta aaccgaaaac 120  
tctggcaggg tcaaccctt tcatggcggn ttgaaagctt gaagcgcctt aagttactcc 180  
caagcttggt gcgnttgccg ttggggcgcg gggaaaagtt gaaaacacgg gcgntttgtt 240  
gcccgccccg cgggcggttt nttacgccat cctgggaaaa ctttcagggt tggctgctta 300  
cnaaacggg 310

<210> 237  
<211> 315  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1)...(315)  
<223> n=A,T,C or G

<400> 237  
gcacgagtnt ttgttattta natnttgctt tgtttaangg aagaacacaa naatgccttg 60  
ctaaagggat tctgtttggt tgcangctgc nagcggggaa aaaatcnaa tgtatnttgc 120

```

acaacangat tttttagaan tcagaactat gacatgaagt canncagggc actctacgac 180
tgaatttgcg gtgctgcctt cacangctcc ttntctcgctc tntnctggca ncngtgactc 240
ntacacgtcc tgganantan cctccctana aggaacgact ccgacacccc cccnntaccc 300
ctnaangttc atcng 315

```

```

<210> 238
<211> 510
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> (1)...(510)
<223> n=A,T,C or G

```

```

<400> 238
ngcacgagtn tttgttattt atatattgct ttgtttaaag gaagaacaca aaaatgccct 60
gctaaagggg ttctgttttg ttgcaggctg cnngcgggga aaaaatcaaa gtgtattttg 120
cagaaaatga ttttttanaa gtcagaacta tgacatgaag tcaagcaggg cactctagga 180
ctgaatttgc tgtgctgcct tcatatgctc cttgctcgct cttttctggc agctgtgact 240
cncacaggtc atggaganta tcattcccta aaaggaacaa cnccgatatt catctttatc 300
cattaagtnc atctgtccca ttctatgtng tggatgctaa cttttgatca ttgatngtga 360
tnccatggac atntancatc anctttcana ncctnggatc tttgacnagt cttattantn 420
agantccaac tantacgatg ccganttana aatgctggnt ntccaattcc tactcaaata 480
nccnacatga acttccantc cccttgcnna 510

```

```

<210> 239
<211> 209
<212> DNA
<213> Homo sapiens

```

```

<400> 239
ggtgcttttc ccttctactc gtcttctcgc ctggcaggag aagctccgc tactggttgc 60
ccttctacca ctgtcgacac caccaactgc agtgagccag tgtccgaggc tccagccaga 120
aacaggtagc agccatgccg gataccaaac gccacactt aagagcctga aatgacctga 180
cgccacctcc gcatgcttta cctactgag 209

```

```

<210> 240
<211> 610
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> (1)...(610)
<223> n=A,T,C or G

```

```

<400> 240
ggcacgaggt ttctggctgg agcctcggac actggctcac tgcagttggt ggtgtcgaca 60
gtggtangag ggcaaccagt aacgggagct tctcctgcca ggcaggaaga cgagtagaag 120
ggagcggcat gctggaggct ggagcctgag cccctggggc tcgccttgct gtgtttgttg 180
gtgacgtggg aactgcagc tcggccagag tggtaaaaaa tgccttggtg tacgcttttc 240
tggttttgcc cgtctatctg ctccaagcca ggctgganga ngagganaag gaatcacctg 300
tggtacgctg gagcctgcat gtggcgtgac tctgcaactc gcctcgtgtg actgatggca 360
gccacggaga ctgcagctcg acaggagtg aggttctca ntggcttgaa agctcagctg 420

```

```

actcccacga aatttgccgg aaactcaagg ctgtcagtga cnttcgtggc gccaaagactt 480
aancangcgc gttgcatgca tccggccagt gtctgtgcca cgtgccctga cnccaccttg 540
anataancac ccggaacgcg cnnccgcgag gccgcgcgca cacgnccggg cancaacttg 600
gctggcttcc                                     610

```

```

<210> 241
<211> 474
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> (1)...(474)
<223> n=A,T,C or G

```

```

<400> 241
ggcacgaggt ttctggctgg agcctcggac actggctcac tgcagttggt ggtgtcgaca 60
gtggtangag ggcaaccaat aacgggagct tctcctgcc aaggcagaaga cgantagaan 120
ggancggcat gctggangct ggancctgan cccctggggc tcccttgctg tgtttggtgg 180
tgacgtggga cactgcagct cggccagant ggtaaaaatg tcctgggtgta cgcttttctg 240
gctttgcccg tctatctgct ccaagccacg ctggaagang agganaagga ntcacctgtg 300
gtacgccgga gcctgcatgt gggngtgact ctgcaactcg cctcgtgtga ctgatggcac 360
ccacggacac tgccactcta cagnaatga ggcttctccn tggactngaa agctcanctt 420
nactccncc aagtttgncc gaactcaagg ctntcactna acttcgtggc gccca 474

```

```

<210> 242
<211> 415
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> (1)...(415)
<223> n=A,T,C or G

```

```

<400> 242
ngcgggggnt tccaccagct cgtgtgcaca agtngcgcca cacaacatg cgcaggcact 60
gcatgtcatc natgtgcttc gccgtggttc tggaaacagc agtagaagat ggcgttcggg 120
tcgcgaccaa attcgacgtc ntggatgctc ttgcgcaaga angtcacgta cgggatcggc 180
ccgatggatc cgctnaagcg ccgaaaggcc ctgacttgca aaccgcggct cacagaaccg 240
gcaccaccgg cgccctccgc cnacaaaagt cgagcggcct ccgacacaca ctccctcaca 300
tccccgtcnc gcaactcggc ngtttctagc tccgccacgg ttgtcagcgg caccgcgggc 360
gccnagctgc cggcggcatc cgttgcacac agcacacacg gatccgctct cgtgc 415

```

```

<210> 243
<211> 841
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> (1)...(841)
<223> n=A,T,C or G

```

```

<400> 243

```



```

aacgaggtgt cgatgagcgc gaacaatcgc cctccttcat ctctacctga tggatgaactt 60
cgctcctaca gccgagccaa tgaagacgaa tggctgctgc cgaggatggg agtctcacta 120
gagcacgcgc cgctggacaa ctcatcgact tgtacgcttc cggtagctta gccattcag 180
ctccactgac gacagagacg gagctggcca ctgccatctc gacgcagcgc gacaaggagc 240
agcttcgggc gccgtatgca tcaactcgaag agaaccagga gcagccgga gcaggangcg 300
ctgcacggta caggcacttt cggcgcttca gcggatccat cgggcccgat ccgtagctca 360
ccttcttgcg caagaacatc caggacgtcg aattcggtcg cgaaccgaat gccatcttct 420
actcgctctt ccaggaccgc gcgaagcaca ttgatgacat gcagtgcctt gcgcatgttt 480
gtgcggcgct accttgggtc acacgaacga nggcaaccaa cccgcccag gtgcgcctct 540
atgcattcct gttctgttcc ggtgtgcatg gccggatgtg gaccgtganc ttggtgaatc 600
ggctggtgca tgaagactta ccgctctcnt caagggcgaa cgcncctcan ttcgganaag 660
gaacaaaacc ccccnnaag aacggcantt gcancntttt ccccgctgc cggctcttct 720
ccattcgggn attctctntc tcnaaaant ccgnaaatc ttctttcggg ttctccctcg 780
tttttatttg ccttcccgc cacttgggtt gttttacatc ctacaanct ttttttctc 840
c 841

```

```

<210> 244
<211> 761
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> (1)...(761)
<223> n=A,T,C or G

```

```

<400> 244
aacgaggtgt cgatgagcgc gaacaatcgc cctccttcat ctctacctga tggatgaactt 60
cgctcctaca gccgagccaa tgaagacgaa gtggctgctg ccgaggatgg gactctcact 120
agagcacgcg gcgctggaca actcatcgac ttgtacgctt ccgtagctt agccattca 180
gctccactga cgacagagac ggagctggcc actgccatct cgacgcagcg ggacaaggag 240
cancttcggg cgcgtagtgc atcactcgaa gagaaccagg agcagccgga agcaggaggc 300
gctgcacggg acaggcactt tcggcgcttc agcggatcca tcgggcccga cccgtacgtc 360
accttcttgc gcaagaaaca tccaggacgt cgaattcggt cgcgaccga atgccatctt 420
ctactcgctc ttccaggacc cggcgaagca catttgatga actgcagtgc ctgcgcatgt 480
ttgttgccgc gctacctggg tgcacncgan cgaaggcaac aaccgcgcc angttgccgc 540
tctatgcatt cctgtctgtg ccggtgttgc atggccggat gtgganctg ancttgtgaa 600
tcgctgggt gcataagga cttaccgctc tcgtcaaggg cgaacgcgcc atcaattccg 660
gaaaaggaa naaaacccc cccaangac gnaatttgc ancttttccc ncncctgccg 720
gctcttctcc antnccgggt tctctttctc anaaaattcc c 761

```

```

<210> 245
<211> 710
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> (1)...(710)
<223> n=A,T,C or G

```

```

<400> 245
aacgaggtgt cgatgagcgc gaacaatcgc cctccttcat ctctacctga tggatgaactt 60
cgctcctaca gccgagccaa tgaagacgaa gtggctgctg ccgaggatgg gactctcact 120
agagcacgcg gcgctggaca actcatcgac ttgtacgctt ccgtagctt agccattca 180
gctccactga cgacagagac ggagctggcc actgccatct cgacgcagcg ggacaaggag 240

```

```

cagcttcggg cgccgtatgc atcaactcgaa gagaaccagg agcagccgga agcaggaggc 300
gctgcacggg acaggcactt tcggcgcttc agcggatcca tcgggcccgaat cccgtacgtc 360
accttcttgc gcaagaacat ccaggacgtc aaattcgggc gcgaccgaat gccatcttct 420
actcgctctt ccaggaaccg gcgaagcaca ttgataacat catgcctgcc catgtttgtt 480
gcgggccctcc tgggtgcnca cgaancgaag ggcaacaaac ccgcgccagg tngccgctct 540
tatgcattcc ttgtctgttc cggtnntgca tggcccggan nttggaaccg tnantctggt 600
nnaatcggct ggtgcattga aggaacttac cgctctcgtc aagggccgaa cgcnccttcc 660
agttcggana aaggancgaa aacccccccn naaggaacgg cnttgcnnng 710

```

&lt;210&gt; 246

&lt;211&gt; 704

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; (1)...(704)

&lt;223&gt; n=A,T,C or G

&lt;400&gt; 246

```

aacgagggtg cgatgagcgc gaacaatcgc cctccttcat ctctacctga tgggtgaactt 60
cgctcctaca gccgagccaa tgaanacgaa ntggctgctg ccgaggatgg gagtctcact 120
aaagcacgcg gcgctggaca actcatcgac ttgtacgctt ccggtagctt agccattca 180
gctccactga cgacaganac ggagctggcc actgccatct cgacgcagcg ggacaaggga 240
gcagcttcgg gcgcctatg catcactcga agagaacagg agcagccgga agcaggaggc 300
gctgcccggg acaggcactt tcggcgcttc ancggatcca tcgggcccgaat cccgtacgtc 360
accttcttgc gcaanaacat ccaggacgtc gaattcgggc gcgacccgaa ttgccatctt 420
ctactcgctc ttccaggagc cggcgaagca cattgatnaa attgcattgc ctgcgcatgt 480
ttgtgcgggg ctctctggtg ccccgancga agggcnacaa ccccgcgcca ggggtgcncct 540
ctatgcattc ctntctgttc cgggtgttgcn tgggcgggat ttgaaccgtg aancttggtg 600
aatccgnttg gtgcattaag aacntaaccg ttctctgtca ggggcnnacc ggnccttnc 660
aatttcggaa aaangaacca aaanccccc cnccaagga aacn 704

```

&lt;210&gt; 247

&lt;211&gt; 618

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; (1)...(618)

&lt;223&gt; n=A,T,C or G

&lt;400&gt; 247

```

ggccgccagt gtgatggata tcgaattcaa cgagggtgtcg atgagcgcga acaatcgccc 60
tccttcactc ctacctgatg gtgaacttcg ctctacagc cgagccaatg aagacgaagt 120
ggctgctgcc gaggatggga gtctcactag agcacgcggc gctggacaac tcactgactt 180
gtacgcttcc ggtagcttag cccttcagc tccactgacg acagagacgg agctggccac 240
tgccatctcg acgcagcggg acaaggagca gcttcggggc ccgtatgcat cactcgaaga 300
gaaccaggaa gcagccggaa gcaggaggcg ctgcacggta caggcacttt cggcgcttca 360
gcggatccat cgggcccgat ccgtacgtca ccttcttgcg caagaacatc caggacgtcg 420
aattcggtcg cgaccggaat gccatcttct actcgctctt ccaggacccg gcgaaagcac 480
attgatgaca tgcagtgcct gcgcatgttt gtngcggcgc tacctggtgc acacgagcga 540
nggcaacaaa cccgcgcccc gggtgcgctc tatgcattcc tgttctgtcc ggggtgtgcat 600
ggccccgatg tggaaccc 618

```

<210> 248  
 <211> 622  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> (1)...(622)  
 <223> n=A,T,C or G

```
<400> 248
gcacgagagc ggatccgtgt gtgctgtgtg caacggatgc cgccggcagc ttggcgcccc 60
cggtgccgct gacaaccgtg gcgagagctag aaactgccga agtgcgcgac ggggatgtga 120
gggagtgtgt gtcggaggcc gctcgacttt tgttggcgga gggcgccggt ggtgccggtt 180
ctgtgagccg cggtttgcaa gtcagggcct ttcggcgctt cagcggatcc atcgggccga 240
tcccgtaagt gaccttcttg cgcaagagca tccacnacgt cgaatttggg cgcaaccga 300
acgccatctt ctactcgctc ttccagaacc cggcgaagca cattgacaac atgcnntgcc 360
tgcgcatgtt tgtgcggcgc tncctgntgc acacgaccga gggtagcaac ccgcgccagg 420
ntgcncctct acgcattcct gtctgcccgg tgtgcgtggc cnggatgtgg accntgagcn 480
ggngantccg ctggtgcntg aagacnttgc cgctctcgct aaggccnacc gccntcgcg 540
gcggaaaaag gancaaaanc ccccgcccaa gaaccggcnc tgcaccgttn tcgcgccctt 600
gctgggtctt tctccttac gg                                     622
```

<210> 249  
 <211> 517  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> (1)...(517)  
 <223> n=A,T,C or G

```
<400> 249
cattcgagct cggtagccgg gatccgattg gtaaaggagg tgccgaacag ccagctggtg 60
ttttcgggtg ggcgggggca gcccacatcg ctgtggtcgt tggcgtagtg gatgcgatgt 120
gccgggacaa acgcgttttc caccacgatg tcatgactgc ctgtgccgcy caggcccagc 180
acatcccagt tgtcctcaat gcggtagtcg gccttgggca ccagaaaagt cacatgctcc 240
aggccaggcg tgccatcacg cttgggcagc agaccgccta gaaacagcca gtcgcaatgc 300
ttggagccgg tggaaaagct ccagcgaccg ttgaacctga atccgccttc cacgggctcg 360
gccttgccag taggcatata ggtcgaggcg atgcgcacgc cgttatcctt gcccacaca 420
tctgtctggg cctggtcggg gaaaaancgc cagctgccaa ggggtgaacg ccgaccaccc 480
cgtaaatacca ggcggtggac atgcagccct ttaccaa                                     517
```

<210> 250  
 <211> 215  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> (1)...(215)  
 <223> n=A,T,C or G

```

<400> 250
nntncattgg gccgacgtcg catgctcccg gccgccatgg ccgcgggatt accgcttgtg 60
accgcttgtg accgcttgtg accgcttgtg accgcttgtg accgcttgtg accgcttgtg 120
accgcttgtg accgcttgtg accgcttgtg accgcttgtg accgcttgtg accgcttgtg 180
accgcttgtg acnggggggtg tctggggggac tatga 215

```

```

<210> 251
<211> 231
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> (1)...(231)
<223> n=A,T,C or G

```

```

<400> 251
ngcgccacc tngtgattga tggtcgttta ctatcaagta tgtacatctt gctctagaca 60
actccnattc agtgggaagaa attgggaaag tatcccgat aagtaatagg nattaggtct 120
ncettantgc ttggtgggat attccncaac tgntccngat cggatcagnc tctgtctngn 180
gaatgtgctc gatcgtnatt ctactnctga gcttctatcc nnacgtggcc t 231

```

```

<210> 252
<211> 389
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> (1)...(389)
<223> n=A,T,C or G

```

```

<400> 252
atgtatcanc nctgttggtg ttncatcttt tgcagtcngt tctaagggcn gataantatc 60
agagatgcta atgcatnttc tgccaggcca ncattgggtg cctatgcgta ctcttcttat 120
cttcctgaag agtcatctct ggnggatgtg tccccccctc tccacagtgt ttgcaagcgt 180
taccacgcgn tgtcggngcc gggaaggctn ncacatecgg gnagacttcc ccncgtntga 240
atcgntctn gaatctccgg cgtcttccct naacctcttg actnggacaa ngncccgtn 300
tcccctntgt gaactngtan ccgccccctc tccccccctc agcctaancg ggaangaaga 360
cngggctnat ctngggcncc acaagaant 389

```

```

<210> 253
<211> 289
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> (1)...(289)
<223> n=A,T,C or G

```

```

<400> 253
nggggccnna tgagcgcgcg taatacnatc actatngggc gaattgggta cgggcccccc 60
tcnagcggcc gccttttntt nttttttnt tntttttnt caaaacaccc tcncncntgg 120
atgganacgt nacctttctc taaccanac ttcacaatnc nantctcagg cagccgctc 180

```

```

aaanccgatg tcangttggn atntcaantn caatcttatt ttngngaatta anctganatt 240
gtggatggtn naccaatcan atacttggn tccgttgaac cctgtgga 289

```

```

<210> 254
<211> 410
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> (1)...(410)
<223> n=A,T,C or G

```

```

<400> 254
attgtgttgg gaacttgtag acagctatat caattgcagt gctatttctc tgagggtattg 60
aatctcantt attataatTT tgaaatccaa ttggcttggga cttcattatt ttccaactaa 120
aaagatgatt gaaggattta tttgaaatgt gtaaagagta atatagattt tatgcttatg 180
tttccttgaa aaaagtaggt aaaattcttc tggaagtgtt actcctaaaa tacaaatgaa 240
catgtcaaga attacataaa ttctttaaac tctccttaan aannaatggc tctatgtann 300
gagngaccct tacagactat taagaattaa cttgcatggc anagactcat ttanattcat 360
gaaatggntc tcactttctt ggtaagatct ggcttggacg tttttggtaa 410

```

```

<210> 255
<211> 668
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> (1)...(668)
<223> n=A,T,C or G

```

```

<400> 255
tttttttttt ttttctgtg ccaggcacta taccactgtg ctaggtgcct tctttgcatt 60
acttcatttc ctcataagct ttctgaggan acagaaagct tgagggtcac gtagctagca 120
tctacataaa ttagttgcta aaaacataca atacgtcttc cggcaggctg tcattagtaa 180
ctgatactac tagttgataa tctcataaac ctacganaan ctaccattta agctgaaaca 240
actgtcaata tcactaanta aaacttaaat ccataaatca actatattct aaaatctgac 300
ttcagttcaa ttaaaaaatc actagttgtt acctacctcc ttctgaaagc cagtacaagt 360
taaatagaaca actcccaggt ttaacaaaca agtggcatct aaaaaaaaga tttaaaaaat 420
aatccactta catatattta aaatggcatt aataaaacaa aatttatcca ataacnaant 480
ggcaaaggaa ggtgtccaat tattacatgt tataaatctt taaattaaac ttttcttngg 540
tttttctntc ctanaataaa tacaancctt tccccgccna accagaaaaa agcaaaaaac 600
aaaacccaaa aactcccagc ncngcttaaa aaacncaaaa aaaataaaan ctctattaaa 660
tgcccnaa 668

```

```

<210> 256
<211> 487
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> (1)...(487)
<223> n=A,T,C or G

```

```

<400> 256
cgnaaccgtn cntttttnat gtgcgcccgc cncagnacca gngccgctac aggccaaggc 60
cggaagcacg ggagaggntt nggaaaaaaa agagtgttta caaagagcat attcgagag 120
ttgggatgag tgaaggggac cagaaggngc agcggtaggg acgcgtgaaa ggangcngcg 180
gagaaatgac agcaagaagg gganaagcac acgaaaaggc agtatectcc tcccccttt 240
tcgaggactg ccgcatcttt gttttctgcc cattccagtc accgaanaag atcccaaana 300
aagaagaaaa gaancagagg tgcacttcgc ttcataatttc nctcgctttc ttttctgnct 360
tcacnagttc tgcaggattg cccttgctct cttccgagca catctacgca cgnatgaggc 420
tcggcaggtc aagccnacaa aacnctcgca ctctctttt tctttgcnnng tctgngtggt 480
angngng                                     487

```

```

<210> 257
<211> 502
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> (1)...(502)
<223> n=A,T,C or G

```

```

<400> 257
cctttgaaag nccngctnaa ttcnngnanc cccngatca gcaccaggga gctacaacna 60
aggccggaag caggggattt ngccggaaaa aaaagagtgc ttacaaagag nttatccnca 120
nagatgggat gagtgaaggg gacgagaagg tgcagcggta gggacgcgtg aaaggaggca 180
gcgagaaaat gacagcaaga aggggagaag cacacgaaaa ggcagtatcc tcctcccccc 240
ttttcgagga ctgccgcac tttgttttct gcccattcca gtcaccgaaa aagatcccaa 300
agaaagaaga aaagaaacag aggtgcactt cgcttcatat ttcgctcgct ttcttttctg 360
tttccacaag tctgcaggat tgcccttgct ctcttccgag cacatctacg cagtatgag 420
gctcggaggc caagccaaaa aaacgcttgc actcctcttt ttctttgcgt gtctgtgtgt 480
atgtggaatt ccgcggcncc gc                                     502

```

```

<210> 258
<211> 510
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> (1)...(510)
<223> n=A,T,C or G

```

```

<400> 258
actcgnact cgatncanta caagagnnta tgnattcgaa ngtgcccccg catcagcacc 60
aggagctac aacgaaggcc ggaagcaggg gagagggccg gaaaaaaaag agtgcttaca 120
aagagcatat ccgcagagtt gggatgagtg aaggggacga gaaggtgcag cggtagggac 180
gcgtgaaaagg aggcagcggg gaaatgacag caagaagggg agaagcacac gaaaaggcag 240
tatcctctct ccccttttct gaggactgcc gcactcttgt tttctgcca ttccagtcac 300
cgaaaaagat cccaaagaaa gaanaaaaga aacagagggtg cacttcgctt catatttcgc 360
tcgctttctt ttctgtcttc caagtctgca ggattgccct tgcctcttcc cgagcacatc 420
tacgcacgta tgaagctcgg aggtcnnngc aaaaaaacgc ttgcactcct ctttttcttt 480
gcnagtctgt gtgcatgngg gaaatnctna                                     510

```

```

<210> 259

```

<211> 292  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1) ... (292)  
<223> n=A,T,C or G

<400> 259  
gannngagtc acgaaaaggc agtatcctcc tcccccttt tcgaggactg ccgcattctt 60  
gttttctgcc cattccagtc accgaaaaag atcccaaaga aagaagaaaa gaaacagagg 120  
tgcatttcgc ttcattttc gctcgtttc ttttctgtc tcacaagtct gcaggattgc 180  
ccttgctctc ttccgagcac atctacgcac gtatgaggct cggagggtcaa gccaaaaaaa 240  
cgcttgcact cctctttttc tttgcgtgtc tgtgtgtatg tggaattcct tg 292

<210> 260  
<211> 582  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1) ... (582)  
<223> n=A,T,C or G

<400> 260  
gcacgagggt ggggtgtact gtgtataata actccagatc cttgaccaag tttggagagt 60  
cacttatggc catttgaac caaatgaagg atcaaaggac taattatttt gaatacctct 120  
gagtgttttc cccaagcttg agaagagttt cattcagcta taaaatgctc attgtgcaaa 180  
tgagtggttt ccatgctgta taattaaagc attgccttta ataattttt attaccttta 240  
gcttgctctt ttaatttgag gaaaatccaa acaattttaa gtaaaacgtg ataaagacag 300  
tttttcngga gananaaggg nagatcgcta tgtttattcc acttaatatc tatatcaaat 360  
atgtgtatca aaagcagact ctcaatttaa aaatattctt ctaatggcna gaatcttttn 420  
cctagattga gagtccagagc tcacatagna tnactgctgg taaatagaca cttagactat 480  
agagctnagc tnaagttcca actanccaac tgcatttctg aatatgcttt ttattnaaag 540  
gccagnnctt ttgccttttt nccnccctaa tnccttctat tg 582

<210> 261  
<211> 783  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1) ... (783)  
<223> n=A,T,C or G

<400> 261  
gcacgaggga aaatacagag ggtatttttac catggacagg caaccatttt ttccaggaca 60  
actctttgca gcagagagct attctctttc ttttgcttta cactctcaac ctcaactctc 120  
gagtgtctgc atcctanttt tccatggcca taagataagg aaccatgagt gttactctag 180  
atgaggctgt ttcattgtgg gagctcatcc aggatccaag gtagattcat cagaagggtta 240  
agtataggag tgggaaccca aatctctact tttattttga ggccttctct cctcaatttt 300  
aaattgtaaa atcaaactta aaactgggta tctgatggcc agttaaaga ctgggtatct 360  
gattgccagt taagagatgg tcatttatgc tcaccacat tctcaagacg cagggtagggt 420  
gacangcttg ctgggggaatg ctgancgaat cccccaatgc cttcaggatt ctggggaatg 480

```

tggtctctgnt ttaaactggn tgactttttac aaagagccta cccgtcatgg ggggactggg 540
aagaaaaccc anangcagnt tctggcccan ggttacaccc ccanggn tac cttgaaggnt 600
ttttggacat acctnttncc cccctnttac tgnttcatta gggcntcnnc aaccaantt 660
tccaagttnt ggccctttna aaantttttt nttttccntt tccanggacc cccctggntt 720
cctggnnccc cctttttata nccaaccttg ccnggnattt tttcnnttn aaagggaaat 780
aat

```

783

&lt;210&gt; 262

&lt;211&gt; 741

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; (1)...(741)

&lt;223&gt; n=A,T,C or G

&lt;400&gt; 262

```

tgaacctan tgggcccggc cccctcgagt cgacgggtac gataagcttg atatcgaatt 60
cggcacgagt gtatatcttg ttattatacc ccagattnaa gtgtatattc ttaggcagta 120
gttcttggtta acatccttac tacataaaat ccacttacta ttttaagtatt attctaacag 180
gaggtagaat agctgcctta aaaaatgtag tgatcgaatg gcagtttttc tgctgaatgg 240
aaattactga cacaaaattht ggthtttgga gacattttcc tccttggtgt tgagttttcc 300
cattcacgga tagggcataa agcttggttt atagttgagg ggtgcaaaag gggaaatagga 360
ttgggaaaat acagtgttcc agcaaagggtc tgacaaggta catcttgagg aggattccta 420
ttctgctang tggcactgta ngtcttgaaa tactgtgtac tttccagaca aaggatagag 480
aaaaagacct tcactgggtg ggggagaaga aaaccttgt tcctagaaaa atcacaaaaa 540
aggcatcctt tancctatat tcccagnttt actggngcat ttgcttgatg tgactgacnc 600
ngattatttc ctttnactgg naaaaattcc tgcncctttg gatatnaang ggggnaccng 660
gaaaatnggg ggcnttgggg aaggaaanaa aaaaaattgg agggaccnaa ctttggaana 720
tgggntgctt nangccttaa g

```

741

&lt;210&gt; 263

&lt;211&gt; 437

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; (1)...(437)

&lt;223&gt; n=A,T,C or G

&lt;400&gt; 263

```

ggcacgagag aatgtgttca cagacactat tttatannta tctgatgtgt actgtgtctg 60
gtggatgtga aagccatact tcttaaactc gatttgaaaa gcaaatctga ttatcacagc 120
cataattaaa tttggccagc cttccttcc cctccctcc ttcacttcc tcttccctc 180
cgctcgtgc cgaattcggc acgagcctga cctcactacc aaaaaaaaaa aaattcaaag 240
tgcttgagg ttccaggcat tcttagctct atttacttac tttccacctc aaatggcctt 300
agaattcaaa ttctgnanaa aatggattgc catanataat ccaatgaaaa tgggtcatat 360
tttgccatta atagaatcac agtcnacaag ggactaatag aattagtcac ttangtatcn 420
ttagatttgg gagacnn

```

437

&lt;210&gt; 264

&lt;211&gt; 706

&lt;212&gt; DNA



<213> Homo sapiens

<220>

<221> misc\_feature

<222> (1)...(706)

<223> n=A,T,C or G

<400> 264

```
gcacgagcac cccaaggttt taggacaaaa tgggatgagt gaattcatgg cttgacagac 60
tgaacagaaa aatgaggctc cgtgctccat attcatgtgc atctgcccct catggtgaca 120
tgctaattgg ttggccggtg cacaagacaa ggaagtgcag gtttcctgtt gctcacacag 180
tgcttcctgt ctgctgtggc aggagccggg aggaaggagg cgagccaaga ggggtgctgc 240
ccaccggaaa cgatggcgcg aggcgcgaga gctaaatggg ggcctctcca gggagtgtc 300
tgttcacggc tccatcgctg ttagtaagta tcttgtgatt tcggaattta aatgagggtg 360
tgtttaacct gcataacatc tggcttttaa aatctgactt tattttcctt ttatttctgt 420
gcacggtc aggcacactt agtggtggct taggtgttga agtcaggtta ccaaacagca 480
cgccctctct ttattctcag gctgcgtgtt tcattgattc tgaaggtcag atggctgtgt 540
tcaagttctg ttagtatatt ggtgtcagaa atgaaaagat gatgtaacct ttataactt 600
cttaaaggct catatcatgt caggaaatta acctgtacga gttatggaca aatgcccata 660
ctgatgattt tcanccatga aatgaatna aagggganaa gggcca 706
```

<210> 265

<211> 717

<212> DNA

<213> Homo sapiens

<400> 265

```
ggcagagca gcattacggt ttatacacat gtccacaact cagcattgct ttcaaaatag 60
gaacacttta ttagtaaaga ggaagaaatt gcctaacacag actcagtgtc ttcccataa 120
caatcatctg ccaagccgca ggcctaacca ggaaatccca tttccttttg gcgttggtgc 180
ctccaccaac agatacaacc ctgatgccaa atgttgtatg gtttgtaggt gttgtgagcc 240
aatgagggca tgcctagggc caaaggctgc cctttggaat gagggcaagg tcgtagactc 300
catcaaaaaa caaatgcatc ctctccaaa atcaaatgct caacacatgc agcctttcgt 360
atgccatct cccctttact cattttcatg gctgaaaatc atcaggatgg gcatttgtcc 420
ataactccta caggttaatt tcctgacatg atatgagcct ttaagaagtt ataaagggtt 480
acatcatctt ttctattctg acaccaatat actaacagaa cttgaacaca gccatctgac 540
cttcagaatc aatgaaacac gcagcctgag aataaagaga gggcgtgctg ttggttaacc 600
tgacttcaac acctaaagcca ccactaagtg tgcctgagcc gatgcacaga aataaaagga 660
aaataaagtc agattttaaa aagccagatg ttatgcaggg taaacacaac ctcatta 717
```

<210> 266

<211> 362

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> (1)...(362)

<223> n=A,T,C or G

<400> 266

```
ggcagagggt tagatttaac ttccacagat gactcagcag aggataacta ctaatcagag 60
tacaacatca aaactgtaac cagtataatc actggattat gagcaactca aaatagctcc 120
agtttccaaa gggccataaa ctgcacatag cagtactatg tgcaattaac acataattta 180
ttatgaaaat gtggacatgc caggtaagta aggggattta ggttgacttt ttataatact 240
ttaaatttga aatgccattt ctgtggattg gatgacatct tccagggtgt ntaatnctgg 300
```

gntacctnct gatanatcct gananaaaga ggtanacacca gcgtctatca nacctcaata 360  
ca 362

<210> 267  
<211> 692  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1)...(692)  
<223> n=A,T,C or G

<400> 267  
ggcagcaggt tagatttaac ttccacagat gactcagcag aggataacta ctaatcagag 60  
tacaacatca aaactgtaac cagtataatc actggattat gagcaactca aaatagctcc 120  
agttttccaaa gggccataac tggccctttt aanacttttn gcaattaaca cataatztat 180  
tatgaaaatg tggacatgcc aggtaagtaa ggggatttag gttgactttt tataataactt 240  
taaatttgaa atgccatttc tgtggattgg atgacatctt ccagggtgctt taatttggtt 300  
tacctcctga tagatcctga cagaaagagg naggcaccagc gtctatcaaa cctcaatata 360  
gngtgtgaaa cacangagag cctgcttttg tcnacacggg gaaacacatt gttatcacia 420  
cacacaaaag gcaanctncc aatgggggnan ncttacctgn cctctcatat tgggggcaan 480  
gaaaangggg cccccanagtg gctgagtana tccccaaaaa cncactan tggtcagnnt 540  
gcttcccccac acagccagat gactgaattt agcccaagct gcagtctcaa aaccagcttt 600  
ctgacaatca gtaacaagaa catactggtc tgttgacgtg agctcaagtg ttgggtgttc 660  
agtcaaaaanc catggatgcc aatcatctcc ca 692

<210> 268  
<211> 605  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1)...(605)  
<223> n=A,T,C or G

<400> 268  
cgtgccgaat tcggcagcag ngcacatatc agtactatgt gcaattaaca cataatztat 60  
tatgaaaatg tggacatgcc aggtaagtaa ggggatttan gttgactttt tataataactt 120  
taaatttgaa atgccatttc tgtggattgg atgacatctt ccagggtgctt taatttggtt 180  
tacctcctga tagatcctga cagaaagagg tagcaccagc gtctatcaaa cctcaatata 240  
gttgtaaaac acagagagcc tgcttgccca cacatggaga aacattgtta tcacaagaca 300  
cagaaggcaa acttccaatc tggcatactt nctgtctctc tcatatttgg ggcaatgaga 360  
atggtggacc agatggcttg antagatgcc aaagaacacc canactgggc agcatgcttn 420  
cccagacagc cngaagactg aaatttantc ccagctgcag ncttaaacc tttttttgac 480  
nttcogtaac cagaccatac ttttttttct gatgcttttc ttaacttcat cttttccaat 540  
taaattcatt agtnnaacc taaanggggc cgtttttccg aaaaattttc nttntnttt 600  
ccccc 605

<210> 269  
<211> 535  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1)...(535)  
<223> n=A,T,C or G

<400> 269  
gcacgaggng caacccacagg gtgggggtctc tgggatgaac ctggagacct gagcttgcac 60  
agcttccttg gtaaattgag gaggcattga ccacaagatt gccaaagctcc ttcttatcca 120  
aacttgatat tgtagattc catgatccag ttcatcacgg ttgatggctg aatctcatgc 180  
actanaaaaa ggtaatatata aaganaaaaa tanaangatn ttcaagtga tataaanacc 240  
tttaattctca ntctttctag ttcaaagaga cggaacaatg agagatgctg gttcatanag 300  
ctgntanatt taacttccac agatgactca ncagaggata actactaatc anagtacaac 360  
atcaaaactg taaccagtat aatcactgga ttatgagcaa ctcaaaatag ctccagtttc 420  
caaagggcca taaactgcca tatcaantac tatgtgccat taaccataa tttattatga 480  
aaatgtggac atgccangtn agtaagggga tttaggggtga ctttttatna tactt 535

<210> 270  
<211> 803  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1)...(803)  
<223> n=A,T,C or G

<400> 270  
gcacgagggc aacccacagg tgggggtctct gggatgaacc tggagacctg agcttgcaca 60  
gcttccttgg taaattgagg aggcattggc cacaagattg ccaagctcct ttctatccaa 120  
acttgatatt gttagattcc atgatccagt tcatcacggg tgatggctga atctcatgca 180  
ctagaaaaag gtaatatata aaaaaaaaaat aaaaagatat tcaagtgaat ataaagacct 240  
ttaatctcag tctttctagt tcaaagagac ggaacaatga gagatgctgg ttcatagagc 300  
tgtagatttt aacttccaca gatgactcag cagaggataa ctactaatca gagtacaaca 360  
tcaaaactgt aaccagtata atcactggat tatgagcaac tcaaaatagc tccagtttcc 420  
aaagggccat aaactgcaca tatcagtact atgtgcaatt aacacataat ttattatgaa 480  
aatgtggaca tgccaggtaa gtaaggggat ttaggttgac tttttataat actttaatt 540  
tgaaatgccca tttctgtgga ttggatgaca tcttcacggg gcttttaattt ggtttacctc 600  
ctgatagatc ctgacagaaa gaggtagcac cagcgtctat caaacctcaa tacagttgta 660  
aaacacagag agcctgnttt gcctacncat ggagaacatt gttatcaca gacacagaag 720  
ggaactcca tctggctact tacctggctt tatttttggg gcaatganaa tngggggacc 780  
aatggnatgan tanatgccaa aaa 803

<210> 271  
<211> 836  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1)...(836)  
<223> n=A,T,C or G

<400> 271  
gcacgagggc aacccacagg tgggggtctct gggatgaacc tggagacctg agcttgcaca 60  
gcttccttgg taaattgagg aggcattggc cacaagattg ccaagctcct ttctatccaa 120  
acttgatatt gttagattcc atgatccagt tcatcacggg tgatggctga atctcatgca 180

```

ctagaaaaag gtaatatata agaaaaaaat aaaaagatat tcaagtgagt ataaagacct 240
ttaatctcag tctttctagt tcaaagagac ggaacaatga gagatgctgg ttcatagagc 300
tgtagattt aacttccaca gatgactcag cagaggataa ctactaatca gagtacaaca 360
tcaaaactgt aaccagtata atcactggat tatgagcaac tcaaaatagc tccagtttcc 420
aaagggccat aaactgcaca tatcagtact atgtgcaatt aacacataat ttattatgaa 480
aatgtggaca tgccaggtaa gtaaggggat ttaggttgac tttttataat actttaaatt 540
tgaaatgcc a tttctgtgga ttggatgaca tcttccaggt gctttaattt ggtttacctc 600
ctgatagatc ctgacagaaa gangtagcac cagcgtctat caaacctcaa tacagttgta 660
aaacacagag agcctgcttt gnctacacat ggagaaacat tgtatcacia gacacagnaa 720
ggcaacttcc atctgggata ctacctgtct ctctatttgg ggcattganat ggggacaatg 780
ntgananatg caanacacca atgngagctg ntccnagc cnatatgatt ntccat 836

```

&lt;210&gt; 272

&lt;211&gt; 203

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; (1)...(203)

&lt;223&gt; n=A,T,C or G

&lt;400&gt; 272

```

ggagaattgg gcccgctcang ggtgcattct gcatcacctg anttcnaaat ctnagtcaat 60
cnnctacta atantatcaa catnatttna acctgatctc cactgcttng tnattttcnn 120
ttcactgncc ctntcactng aacntctntt cacacagcca cccccatta tctggntggc 180
acctcncca aatncncct naa 203

```

&lt;210&gt; 273

&lt;211&gt; 594

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; (1)...(594)

&lt;223&gt; n=A,T,C or G

&lt;400&gt; 273

```

attcgggccn ctggatnctg gctcgagcgg ccgccgctgt gatggatc tgcanaatc 60
ggcttctgga gagagctttn tttttgatgg ttgcangtac tctcgatgga gttgggtggg 120
gtgggttatct ctctctggtt gtctttctgt ataaanttct tgcntgact nectanctn 180
ctccccctg gtccttcctc tagngtaaca nctggtaatc cctntcttct ttgctctect 240
tnttctect ganegatttc ctctntttgt ccactctcag gnanaaccct gntggtcagt 300
gttcattgact tcngaagnt cgacccgcna aatagggncn cacggatnat gttgaancng 360
ggaagggagn gtccaanttc tctgttccan aggcctnagc tagaganaat gatgggagan 420
ggtttactga gatcatngnn tcttctcgaa gatatnnttt aggggtgtcc cccataagng 480
aatttctcan cttcaaatct tctaatacat tactgaacan ctgncatttg ttacgccaca 540
nattgnaatt ctccatntct ttttagaaac nattncaagg tcatttattt ccct 594

```

&lt;210&gt; 274

&lt;211&gt; 229

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

<221> misc\_feature  
 <222> (1)...(229)  
 <223> n=A,T,C or G

<400> 274  
 ctactcactg tccggccatt tggncctctg natgcatnct caagcagcnc gccantatga 60  
 tnnatatctg cacanttcag cttctngaga aaactatgtt ttaaacagtt gcntanactt 120  
 anaatanaaa tcgagtaagg tntagatnan tctctaacga tngaattatt ntacanaggg 180  
 gtanncgatn accaggagta nctaganttg ancancancc taggtcnga 229

<210> 275  
 <211> 651  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> (1)...(651)  
 <223> n=A,T,C or G

<400> 275  
 atatctgntg aatacggntt cctgnaaaaa ggtntnattt agatgggtga gtccgactca 60  
 gcgatgcgac ttggtgggtg tggtcantct cttatgggtg agattgttca tgatatcatg 120  
 ccctgagatg cctggactnn cctcacggga gatcctagac ggtgntancc cctgagagtc 180  
 tctctctccc tgctctccta acttctccta atgatccctc cnattgtcta ctgtccnatt 240  
 gaacccttct tgcttatgta tncaatcntt nacgggtgtcc ctgctnantt tttganacga 300  
 ngctcataat ggaacngggga aggatagtnt gaataatntc ctgtataccc acgccnacnt 360  
 ctacnctntg atctgacacg gtatactgat ttgtgctgtt cncttcacca ttccantttc 420  
 taccttcgcg tcatatgctc tgtangctac accctctgtg actgctttct cagttacgtg 480  
 caacaaggtn ttcatatctn gaactcttac accattctag anggatcncc cctcgganaa 540  
 antttggaan aacaagcaag ancanaatnc ctctctnctg ntacacnanc cggcttncgt 600  
 atcctcgtnn aaggaattcc cgcgtttcct gggctttaan tctcctaaac t 651

<210> 276  
 <211> 392  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> (1)...(392)  
 <223> n=A,T,C or G

<400> 276  
 accccccccg aattacgntg gccnatntaa aagtncatca ngcctccang caacntatcn 60  
 tttcattacc acccacactc ctggttnnggg anggangtgg naatccttca ccatnctaata 120  
 gtatgtgggtg ctctcatgcn ggtacgtata atctanncgt cccctnaaat cggatgcttc 180  
 tgtaatcnnc agtcacnaaa ccacanggan caactgaaac angatttggtc taacagccaa 240  
 tgtctgggcc ctcnnaatc cctnnaatat ctctacacc tgtagtanna atnaactacn 300  
 ctacnctatt nnacacacgn tttagggtgt annaccaagc ccntattgag tgaaatcggt 360  
 tntatngtat naaatgccaa aagntgcggt aa 392

<210> 277  
 <211> 212  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> (1)...(212)  
 <223> n=A,T,C or G

<400> 277  
 gggtttgcggg natgaanttt gnaanaatna acttttagnga taaccacccc accaatnctt 60  
 nctnagtatt tgncaacctn aaaactacag ctctctccag atagactntn ccttnctgat 120  
 ttcaactctc cttggactgg tcagcctgaa ggggtggaat gactcaccaa cgctactaat 180  
 nccttnttna ctgtgccttn attttttcgc ct 212

<210> 278  
 <211> 269  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> (1)...(269)  
 <223> n=A,T,C or G

<400> 278  
 nnntccatcc taataccact cactatcggg ctggaancgg ccgcccgggc acgtntcttn 60  
 tgngacagga tctgaatnaa ggggtggttg taacttnact naaaattctg aaatgaccc 120  
 gcatcagaca ggggttctccg tntanaatan agtttccctg ttagttatcn agcctgggca 180  
 ggggangana gattcgagga cntntgaaat gaaggnatta tttaggatgg gtgactcatt 240  
 ccnaccnttc ncgctnacca gnccganga 269

<210> 279  
 <211> 266  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> (1)...(266)  
 <223> n=A,T,C or G

<400> 279  
 gttggtgant cngtttgng tcttctcgtt gntnggtgtt tgggtgtgtg nnttggtgtn 60  
 gggtngtntt tntggagaga gttgtagttc gtgaggggtg cagtgtactt actatggagc 120  
 ctaaggangt gngctaactt anantgatna ctttgctcat actgccctgc cctnaatgcc 180  
 nngcttgctt caccctggtg ccnaaccnna tcgaacacct aacagtctag taggcttctt 240  
 gctntancag actnctcttg aggatc 266

<210> 280  
 <211> 317  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> (1)...(317)  
 <223> n=A,T,C or G

<400> 280  
acactgttnag gtgtntggaa ntgntgtagg catagncttt ntggcacaga gttggagccg 60  
tgaggcatag cntgtactta ctatggagcc taaggangga gctaacttat antnatnact 120  
ttgtcctaac tgccttcttc tnaatgecta ngcttgcttc accctgntgc cttacnnnat 180  
cgaacaccta cgcggtctat aggtctcttg ctctatcagg actnctcttc nagcttcntc 240  
gcctcanttg actcactgtg ctcggtcgtt ctactgngat ccagncgctc atnaacctna 300  
cttnggacgc aggtcat 317

<210> 281  
<211> 174  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1) ... (174)  
<223> n=A,T,C or G

<400> 281  
gnggtcatat tatacatcta aggcattggcc aactccacgc cattatnaat tccatcgtac 60  
tgtccgcagt cactacttat aacctagatt aatagtgcct ggccccggac ngctctgtgca 120  
atctnccgcc ataccaattn cgatccncan accncgatna cactcctcct tact 174

<210> 282  
<211> 169  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1) ... (169)  
<223> n=A,T,C or G

<400> 282  
atcgcagctt gtacgatcgt catataacgc gcatgtgcgg atcgcttcag cgccgcccga 60  
ctgtcagaag gangagatct tttttatcac ttgtttgttt gactatanat aanancgact 120  
acagcattga tgtgtgtcct caaganttgt ctgggtctga naaagctga 169

<210> 283  
<211> 157  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1) ... (157)  
<223> n=A,T,C or G

<400> 283  
ggnntntctaa gatcgcagtt gtacgatcgt catatnacgc gcatgtgcgn atcgcttcac 60  
gtcgcncggc tgtccaggan atgcatntca acataatgtg cactctatat gggtattgat 120  
taatacgagn tangagcana tatcngatac aacacaa 157

<210> 284  
<211> 133  
<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> (1)...(133)

<223> n=A,T,C or G

<400> 284

```
ggngtggtgt nagatacgca ngctgggacg aatcgnntca tagtacggcg catgtgttga 60
tcaattctga aaatccatcc cggcgcgctc ancatgcact anagggcaat cgcctatatg 120
antcgtatta caa 133
```

<210> 285

<211> 194

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> (1)...(194)

<223> n=A,T,C or G

<400> 285

```
ntntgngtga tgatacccaa gctggntacc nactngantc caattaccgg ctcantntgc 60
tngaaacngc ttcgatngnc tctggcatg tacttgaaac aggnatanata tctaatagnn 120
tacngtgtnn ttttcnatca tacagnttnt atattncact nccnccatt cntttctant 180
ctctctctcc ntat 194
```

<210> 286

<211> 134

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> (1)...(134)

<223> n=A,T,C or G

<400> 286

```
gaggggnntat gataccaagc tggtagcanc ccgtcactat nacggcccag tgtgtggatc 60
cgctanctgg tcncgcgatg tctacncaca cngaaactgc ctctcgnaa gatctcctct 120
cctctccnaa gaga 134
```

<210> 287

<211> 119

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> (1)...(119)

<223> n=A,T,C or G

<400> 287

```
tnggggtatat ccagttgtac actggncata tacgcgcatt atgatcgttt cacgcccga 60
gtacggcatc attacganat ggnctcattc gtttaccttt ntcgctggac acaagcgtc 119
```

<210> 288



<211> 170  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1)...(170)  
<223> n=A,T,C or G

<400> 288  
gggntgagat acncaagttg gtacgagtcg gatcatatna cggncgccat tttctggaat 60  
ccgcttacgt ggtcccggcg aagtactttt tcatgccttg caaaatngcg ttactgcact 120  
ancttgctta acctatgagt ggggtctttc atacccttc tntcatggaa 170

<210> 289  
<211> 126  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1)...(126)  
<223> n=A,T,C or G

<400> 289  
ggccaattgg ggcctctana tgcntgctcg aacgggcgcc aatttnatgg atatctccaa 60  
aattcggctt accntggtcg cggncnaagt acttaactca atccatctnt cactcaggat 120  
naatgc 126

<210> 290  
<211> 126  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> (1)...(126)  
<223> n=A,T,C or G

<400> 290  
ggccaattgg ggcctctana tgcntgctcg aacgggcgcc aatttnatgg atatctccaa 60  
aattcggctt accntggtcg cggncnaagt acttaactca atccatctnt cactcaggat 120  
naatgc 126

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